

AME Medical Review 2A006

# KEY LEADERS' OPINION ON MICRORNA AND MYOCARDIAL INFARCTION

HONORARY EDITORS: LARS MAEGDEFESSEL  
MENNO HOEKSTRA  
YVAN DEVAUX

EDITORS: CHENGJIAN YANG  
ALBERTO DOMINGUEZ-RODRIGUEZ  
NIKOLAOS PAPAGEORGIOU

ASSOCIATE EDITORS: ZHIJUN HAN  
JOHANNES MAIR  
ANDREA ROGNONI  
JOHNSON RAJASINGH



[www.amegroups.com](http://www.amegroups.com)

AME Medical Review 2A006

# KEY LEADERS' OPINION ON MICRORNA AND MYOCARDIAL INFARCTION

HONORARY EDITORS: LARS MAEGDEFESSEL

MENNO HOEKSTRA

YVAN DEVAUX

EDITORS: CHENGJIAN YANG

ALBERTO DOMINGUEZ-RODRIGUEZ

NIKOLAOS PAPAGEORGIU

ASSOCIATE EDITORS: ZHIJUN HAN

JOHANNES MAIR

ANDREA ROGNONI

JOHNSON RAJASINGH

# AME Publishing Company

Room C 16F, Kings Wing Plaza 1, NO. 3 on Kwan Street, Shatin, NT, Hong Kong

Information on this title: [www.amegroups.com](http://www.amegroups.com)

For more information, contact [books@amegroups.com](mailto:books@amegroups.com)

Copyright © AME Publishing Company. All rights reserved.

This publication is in copyright. Subject to statutory exception and to the provisions of relevant collective licensing agreements, no reproduction of any part may take place without the written permission of AME Publishing Company.

First published in 2019

Printed in China by AME Publishing Company

Editors: Chengjian Yang, Alberto Dominguez-Rodriguez, Nikolaos Papageorgiou

Cover Image Illustrator: Zhijing Xu, Shanghai, China

## **Key Leaders' Opinion on MicroRNA and Myocardial Infarction**

(Hard Cover)

ISBN 978-988-78920-9-0

AME Publishing Company, Hong Kong

---

AME Publishing Company has no responsibility for the persistence or accuracy of URLs for external or third-party internet websites referred to in this publication, and does not guarantee that any content on such websites is, or will remain, accurate or appropriate.

The advice and opinions expressed in this book are solely those of the authors and do not necessarily represent the views or practices of the publisher. No representation is made by the publisher about the suitability of the information contained in this book, and there is no consent, endorsement or recommendation provided by the publisher, express or implied, with regard to its contents.

# Key Leaders' Opinion on MicroRNA and Myocardial Infarction (FIRST EDITION)

## HONORARY EDITORS

---

### Lars Maegdefessel

Molecular Vascular Medicine, Karolinska Institute, Center for Molecular Medicine L8:03, 17176 Stockholm, Sweden; Experimental Vascular Surgery and Vascular Biology Laboratories, Technical University Munich, Klinikum rechts der Isar, Department of Vascular and Endovascular Surgery, Munich, Germany

### Menno Hoekstra

Division of BioTherapeutics, Leiden Academic Centre for Drug Research, Gorlaeus Laboratories, Einsteinweg 55, 2333CC Leiden, The Netherlands

### Yvan Devaux

Cardiovascular Research Unit, Luxembourg Institute of Health, 84 Val Fleuri, L-1445 Luxembourg, Luxembourg

## EDITORS

---

### Chengjian Yang

Department of Cardiology, Wuxi Second Hospital, Nanjing Medical University, Wuxi, Jiangsu, China

### Alberto Dominguez-Rodriguez

Hospital Universitario de Canarias, Department of Cardiology, Ofrs s/n La Cuesta E-38320, Tenerife, Spain

### Nikolaos Papageorgiou

Barts Heart Centre, St Bartholomew's Hospital, West Smithfield, London, EC1A 7BE, UK

## ASSOCIATE EDITORS

---

### Zhijun Han

Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, Jiangsu, China

### Johannes Mair

Department of Internal Medicine III – Cardiology and Angiology, Innsbruck Medical University, A-6020 Innsbruck, Austria

### Andrea Rognoni

Coronary Care Unit and Catheterization Laboratory, "Maggiore della Carità Hospital", Corso Mazzini 18, 28100 Novara, Italy

### Johnson Rajasingh

Department of Internal Medicine, Cardiovascular Research Institute, Department of Biochemistry and Molecular Biology, University of Kansas Medical Center, Kansas City, KS 66160, USA

## EDITORIAL BOARD

---

### Mingzhu Gao

Department of Laboratory Medicine, The Affiliated Wuxi No.2 People's Hospital of Nanjing Medical University, Wuxi, 214002, China

### Yan Jin

Department of Cardiology, The Affiliated Wuxi No.2 People's Hospital of Nanjing Medical University, Wuxi, 214002, China

### Haohao Liu

Department of Laboratory Medicine, The Affiliated Wuxi No.2 People's Hospital of Nanjing Medical University, Wuxi, 214002, China

### Xiaoxiao Liu

Department of Cardiology, The Affiliated Wuxi No.2 People's Hospital of Nanjing Medical University, Wuxi, 214002, China

### Yuanyuan Sang

Department of Cardiology, The Affiliated Wuxi No.2 People's Hospital of Nanjing Medical University, Wuxi, 214002, China

### Junhong Wang

Department of Geriatrics, First Affiliated Hospital with Nanjing Medical University, Nanjing, 210029, China



**Ke Wang**

Key Laboratory of Nuclear Medicine, Ministry of Health,  
Jiangsu Key Laboratory of Molecular Nuclear Medicine,  
Jiangsu Institute of Nuclear Medicine, Wuxi, Jiangsu,  
214063, China

**Shuya Wang**

Department of Cardiology, The Affiliated Wuxi No.2  
People's Hospital of Nanjing Medical University, Wuxi,  
214002, China

**Yan Wang**

Department of Laboratory Medicine, The Affiliated Wuxi  
No.2 People's Hospital of Nanjing Medical University,  
Wuxi, 214002, China

**Lizhu Zhang**

Department of Cardiology, The Affiliated Wuxi No.2  
People's Hospital of Nanjing Medical University, Wuxi,  
214002, China

**Peiling Zhou**

Department of Cardiology, The Affiliated Wuxi No.2  
People's Hospital of Nanjing Medical University, Wuxi,  
214002, China

**AUTHORS**

---

**Robert Adam**

Department of Cardiology, Basingstoke and North  
Hampshire Hospital, Aldermaston Road, Basingstoke,  
Hampshire, RG24 9NA, UK

**Mariama Akodad**

Cardiology Department, Hôpital Arnaud de Villeneuve,  
CHU de Montpellier, UFR de Médecine, Université  
Montpellier 1, 371, avenue du Doyen Gaston Giraud, 34295  
Montpellier cedex 05, France; PhyMedExp, University of  
Montpellier, INSERM U1046, CNRS UMR 9214, 34295  
Montpellier cedex 5, France

**Brian H. Annex**

Division of Cardiovascular Medicine, Robert M. Berne  
Cardiovascular Research Center, University of Virginia,  
Charlottesville, USA

**Wayne Balkan**

Interdisciplinary Stem Cell Institute, University of Miami  
Miller School of Medicine, Miami, FL, USA

**Kirstine Belling**

Novo Nordisk Foundation Center for Protein Research,  
Faculty of Health and Medical Sciences, University of  
Copenhagen, Copenhagen, Denmark

**Michael A. Bellio**

Interdisciplinary Stem Cell Institute, University of Miami  
Miller School of Medicine, Miami, FL, USA

**Stefan Blankenberg**

Univeristy Heart Center Hamburg, Clinic for General and  
Interventional Cardiology, Hamburg, Germany; German  
Center for Cardiovascular Research, partner site Hamburg/  
Lübeck/Kiel, Hamburg, Germany

**Angelo Sante Bongo**

Coronary Care Unit and Catheterization Laboratory,  
"Maggiore della Carità Hospital", Novara, Italy

**Søren Brunak**

Novo Nordisk Foundation Center for Protein Research,  
Faculty of Health and Medical Sciences, University of  
Copenhagen, Copenhagen, Denmark

**Beata Burzynska**

Institute of Biochemistry and Biophysics, Polish Academy  
of Sciences, Warsaw, Poland

**Thuy Cao**

Department of Internal Medicine, Cardiovascular Research  
Institute, University of Kansas Medical Center, Kansas City,  
KS 66160, USA

**Chiara Cavallino**

Coronary Care Unit and Catheterization Laboratory,  
"Maggiore della Carità Hospital", Novara, Italy; Division of  
Cardiology, Sant'Andrew Hospital, Vercelli, Italy

**Dennis V. Cokkinos**

Heart and Vessel Department, Biomedical Research  
Foundation Academy of Athens, 115 27 Athens, Greece

**Hilary A. Collier**

Department of Molecular, Cell and Developmental Biology, University of California, Los Angeles, CA, USA; Department of Biological Chemistry, David Geffen School of Medicine, Los Angeles, CA, USA

**Paula A. da Costa Martins**

Department of Cardiology, CARIM School for Cardiovascular Diseases, Faculty of Health, Medicine and Life Sciences, Maastricht University, Maastricht, the Netherlands; Department of Physiology and Cardiothoracic Surgery, Faculty of Medicine, University of Porto, Porto, Portugal

**Elena De Falco**

Department of Medico-Surgical Sciences and Biotechnologies, Sapienza University of Rome, Rome, Italy

**Maurice W. J. de Ronde**

Department of Vascular Medicine, Department of Clinical Epidemiology, Biostatistics and Bioinformatics, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands

**Leon J. De Windt**

Department of Cardiology, CARIM School for Cardiovascular Diseases, Faculty of Health, Medicine and Life Sciences, Maastricht University, Maastricht, the Netherlands

**Yvan Devaux**

Cardiovascular Research Unit, Luxembourg Institute of Health, Luxembourg, Luxembourg

**Pieter Adrianus Doevendans**

Department of Cardiology, Experimental Cardiology Laboratory, University Medical Center Utrecht, the Netherlands; Netherlands Heart Institute (ICIN), Utrecht, the Netherlands

**Alberto Dominguez-Rodriguez**

Hospital Universitario de Canarias, Servicio de Cardiología, Tenerife, Spain; Facultad de Ciencias de la Salud, Universidad Europea de Canarias, La Orotava, Santa Cruz de Tenerife, Spain

**Bethany Doran**

Duke Molecular Physiology Institute, Durham, NC, USA

**Ingo Eitel**

Department of Cardiology, Angiology, Intensive Care Medicine, Medical Clinic II, University Heart Center of Lübeck, Lübeck, Germany; German Center for Cardiovascular Research (DZHK), partner site Hamburg/Kiel/Lübeck, Lübeck, Germany

**Hamid El Azzouzi**

Department of Cardiology, Experimental Cardiology Laboratory, University Medical Center Utrecht, the Netherlands; UMC Utrecht Regenerative Medicine Center, University Medical Center Utrecht, the Netherlands

**Hans-Josef Feistritzer**

University Clinic of Internal Medicine III, Cardiology and Angiology, Medical University of Innsbruck, A-6020 Innsbruck, Austria

**Lasse Folkersen**

Center for Biological Sequence analysis, Technical University of Denmark, Lyngby, Denmark

**Giacomo Frati**

Department of Medico-Surgical Sciences and Biotechnologies, Sapienza University of Rome, Rome, Italy; Department of AngioCardioNeurology, IRCCS Neuromed, Pozzilli, Italy

**Emeline Goretti**

Cardiovascular Research Unit, Luxembourg Institute of Health, Luxembourg, Luxembourg

**Sabina P. W. Guenther**

Transplant and Stem Cell Immunobiology Laboratory, Department of Surgery, University of California, San Francisco, CA, USA; Transplant and Stem Cell Immunobiology Laboratory, University Heart Centre Hamburg, University of Hamburg, Cardiovascular Research, Hamburg, Germany; Cardiovascular Research Center (CVRC), University Medical Center Hamburg-Eppendorf, Hamburg, Germany; German Centre for Cardiovascular Research (DZHK) e.V., University Medical Center Hamburg-Eppendorf, Hamburg, Germany; Department of Cardiac Surgery, University Hospital Munich, Ludwig-Maximilian-University, Munich, Germany

**Emer E. Hackett**

School of Biochemistry & Immunology, Trinity Biomedical Sciences Institute, Trinity College, Dublin, Ireland

**Nazanin Hakimzadeh**

Department of Biomedical Engineering & Physics, Department of Cardiology, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands

**Joshua M. Hare**

Interdisciplinary Stem Cell Institute, University of Miami Miller School of Medicine, Miami, FL, USA

**Surovi Hazarika**

Division of Cardiovascular Medicine, Robert M. Berne Cardiovascular Research Center, University of Virginia, Charlottesville, USA

**Menno Hoekstra**

Division of Biopharmaceutics, Leiden Academic Centre for Drug Research, Gorlaeus Laboratories, Einsteinweg 55, 2333CC Leiden, The Netherlands

**Dominic Kelly**

Department of Cardiology, Basingstoke and North Hampshire Hospital, Aldermaston Road, Basingstoke, Hampshire, RG24 9NA, UK

**Marek Kiliszek**

Department of Cardiology and Internal Diseases, Military Institute of Medicine, Warsaw, Poland

**Gert Klug**

Cardiology and Angiology, University Clinic of Internal Medicine III, Medical University of Innsbruck, Anichstraße 35, A-6020 Innsbruck, Austria

**Nicolle Kränkel**

Department of Cardiology, Charité Universitätsmedizin, Campus Benjamin Franklin, Berlin, Germany; German Center for Cardiovascular Research, partner site Berlin, Berlin, Germany

**Yuhuang Li**

Department of Medicine, Karolinska Institutet, Solna, Stockholm 17176, Sweden

**Gregory Y. H. Lip**

University of Birmingham Institute of Cardiovascular Sciences, City Hospital, Birmingham, UK

**Agata Maciejak**

Institute of Biochemistry and Biophysics, Polish Academy of Sciences, Warsaw, Poland

**Urszula Mackiewicz**

Department of Clinical Physiology, Medical Centre of Postgraduate Education, Warsaw, Poland

**Michal Maczewski**

Department of Clinical Physiology, Medical Centre of Postgraduate Education, Warsaw, Poland

**Lars Maegdefessel**

Department of Medicine, Karolinska Institutet, Solna, Stockholm 17176, Sweden

**Johannes Mair**

Department of Internal Medicine III – Cardiology and Angiology, Innsbruck Medical University, A-6020 Innsbruck, Austria

**Francisco Marín**

Department of Cardiology, Hospital Clínico Universitario Virgen de la Arrixaca (IMIB-Arrixaca), Universidad de Murcia, Murcia, Spain;

**Mathias Mericskay**

Université Paris-Saclay, Université Paris-Sud, Signalisation et Physiopathologie Cardiovasculaire, Inserm UMR-S 1180, LabEx LERMIT, DHU TORINO, Faculty of Pharmacy, F-92296 Chatenay-Malabry, France

**Bernhard Metzler**

University Clinic of Internal Medicine III, Cardiology and Angiology, Medical University of Innsbruck, A-6020 Innsbruck, Austria

**Mithun Mitra**

Department of Molecular, Cell and Developmental Biology, University of California, Los Angeles, CA, USA; Department of Biological Chemistry, David Geffen School of Medicine, Los Angeles, CA, USA

**Jochen D. Muehlschlegel**

Department of Anesthesiology, Perioperative, and Pain Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, USA

**Esteban Orenes-Piñero**

Proteomic Unit, Instituto Murciano de Investigación Biosanitaria Virgen de la Arrixaca (IMIB-Arrixaca), Universidad de Murcia, Murcia, Spain

**Lara Ottaviani**

Department of Cardiology, CARIM School for Cardiovascular Diseases, Faculty of Health, Medicine and Life Sciences, Maastricht University, Maastricht, the Netherlands

**Samir Ounzain**

Experimental Cardiology Unit, Department of Medicine, University of Lausanne Medical School, Lausanne, Switzerland

**Nikolaos Papageorgiou**

Barts Heart Centre, St Bartholomew's Hospital, London, UK

**Thierry Pedrazzini**

Experimental Cardiology Unit, Department of Medicine, University of Lausanne Medical School, Lausanne, Switzerland

**Jan J. Piek**

Department of Cardiology, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands

**Yigal M. Pinto**

Department of Experimental Cardiology, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands

**Sara-Joan Pinto-Sietsma**

Department of Vascular Medicine, Department of Clinical Epidemiology, Biostatistics and Bioinformatics, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands

**Sheeja Rajasingh**

Department of Internal Medicine, Cardiovascular Research Institute, University of Kansas Medical Center, Kansas City, KS 66160, USA

**Johnson Rajasingh**

Department of Internal Medicine, Cardiovascular Research Institute, Department of Biochemistry and Molecular Biology, University of Kansas Medical Center, Kansas City, KS 66160, USA

**Francesco Rametta**

Division of Cardiology, Sant'Andrew Hospital, Vercelli, Italy

**Milena Rizzo**

Institute of Clinical Physiology (IFC), National Research Council (CNR), Pisa, Italy; Tuscan Tumor Institute, Florence, Italy

**Andrea Rognoni**

Coronary Care Unit and Catheterization Laboratory, "Maggiore della Carità Hospital", Novara, Italy

**François Roubille**

Cardiology Department, Hôpital Arnaud de Villeneuve, CHU de Montpellier, UFR de Médecine, Université Montpellier 1, 371, avenue du Doyen Gaston Giraud, 34295 Montpellier cedex 05, France; PhyMedExp, University of Montpellier, INSERM U1046, CNRS UMR 9214, 34295 Montpellier cedex 5, France

**Francesco Russo**

Novo Nordisk Foundation Center for Protein Research, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark

**Louis A. Saddy**

Department of Anesthesiology, Perioperative, and Pain Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, USA

**Junichi Sadoshima**

Department of Cell Biology and Molecular Medicine, Rutgers New Jersey Medical School, Newark, NJ, USA



**Sonja Schrepfer**

Transplant and Stem Cell Immunobiology Laboratory, Department of Surgery, University of California, San Francisco, CA, USA; Transplant and Stem Cell Immunobiology Laboratory, University Heart Centre Hamburg, University of Hamburg, Cardiovascular Research, Hamburg, Germany; Cardiovascular Research Center (CVRC), University Medical Center Hamburg-Eppendorf, Hamburg, Germany; German Centre for Cardiovascular Research (DZHK) e.V., University Medical Center Hamburg-Eppendorf, Hamburg, Germany

**Ivonne Hernandez Schulman**

Interdisciplinary Stem Cell Institute, Division of Nephrology and Hypertension, University of Miami Miller School of Medicine, Miami, FL, USA

**Sebastiano Sciarretta**

Department of Medico-Surgical Sciences and Biotechnologies, Sapienza University of Rome, Rome, Italy; Department of AngioCardioNeurology, IRCCS Neuromed, Pozzilli, Italy

**Frederick J. Sheedy**

School of Biochemistry & Immunology, Trinity Biomedical Sciences Institute, Trinity College, Dublin, Ireland

**Joost Petrus Gerardus Sluiter**

Department of Cardiology, Experimental Cardiology Laboratory, University Medical Center Utrecht, the Netherlands; UMC Utrecht Regenerative Medicine Center, University Medical Center Utrecht, the Netherlands; Netherlands Heart Institute (ICIN), Utrecht, the Netherlands

**Thomas Stiermaier**

Department of Cardiology, Angiology, Intensive Care Medicine, Medical Clinic II, University Heart Center of Lübeck, Lübeck, Germany; German Center for Cardiovascular Research (DZHK), partner site Hamburg/Kiel/Lübeck, Lübeck, Germany

**Holger Thiele**

Department of Cardiology, Angiology, Intensive Care Medicine, Medical Clinic II, University Heart Center of Lübeck, Lübeck, Germany; German Center for Cardiovascular Research (DZHK), partner site Hamburg/Kiel/Lübeck, Lübeck, Germany

**Dimitris Tousoulis**

Department of Cardiology, Hippokration Hospital, University of Athens, Athens, Greece

**Deepak Voora**

Duke Center for Applied Genomics & Precision Medicine, Durham, NC, USA

**Effimia Zacharia**

Department of Cardiology, Hippokration Hospital, University of Athens, Athens, Greece

**Tanja Zeller**

University Heart Center Hamburg, Clinic for General and Interventional Cardiology, Hamburg, Germany; German Center for Cardiovascular Research, partner site Hamburg/Lübeck/Kiel, Hamburg, Germany

**Cover Image Illustrator**


---

Zhijing Xu, Shanghai, China

**Executive Typesetting Editor**


---

Li Wang, AME Publishing Company

## Will scholarly journals perish?

Will scholarly journals perish? This is a question that has puzzled me for years.

The introduction of online journals has resulted in the inevitable recession of print journals. The uprise of the open access journals has been changing the structure of scholarly journals ceaselessly. What keeps turning over in my mind is the open access of clinical trials data. What would the bigger picture be if open access to clinical trials data became the mainstream?

It is interesting that with the primary bottleneck being the availability of open data, the Big-data Clinical Trial (BCT) seems to stay where it was in spite of the increasingly popularity of “Big Data” among scientists. It has due to the fact that the fact that without open data, a statistical analysis is restricted to a particular area (or several areas). Even with big enough data, the study can only be termed as “research with big data sets” rather than “big data research”, which are totally different concepts. Big Data is constituted by a plurality of dimensions. On one hand, for an individual (e.g., a patient), the relevant data covering his/her disease course is large enough; on the other hand, for the entire population, as more as individuals (e.g., patients) are expected to be included, to contain all the elements just like the “universe set” in set theory; by doing so, scientists expect to carry out the so-called clinical studies in real-world settings.

Why do real-world-based clinical trials so appealing? It is understandable that there is a likelihood that the results and conclusions will be altered in studies targeting the same issue using the same research method with the sample size changed. Indeed, the probability of such a likelihood is quite high. In many top journals, it is a common phenomenon that some authors tend to validate the results of one study in another population using the same research method. However, if the results are “validated” in one population, it only means that they are “repeatable”. Will the results also be repeatable in the second, third, and populations beyond? If the attempts are not extensively iterated as they should be, this “validation” is, in a sense, equivalent to self-deception.

When clinical research data is open accessed, we can easily integrate data from multiple centers for statistical analysis and meanwhile “validate” the results in multiple populations. If this is the case, then another question arises: can everyone easily publish his/her results/papers in high-profile journals such as the *New England Journal of Medicine*? My answer is NO.

When the open access to clinical research data becomes mainstream, it will be easy to have access to a constantly updated database on the Internet. Simply by clicking on a button, we could obtain the statistical results of the most current data. Another click would display the validation results based on a specific population. The database would be updated at regular intervals (e.g., 1 month or 1 day), and the statistical results would likely also be changed accordingly. At that time, the question may change to “would any researchers publish their findings in a journal?” Well, even if someone is still keen to write such articles, journals may be reluctant to publish them because of the indefiniteness of the findings with the risk of being overturned at anytime.

Eventually here it comes the serious question: will scholarly journals perish? My answer is still NO. In what way then can scholarly journals continue to be relevant?

During my Business Administration course, my teacher distributed to us an article from the Case Study column of the *Harvard Business Review*. In this highly respected journal, articles in this column often present one case first, followed by the comments from two experts. These comments could either support or oppose each other. My teacher asked us to study the case, read through the comments and then form our own point of view on the case. He encouraged us to interpret the case from different perspectives independently, which I found to be a fairly practical exercise.

Thinking of this course brought up a possible answer to me. When the open access to clinical research data becomes mainstream, the entire publishing industry, especially the publication of “scholarly journals”, would eventually experience revolutionary change. It may no longer focus on rigid and cold outcomes but would surely shift focus towards the reflection on problems, the update of insights, and the integration of science and arts.

*AME Medical Review Series* is a product born of this new mentality. As an attempt, we decided to invite international experts to provide their views on a specific topic to share their insights with more clinicians with the aim that this will ultimately benefit more patients. The first chosen topic for the series is the currently controversial one: conventional surgery versus

## VIII

stereotactic body radiotherapy for the early stage lung cancer. As the first book to the series, we hope it will give you a glance of the coming changes.

The book series will be written by a group of individual experts who are willing to contribute medical reviews and comments for those readers who are specially interested in clinical research and medical reviews. While the book in your hand may be about a difficult subject, we do hope we have presented it in an accessible manner. We would be overjoyed if it can, in any way, bring you thought and inspiration.

**Stephen D. Wang**  
Founder and CEO,  
AME Publishing Company

In recent years, research on new types of myocardial infarction markers has been supported by the Ministry of Science and Technology, the National Natural Science Foundation of China, and the Chinese Academy of Sciences. Yang and colleagues have made remarkable effort in this field. The editorial team of this book has been committed to reporting on the discoveries made by the research on new markers of myocardial infarction. Moreover, in the past few years, it has also shared a number of team research results in various academic journals.

As human beings continue to understand the laws of life, the focus and fashions of scientific research are constantly changing. Recently, the field of life sciences has undergone tremendous change, from Genomics to Proteomics to RNomics. The study of non-coding RNA represented by microRNA has become a hotspot for research and an expanding frontier in the life science disciplines of cell biology, developmental biology, neurobiology, and molecular immunology among others. The editorial member of this book have conducted an in-depth exploration of miRNAs in plasma samples of patients with acute myocardial infarction. This book contains all the research results to date, which include the relationship between acute myocardial infarction and miRNA, the function and mechanism of common miRNAs related to myocardial infarction, such as miRNA-1, 138, 22, 197, 233, etc. and research findings and remarks by researchers at home and abroad in many related fields.

Currently, there are still many gaps in our understanding of the study of new myocardial infarction markers. The purpose of this book is to illuminate more of this uncertainty, and to inspire peers engaged in their own research on new types of myocardial infarction markers. It is hoped that it can act as a great guiding influence for medical workers, and it play a significant role in the continuing research of new myocardial infarction markers, the analysis of microRNA in patients with acute myocardial infarction, and the improvement of diagnosis and treatment of a AMI, while promoting the further development of research in related fields both here and abroad.

The editorial members and the research team should continue to accumulate and update the evidence in this area, provide feedback to the reader, so as to offer readers a broader platform for the development of new markers of myocardial infarction. From reading this book, I have experienced the enterprising spirit which has strived for the continued discovery of new markers of acute myocardial infarction. Accordingly, I am very gratified to write this foreword and pleased to offer congratulations in completing the publication of *Key Leaders' Opinion on MicroRNA and Myocardial Infarction*.



**Junbo Ge, MD**

Shanghai Institute of Cardiovascular Diseases,  
Zhongshan Hospital, Fudan University,  
Shanghai, China

## miRNAs in CVD and myocardial infarction

Over the last century, cardiovascular diseases (CVD) like myocardial infarction (MI) and stroke have overtaken infectious diseases as the leading cause of mortality worldwide. Researchers have put tremendous efforts into clinical and epidemiological studies, enabling us to unravel large parts of the etiology of the underlying risk factors leading to cardiovascular pathologies. These factors include ageing, arterial hypertension, diabetes, hyperlipidaemia, obesity and tobacco smoking, all of which result in atherosclerosis that becomes the main trigger for ischemic forms of CVD like MI. Despite some crucial revelations, CVD will likely remain the number one reason of death in the foreseeable future, while our current knowledge even predicts an exacerbation in the prevalence of CVD. Thus, in-depth investigations into key determining molecular and epigenetic mechanisms have been initiated, hopefully enabling us to discover novel and powerful markers of disease as well as potential therapies. Recent advances in the field have enabled us to unravel and functionally assess the role of non-coding RNAs (ncRNAs) in disease development and progression.

Insights into the transcriptomic regulation of different species were able to show that approximately 80% of our genome is getting transcribed. However, only 1–2% of this gets translated into protein, generating a large number of ncRNA transcripts. This ncRNA was long believed to be a sequencing artefact or simply non-functional ‘junk DNA’. Over the past 10–15 years however, it has been indicated that ncRNA are key mediators and ‘fine-tuners’ in gene expression and epigenetic control, in particular in different stages of disease development and progression. We are still only scratching the surface of understanding the different forms and subclasses of ncRNAs, and the regulatory function they play in the homeostasis of an organism. These ncRNA subspecies include small nuclear and small nucleolar RNAs, microRNAs (miRNAs), Piwi-interacting RNAs, Y-RNAs and the extensive group of long-noncoding RNAs, which include long-intergenic-noncoding RNAs and natural antisense transcripts. Another interesting form of ncRNAs are circular RNA, which appear to be promising biomarkers due to their stable expression and extended protection from degradation within the circulation.

Currently, miRNAs appear as the most extensively studied and best characterized ncRNA subgroup. miRNAs are well-conserved and upon maturation in the cytoplasm 18–22 nucleotides short transcripts that can act as key post-transcriptional regulators of gene expression not only in humans, but also plants, nematodes, yeast, and other animals. By now, miRNAs have been identified to play major roles in almost every biological process via mediating the translation of target messenger RNAs (mRNAs) and thus their stability.

This present book entitled “*Key Leaders’ Opinion on MicroRNA and Myocardial Infarction*” focuses mainly on the *in vivo* role of miRNAs in the pathogenesis of developing and existing myocardial disease. The therapeutic potential of targeting miRNAs and their contribution as biomarkers in CVD pathologies is thoroughly presented. Numerous preclinical experimental CVD models have indicated that either inhibition or induction/overexpression of a single miRNA can augment or diminish CVD development and/or progression.

miRNA modulation can be powerfully enforced by utilizing so called antimiRs or antagomiRs (synthetic antisense oligonucleotides) that bind and silence miRNA expression, or by using miRNA mimics or pre-miRNAs that act similarly to endogenous miRNAs. Human and animal efficacy data exists that implies the promising role antimiRs might play in disease management. Important features include the frequent conservation of target miRNAs across species and the small molecule size. Several candidate miRNA inhibitors have shown that they can silence their putative miRNA with convincing affinity and specificity. Intriguingly, until now no immunogenic or toxicity issues were reported in human Phase I and II clinical trials. One disadvantage of targeting miRNAs in CVD might be their ubiquitous expression throughout organs and different tissues of the human body. This favors local and cell type specific tools for miRNA modulator delivery, which would limit undesired off-target effects in organ systems in which anti-miRNAs can accumulate to a much higher extent than the targeted cardiovascular system. It will be interesting to see the results of the first miRNA-based therapy trials in CVD. Several candidates exist, with anti-miR-92a in patients with myocardial ischemia being the current frontrunner for a CVD-miRNA-therapy trial in humans. First-in-patient studies and subsequent data is here expected for 2019.



Lars Maegdefessel

**Lars Maegdefessel<sup>1,2</sup>, MD, PhD**

<sup>1</sup>Molecular Vascular Medicine, Karolinska Institute,  
Center for Molecular Medicine L8:03,  
17176 Stockholm, Sweden;

<sup>2</sup>Experimental Vascular Surgery and Vascular Biology Laboratories,  
Technical University Munich, Klinikum rechts der Isar,  
Department of Vascular and Endovascular Surgery, Munich, Germany



Despite the past development of drugs, i.e. statins, that efficiently lower plasma cholesterol levels and thereby reduce the chance of developing atherosclerotic lesions, the risk to die from a cardiovascular event remains very high. More specifically, death of myocardial infarction is still the predominant cause of death worldwide. As such, there is clear need for the development of alternative cardiovascular therapies. In this light, it is of interest to see that many associations have been found between differences in the cellular expression and/or plasma level of a specific class of small non-coding RNA molecules, the ~22 nucleotide long so-called microRNAs, and the occurrence of myocardial infarction. MicroRNAs through binding to their target transcripts negatively impact on the gene expression of a wide variety of proteins, and - in this way - are able to modulate many processes involved in the development of atherosclerotic lesions, the de-stabilization of these lesions, and the generation and recovery from a myocardial infarction. Notably, microRNAs constitute valuable drug targets as they can be readily administered to humans. Furthermore, their activity can also be easily diminished through administration of antagonizing nucleotides, i.e. antagomirs. When taking the aforementioned findings into account, one can foresee a bright future for microRNAs in cardiovascular drug development. Providing insight in the (causal) contribution of microRNAs to cardiovascular disease pathology, i.e. through basic proof-of-principle and clinical studies, will hopefully aid in a speedy development of novel microRNA-based cardiovascular therapies.

**Menno Hoekstra, PhD**

Assistant Professor,

Division of BioTherapeutics,

Leiden Academic Centre for Drug Research,

Leiden, The Netherlands

(Email: Hoekstra@lacr.leidenuniv.nl)

*“MicroRNAs have it all”.*

This book is the first edition of a series of scientific papers from Key Opinion Leaders' in the field of microRNAs and myocardial infarction. The reader will be guided through an extensive review of the available literature related to the role of microRNAs in myocardial infarction.

MicroRNAs (miRNAs) are small single-stranded RNA molecules that regulate target gene (messenger RNA) expression, either through inhibition of translation or activation of degradation. More than 2,500 mature human miRNAs have been characterized so far. They are ubiquitously expressed and are involved in virtually all pathophysiological processes.

In the heart, miRNAs have been extensively described. Some of them have been shown to be enriched in skeletal muscle, the so-called myomiRs (miR-1/133a-b/206/208a-b/486/499). These miRNAs have attracted a lot of attention for their potential to diagnose myocardial infarction or to aid in fighting heart failure. Since their discovery in the circulation (1), miRNAs have been the topic of a plethora of studies, which have been facilitated by the development and commercialization of multiple kits and molecular biology tools. These tools allowed to measure circulating miRNAs and manipulate their expression levels in cells, thereby allowing to test both their biomarker value and their therapeutic potential. Hence, miRNAs have appeared as promising candidates for Theranostics\*, with some potential to advance the development of personalized and precision medicine. In the context of myocardial infarction, miRNAs have the potential to aid implementing personalized healthcare since they have some diagnostic and prognostic value and are involved in the development of heart failure that sometimes occurs after myocardial infarction (2).

However, much remains to be done for a complete knowledge of the role of miRNAs in atherosclerosis, plaque rupture, left ventricular remodeling, heart failure (...). Cost-effective molecular diagnostic kits have to be developed to allow a reliable measurement of circulating miRNAs. The therapeutic value of novel strategies to regulate the expression levels of miRNAs has to be tested.

This book gathers essential information on the emerging role of miRNAs in myocardial infarction, from biomarker to functional and translational studies. It provides directions for future research and reveals challenges that remain to be addressed before miRNAs can find their practical application for the benefit of myocardial infarction patients (3).

## References

1. Mitchell PS, Parkin RK, Kroh EM, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 2008;105:10513-8.
2. Goretti E, Wagner DR, Devaux Y. miRNAs as biomarkers of myocardial infarction: a step forward towards personalized medicine? *Trends Mol Med* 2014;20:716-25.
3. Goretti E, Devaux Y. Which future for circulating microRNAs as biomarkers of acute myocardial infarction? *Ann Transl Med* 2016;4:440.

---

\* Theranostics is a new biomedical field where targeted diagnostic tests are combined with specific targeted therapy.



Yvan Devaux

**Yvan Devaux, PhD**  
Cardiovascular Research Unit,  
Luxembourg Institute of Health, Luxembourg;  
Cardiolinc Network  
(Email: [yvan.devaux@lih.lu](mailto:yvan.devaux@lih.lu))

Myocardial infarction is pathologically defined as myocardial cell death due to prolonged ischemia, which is the most severe manifestation of coronary artery disease. A sudden rupture of plaque and formation of thrombus leads to acute myocardial infarction. Once the oxygen supply is occluded, the onset of myocardial infarction is initiated as little as 20 min after and the complete myocardial cell necrosis happens in a few hours. Prolonged ischemia leads to the loss of heart contractility due to the poor proliferation capability of the myocardial cell. The timely revascularization of the occluded artery is the key for myocardial infarction therapy. However, these treatments only reduce the severity of coronary artery disease, rather than restoration of the contractility of the infarcted heart. Therefore, novel therapeutic strategies to reduce the myocardial cell death and/or stimulate heart regeneration are highly desirable for the future.

Cardiac cell death plays a critical role in the pathogenesis of myocardial infarction, due to the terminal differentiation and loss of regenerative ability of cardiomyocytes. Myocardial infarction usually involves three main types of cell death process including apoptosis, necrosis, and autophagy. MicroRNAs are defined as single-stranded non-coding RNAs with around 22 nucleotides. Cardiac cell death processes are regulated by a variety of molecules, among which microRNAs have shown outstanding regulatory functions. Recently, a large body of research has emphasized the importance of microRNAs in regulating apoptosis, necrosis, and autophagy in cardiomyocytes, which play a decisive role in myocardial infarction. Nevertheless, more regulatory pathways and the link between different pathways of microRNAs regulating cardiac cell death need to be clarified urgently.

The significance of microRNAs in regulating myocardial infarction has been well emphasized by multiple studies. Several microRNAs are downregulated/upregulated depending on the type of myocardial injury. The significant changes in their expression pattern upon myocardial infarction highlights their contribution in regulation of pathogenesis of myocardial infarction.

The regenerative capacity largely declines within seven days after birth and remains very low in the adult heart. Thus, the activation of endogenous heart regeneration and the triggering of cardiomyocytes renewal could provide new clues for the therapy to treat myocardial infarction. Different approaches have been proposed to regenerate new cardiomyocytes: (I) to promote resident cardiomyocytes proliferation by inducing them to re-enter the cell cycle; (II) to activate endogenous stem cells or progenitors such as cardiac stem cells differentiation; (III) to stimulate endogenous regeneration through direct reprogramming from cardiac fibroblasts into cardiomyocytes. It has been demonstrated that microRNAs are critical regulators of these processes and exhibit as potential new therapeutic targets for myocardial infarction.

Circulating microRNAs in the blood have recently emerged as potential biomarkers for the diagnosis or prognosis of myocardial infarction due to their stability and specificity in plasma. A large body of studies explored the fact that microRNAs are leaked from the heart into the circulation after myocardial injury, during which their expression is elevated and dynamic. Circulating microRNAs are stable and can be easily quantified by real-time PCR assay. Among these abundant microRNAs in the heart, four cardiac-enriched microRNAs (miR-208, miR-499, miR-1, and miR-133) are consistently found to be increased in the plasma of acute myocardial infarction patients. Although the potential value of microRNAs as biomarkers has been established in small-scale studies, it is difficult to validate them in large cohorts of patients with myocardial infarction. In addition, the methods of microRNAs detection need to be optimized. The standardized assays for the detection of microRNAs in patients may reduce the inconsistency and microRNAs may become potential biomarkers for diagnosis of myocardial infarction patients. More prospective studies are underway to assess the diagnostic value of microRNAs as biomarkers. The sensitivity and specificity of circulating microRNAs have an attractive prognostic value in response to myocardial infarction, and relevant research is being designed and carried out.

Accumulating evidence reveals that microRNAs function as pro- or anti-myocardial infarction factors through their influence on myocardial cell death and cardiomyocyte regeneration pathways. So far, significant progress has been made to unveil the microRNAs-regulated signaling pathways of myocardial infarction, which has improved our understanding of heart pathogenesis.

**Alberto Dominguez-Rodriguez, MD, PhD, FESC**

Department of Cardiology, Hospital Universitario de Canarias,  
Ofra s/n La Cuesta E-38320, Santa Cruz de Tenerife, Spain;  
Facultad de Ciencias de la Salud, Universidad Europea de Canarias,  
La Orotava, Santa Cruz de Tenerife, Spain  
(Email: [adrvdg@hotmail.com](mailto:adrvdg@hotmail.com))

An increasing number of studies have recently demonstrated that the human genome is dynamically transcribed and leads to the production of a complex world of RNAs. Of those, there is a wide spectrum of RNAs that do not encode proteins, named as non-coding RNAs (microRNAs, miRNAs), whose role remained unclear for many years. Evidence suggests that miRNAs are associated with human disease. Recent studies indicate that altered expression and function of miRNAs is strongly associated with cardiovascular disease including arrhythmias, hypertrophy and myocardial infarction.

Several methods have been developed to study the expression and quantification of miRNAs, but each individually still has practical issues and technical limitations which need to overcome in order to adapt the use of miRNAs in routine clinical practice. Despite the recent advances and promising results associating miRNAs with myocardial infarction, there is still a large proportion of studies that failed to show superiority of miRNAs over classic biomarkers. miRNAs could be of value as adjunctive and combinative biological tools, focusing particularly on the early diagnosis of acute myocardial infarction and to distinguish unstable angina from other non-cardiac causes of chest pain. However, there is need for more clinical studies to evaluate the role of miRNAs in myocardial infarction as stable and sensitive biomarkers.

In the present book, we highlight the potential role of miRNAs as diagnostic tools as well as possible therapeutic targets in cardiovascular disease and more specifically in myocardial infarction. We hope that this book will trigger further research on the role of miRNAs in this field.



Nikolaos Papageorgiou

**Nikolaos Papageorgiou, MD, PhD, FESC**  
Electrophysiology Department,  
Barts Heart Centre,  
St. Bartholomew's Hospital,  
London, UK  
(Email: drnpapageorgiou@yahoo.com)



Over the past decades cardiac biomarkers have become a cornerstone in the diagnosis of acute myocardial infarction (AMI) and cardiac injury of other causes, and more recently of heart failure as well. We are beginning an era when it may be possible for biomarkers to direct treatment to optimize patient care, which must be the final goal with all routinely used cardiac biomarkers. A currently very popular research topic is circulating plasma micro ribonucleic acid (miRNA) testing. This book excellently compiles the current knowledge on miRNA testing in patients with AMI and other cardiac diseases. miRNAs have been first described in the early 1990s, they are small (typically less than 25 nucleotides), single-stranded, endogenous, non-coding RNAs that post-transcriptionally regulate gene expression by destabilizing messenger RNA (mRNA) or translation repression, thereby preventing proteins synthesis. More than 1,000 miRNAs have been identified in the human genome so far. Eventually miRNAs are also secreted from cells into blood, but the biological function of circulating miRNAs still remains to be established. Nonetheless, circulating miRNAs are interesting and attractive candidates for routine laboratory applications, and panels of miRNAs could provide clinically useful information for AMI diagnosis, risk stratification and treatment monitoring. Thus, miRNAs have already been evaluated as diagnostic and prognostic biomarkers in a variety of human diseases, in particular cancer or cardiovascular disorders, which yielded numerous associations between miRNAs and different types of diseases. Although examples exist where deregulated expression of a single miRNA is indicative for a disorder, the simultaneous analysis of the expression of multiple miRNAs usually provides more accurate information. But the interpretation of large panel test results may be challenging requiring expert knowledge in bioinformatics, which clearly is a limitation for routine use.

The next steps for making the way of miRNA testing from research to routine use in cardiovascular diseases will be to replicate and confirm promising available clinical data of the discovery phase in large-scaled, ideally multi-centre clinical trials using a small panel of candidate miRNAs with appropriate pre-analytics and analytics in a clinically relevant patient and control population as validation. Analytical issues still need to be overcome before routine use and explain the poor replicability of circulating miRNA studies. Currently miRNA testing is still tricky, time consuming and purification steps, methods, and normalization remain to be harmonized or standardized and analytical steps automatized. These analytical issues make it very difficult to compare the sample and method dependent published clinical study results. Quantitative reverse transcription PCR (qRT-PCR) still is the most widely used method, which precludes heparin or citrate as anticoagulants of blood samples as both can inhibit PCR amplification. In addition, although circulating miRNAs are remarkably stable molecules, e.g. hemolysis during blood sampling has to be avoided and whole blood must be processed immediately for platelet free plasma to avoid in-vitro miRNA release from blood cells. Alternatively specific collection tubes in which cells of whole blood samples are directly lysed and miRNA expression is thereby stopped are available. However, a high biological variation between individuals appears to be an issue as well.

Finally, circulating miRNAs have emerged as novel biomarkers in cardiovascular diseases. But, as with other heavily investigated novel biomarkers, the coming years will show whether miRNA testing will make the way from research to routine use. This book is an excellent companion for all interested in the field of miRNA testing in cardiovascular diseases.



Johannes Mair

**Johannes Mair, MD**

Department of Internal Medicine III – Cardiology and Angiology,  
Medical University of Innsbruck, A-6020 Innsbruck, Austria  
(Email: [Johannes.Mair@i-med.ac.at](mailto:Johannes.Mair@i-med.ac.at))

Acute myocardial infarction is one of the leading causes of mortality and morbidity worldwide. On the basis of literature it was defined as myocardial cell death due to prolonged ischemia. In the last two decades the progressive developments in the field of mechanical (percutaneous coronary intervention) and pharmacological therapies have contributed to reduce myocardial damage and subsequent remodeling. Several studies published in last years, have revealed the diagnostic value of circulating microRNAs for acute myocardial infarction detection. MicroRNAs are a class of single stranded and non coding RNAs which are involved in some cardiac disorders such as myocardial infarction, cardiomyocyte hypertrophy and, also, in heart failure. However, the diagnostic capacity of miRNAs for acute myocardial infarction is still controversial; some studies demonstrated that microRNAs are involved in regulating cardiac cell death and regeneration after myocardial infarction. Infact a growing number of studies showed the role of microRNAs in regulating apoptosis, necroptosis and autophagy in cardiomyocytes. Some microRNAs such as miR-17, miR-18a, miR-19a, miR-19b has been demonstrated to be associated with cardiomyocytes proliferation; miR-548-3p, miR-509 3p have been demonstrated to be involved in regulation of cell cycle progression. These are only some correlations recently demonstrated. Furthermore an interesting alternative to compensate the cardiomyocytes loss during acute myocardial infarction it could be to stimulate proliferation and differentiation of cardiac stem cells or their progenitors. Some studies *in vitro* showed that overexpression of miR - 1 family enhances cardiac differentiation of cardiac progenitor cells. On the basis of the data we can find in the literature, microRNAs have a great potential to offer a novel potential diagnostic biomarkers in the setting of acute coronary disease and acute myocardial infarction. We expect more large scale, multicenter clinical studies and trials to validate the known data of specificity and sensitivity of microRNAs. This volume reported and summarize some recent review and studies about microRNAs and acute myocardial infarction and I hope it can answer the doubts.



Andrea Rognoni

**Andrea Rognoni, MD, FSCAI**  
Coronary Care Unit and Catheterization Laboratory  
“Maggiore della Carità Hospital”, Novara, Italy

A recent surge in the exploring noncoding RNAs in stem cell research has ignited a field of discovery into many diseases including heart diseases. Heart diseases continue to be among the leading causes of death in Western countries. The most common heart diseases such as Myocardial infarction, atherosclerosis and hypertension which are mainly affect the heart, lungs and the blood vessels. Several independent risk factors have been identified and shown to be responsible for cardiovascular diseases. Although important progress has been made in the treatment of heart failure during the last decade, most interventions relieve symptoms or prevent disease progression. Thus, an improved knowledge and treatment or curative regimen is desperately needed. Recently, scientists have demonstrated that RNA functions not only as an intermediate molecule between DNA and protein, but also plays a critical role in regulating gene expression. Some of the RNAs are functional in cells but do not encode proteins. Hence, these RNAs are called noncoding RNAs (ncRNAs). Approximately 10% of the host genome consist of ncRNAs and are occupied at intergenic or intragenic regions. These ncRNAs play an important role in regulating genes that are involved in controlling the transcriptional or translational pathways. Importantly, ncRNAs are having diverse biological functions like development, differentiation, growth, and metabolism. Among ncRNAs, the short interfering RNAs and microRNAs (miRNAs) have been extensively studied, but their specific functions yet to be identified. In recent years, miRNAs are efficiently studied as one of the important candidates for involvement in most biological processes and have been implicated in many human diseases. Thus, the identification and the respective targets of miRNAs may provide novel molecular insight and new therapeutic strategies to treat diseases.

Number of recent studies has shown that miRNAs are essential for the normal development and physiology of various organs, including the heart. Studies have also started to characterize the link between miRNAs and different aspects of cardiac pathogenesis as well as proliferation, differentiation, function and maintenance of cardiac cells. Moreover, congenital heart anomalies can be associated with the dysregulation of specific miRNAs. The recently developed high-throughput approaches revealed the miRNA size, and their target, and the connectivity of the miRNA-dependent regulatory network. One step further, the expression levels of miRNAs and their decay rates have been identified in individual cell types. These works together help us to understand miRNA-dependent gene regulation to study the response of the entire network. During the past decade, numerous research articles have shown a wide knowledge about the basic mechanisms of miRNAs, biogenesis and its functions in the circulatory system. Although miRNAs are richly expressed in the heart, relatively little is explored about the multi-functional effect of these molecules in the heart.

Circulating miRNAs in patients with MI have been examined very recently. It was reported that miRNAs served as a critical modulating regulator, which participates in almost all aspects of cardiovascular diseases and vascular biology. Studies demonstrated that miRNAs also existed in blood, in the form of circulating miRNAs and are resistant to endogenous ribonuclease activity and can be present in a remarkably stable form during pathological conditions. More importantly, circulating miRNAs are currently explored as biomarkers in a wide range of cardiovascular conditions, including atherosclerotic disease. Based on genome-wide studies, thousands of miRNAs exist, however as of now only a limited number of functional miRNAs have been sequenced, identified and characterized. It takes part actively in regulating splicing, localization, stability, and translation of the target mRNAs. MicroRNAs that are capable of interfering with either complete or partial complementary to the cellular mRNAs, would be a useful treatment strategy for various diseases. I hope that the literature and examples provided here will illustrate the diversity of mechanisms regulating miRNAs while protecting the young and old heart. Certainly, many more needs to be discovered and the full potential of miRNA as therapeutic agents to be revealed. Thus, miRNAs have a promising and an exciting future in the field of research.

In this book, studies have highlighted the mechanisms by which miRNAs regulate various biological functions in model systems and could be a potential molecular therapeutic target for various heart diseases. Moreover, the authors discussed the functional roles of miRNAs and its potential use of diagnostic biomarker for cardiovascular diseases, as well as the limitations and challenges in miRNA-based therapy.



Johnson Rajasingh

**Johnson Rajasingh, PhD, HCLD (ABB)**  
Associate Professor,  
Department of Cardiovascular Medicine,  
University of Kansas Medical Center,  
Kansas City, KS, USA

# Table of Contents

## Emerging Role of miRNA in Myocardial Infarction

- 1 **Which future for circulating microRNAs as biomarkers of acute myocardial infarction?**  
*Emeline Goretti, Yvan Devaux*
- 4 **Micro-RNAs as promising biomarkers in cardiac diseases**  
*Mariama Akodad, Mathias Mericskay, François Roubille*
- 8 **Circulating micro-RNAs as biomarkers of coronary artery disease: is it ready for primetime or still a work in progress?**  
*Surovi Hazarika, Brian H. Annex*
- 11 **Sarco“MiR” friend or foe: a perspective on the mechanisms of doxorubicin-induced cardiomyopathy**  
*Louis A. Saddic, Jochen D. Muehlschlegel*
- 17 **Molecular evidence that exercise training has beneficial effects on cardiac performance**  
*Marek Kiliszek, Urszula Mackiewicz, Michal Maczewski, Beata Burzynska*
- 21 **How to be young at heart? miR-22 as a potential therapeutic target to boost autophagy and protect the old myocardium**  
*Sebastiano Sciarretta, Elena De Falco, Giacomo Frati, Junichi Sadoshima*
- 26 **Another promise against ischemia reperfusion injury: every success raises new questions**  
*Dennis V. Cokkinos*
- 30 **RNAs that make a heart beat**  
*Mithun Mitra, Hilary A. Collier*
- 38 **Circulating fibrocytes serve as a marker for clinical diagnosis**  
*Thuy Cao, Sheeja Rajasingh, Johnson Rajasingh*
- 42 **Measuring soluble CD40 ligand: it is a fancy prognostic biomarker in STEMI-patients?**  
*Alberto Dominguez-Rodriguez*
- 44 **Corin as novel biomarker for myocardial infarction**  
*Hans-Josef Feistritzer, Bernhard Metzler*

## MicroRNA and AMI

- 47 **miRNA-197 and miRNA-223 and cardiovascular death in coronary artery disease patients**  
*Esteban Orenes-Piñero, Francisco Marín, Gregory Y. H. Lip*
- 50 **miR-126: a potential new key player in hypoxia and reperfusion?**  
*Sabina P. W. Guenther, Sonja Schrepfer*



- 53 **miR-21 alters circulating Treg function in vascular disease—hope for restoring immunoregulatory responses in atherosclerosis?**  
*Emer E. Hackett, Frederick J. Sheedy*
- 56 **The role of circulating microRNAs in acute coronary syndromes: ready for prime time?**  
*Gert Klug, Bernhard Metzler*
- 59 **Should we expect novel biomarkers of myocardial infarction?**  
*Marek Kiliszek, Agata Maciejak*
- 61 **The hunt for fatal myocardial infarction biomarkers: predictive circulating microRNAs**  
*Francesco Russo, Milena Rizzo, Kirstine Belling, Søren Brunak, Lasse Folkersen*
- 65 **Correlations between microRNAs and their target genes in skeletal myoblasts cell therapy for myocardial infarction**  
*Andrea Rognoni, Chiara Cavallino, Francesco Rametta, Angelo Sante Bongo*
- 68 **Is the regulation of SIRT1 by miRNA-34a the key to mesenchymal stem cell survival?**  
*Michael A. Bellio, Wayne Balkan, Joshua M. Hare, Ivonne Hernandez Schulman*
- 71 **My heart will go on—beneficial effects of anti-MiR-30 after myocardial infarction**  
*Yubuang Li, Lars Maegdefessel*
- 74 **MicroRNA-499-5p: a therapeutic target in the context of cardiovascular disease**  
*Menno Hoekstra*

## Challenges

- 77 **Early detection of myocardial infarction—microRNAs right at the time?**  
*Nicolle Kränkel, Stefan Blankenberg, Tanja Zeller*
- 81 **Circulating micro ribonucleic acids in cardiovascular disease: a look beyond myocardial injury**  
*Johannes Mair*
- 84 **Clinical utility of novel biomarkers in acute myocardial infarction**  
*Thomas Stiermaier, Holger Thiele, Ingo Eitel*
- 87 **Circulating microRNA biomarkers for cardiovascular risk prediction: are we approaching clinical application?**  
*Maurice W. J. de Ronde, Yigal M. Pinto, Sara-Joan Pinto-Sietsma*

## Future Prospect

- 91 **Exosomes: scytales in the damaged heart**  
*Lara Ottaviani, Leon J. De Windt, Paula A. da Costa Martins*
- 95 **Is there a role for microRNAs as novel predictors of prognosis in myocardial infarction?**  
*Robert Adam, Dominic Kelly*
- 97 **MicroRNAs to take the place of collateral flow index measurements and Rentrop scoring?—Reply to Papageorgiou *et al.***  
*Nazanin Hakimzadeh, Jan J. Piek*

- 99** **Circulating extracellular vesicles containing miRNAs may have utility as early biomarkers for cardiac injury**  
*Bethany Doran, Deepak Voora*
- 102** **Long non-coding RNAs in heart failure: a promising future with much to learn**  
*Samir Ounzain, Thierry Pedrazzini*
- 105** **Long non-coding RNAs in heart failure: an obvious lnc**  
*Hamid El Azzouzi, Pieter Adrianus Doevendans, Joost Petrus Gerardus Sluiter*
- 110** **Association between microRNAs and coronary collateral circulation: is there a new role for the small non-coding RNAs?**  
*Nikolaos Papageorgiou, Effimia Zacharia, Dimitris Tousoulis*



# Which future for circulating microRNAs as biomarkers of acute myocardial infarction?

Emeline Goretti, Yvan Devaux\*

Cardiovascular Research Unit, Luxembourg Institute of Health, Luxembourg, Luxembourg

\*Y Devaux is member of Cardioline network ([www.cardiolinc.org](http://www.cardiolinc.org)).

*Correspondence to:* Yvan Devaux, PhD. Cardiovascular Research Unit, Luxembourg Institute of Health, 84 Val Fleuri, L-1445 Luxembourg, Luxembourg. Email: [yvan.devaux@lih.lu](mailto:yvan.devaux@lih.lu).

*Provenance:* This is a Guest Viewpoint commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Wang KJ, Zhao X, Liu YZ, *et al.* Circulating MiR-19b-3p, MiR-134-5p and MiR-186-5p are Promising Novel Biomarkers for Early Diagnosis of Acute Myocardial Infarction. *Cell Physiol Biochem* 2016;38:1015-29.

Submitted Sep 29, 2016. Accepted for publication Oct 04, 2016.

doi: 10.21037/atm.2016.11.21

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.11.21>

Cardiovascular disease remains the first cause of death and disability around the world according to the World Health Organization ([http://www.who.int/cardiovascular\\_diseases](http://www.who.int/cardiovascular_diseases)). An estimated 31% of deaths worldwide are due to cardiovascular disease and, in many countries, cardiovascular disease is responsible for more than twice as many deaths as cancer (1). Coronary heart disease and stroke are especially devastating and, among coronary heart disease, acute myocardial infarction (AMI), known as “heart attack”, is the most common and the most deadly condition. Obstruction of a coronary artery leads to AMI and to the necrosis of a part of the heart due to rupture of blood supply. An early diagnosis of AMI in patients presenting with chest pain is necessary to rapidly restore blood flow to the heart to limit the extent of myocardial necrosis, which largely impacts patient outcome. Currently, AMI diagnosis is based on electrocardiogram findings and measurements of blood biomarkers of myocardial damage, among which cardiac troponins (cTns) are the most widely used. High-sensitivity troponin assays have been developed, but they suffer from a lack of specificity since elevation of cTn levels can be due to non-cardiac causes. Therefore, there is an unmet need for novel, early and specific biomarkers of AMI.

In the early 2000's, a new class of RNA molecules called microRNAs (miRNAs) emerged. miRNAs are small 20–22 nucleotides-long single-stranded non-coding RNAs able to down-regulate the expression of protein-coding genes,

either through inhibition of the translation of target messenger RNAs or induction of their degradation (2). In the heart, miRNAs are widely expressed and regulate multiple physiological and pathological pathways such as apoptosis, fibrosis or angiogenesis (2).

The discovery by Mitchell and co-workers that miRNAs are present and stable in the bloodstream (3) triggered a wealth of investigations of their biomarker potential. Of note, circulating miRNAs can be either released by dying cells or be actively secreted by living cells, acting as paracrine factors. The former possibility led to the hypothesis that circulating miRNAs emanating from dying cardiomyocytes after AMI might constitute a novel class of biomarkers of AMI. This hypothesis was tested by multiple groups and led to the publication of many reports since 2010 [reviewed in (2)]. From animal studies and small-scale studies conducted in humans, it appeared that many miRNAs are indeed released from dying cardiomyocytes after AMI. In patients with hypertrophic obstructive cardiomyopathy, circulating levels of muscle-enriched miR-1 and miR-133a were significantly increased 15 mins after transcatheter ablation of septal hypertrophy (4), supporting the hypothesis that heart-derived miRNAs may constitute early diagnostic biomarkers of AMI. The excitement around the diagnostic potential of miRNAs for AMI was tempered by large-scale studies in AMI patients and patients with chest pain reporting that circulating

miRNAs fail to provide an incremental diagnostic value over traditional markers including cTns (2,5). This disappointing result was nevertheless limited by the fact that patients in these retrospective studies were initially diagnosed with cTns. Additional prospective studies might bring back some hope, such as a recent study from Wang and colleagues reporting that plasma levels of miR-19b-3p, miR-134-5p and miR-186-5p reached a peak in the 4 hours after admission for AMI, while cTnI showed a peak only after 8 hours (6). All three miRNAs had a robust diagnostic capacity and a 3-miRNA panel discriminated AMI patients from controls with an area under the curve close to 0.90 at admission. However, this study is limited by a low sample size (18 AMI and 20 controls) and the absence of multivariable analyses to address the added diagnostic value of miRNAs on top of existing markers.

While the benefit of using circulating miRNAs for the diagnosis of AMI may be limited, mostly due to the accuracy and rapidity of high-sensitivity cTns assays, there might exist a window of opportunity for prognostication purposes. Indeed, predicting outcome after AMI is still a challenging task. The heart failure biomarkers brain natriuretic peptides (BNPs) are poor predictors of the adverse left ventricular remodelling process leading to heart failure, mainly due to fluctuating plasma levels in the few hours following AMI (7). In two independent groups of AMI patients, plasma levels of miR-150 at admission predicted left ventricular remodelling and provided an added prognostic value over a multivariable clinical model (8). A 4-miRNA panel including miR-150 improved outcome prediction in a cohort of 150 AMI patients (9). In a recent case/control study with 198 patients, circulating levels of miR-22 were independent predictors of cardiovascular mortality in patients with systolic heart failure from both ischemic and nonischemic origin (10).

While the use of circulating miRNAs as prognostic biomarkers after AMI holds some promise, more remains to be done before these relatively novel markers can reach clinical application. Candidate miRNAs have to be extensively validated; they have to provide very accurate predictions at an early stage after AMI; they have to be stable enough in the blood to be reliably detected; they have to be insensitive to medications and other confounding factors such as age or sex; and finally their methods of detection have to be critically improved to be applicable in a clinical setting. A point-of-care device integrating miRNA measurements in a multifactorial computational model delivering a diagnosis in a relatively short period of time would constitute an attractive “New concept in

patient stratification” (Horizon 2020 Work Program 2016–2017. Health, demographic change and well-being. SC1-PM-02-2017) towards personalized healthcare.

In conclusion, although circulating miRNAs are still attracting some interest as diagnostic biomarkers of AMI, the majority of studies conducted so far concluded that miRNAs will have a hard time outperforming the sensitivity and rapidity of high-sensitivity cTn assays. On the other hand, past and recent studies support the use of circulating miRNAs to aid in risk stratification after MI. Future technological developments are needed to translate these research findings into clinical application. Also, more efforts are required before circulating miRNAs can aid controlling the growing burden of heart failure.

### Acknowledgements

Y Devaux received fundings from the Ministry of Higher Education and Research of Luxembourg and the National Research Fund of Luxembourg.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Nichols M, Townsend N, Scarborough P, et al. Cardiovascular disease in Europe 2014: epidemiological update. *Eur Heart J* 2014;35:2929.
2. Goretti E, Wagner DR, Devaux Y. miRNAs as biomarkers of myocardial infarction: a step forward towards personalized medicine? *Trends Mol Med* 2014;20:716-725.
3. Mitchell PS, Parkin RK, Kroh EM, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 2008;105:10513-10518.
4. Liebetrau C, Möllmann H, Dörr O, et al. Release kinetics of circulating muscle-enriched microRNAs in patients undergoing transcatheter ablation of septal hypertrophy. *J Am Coll Cardiol* 2013;62:992-998.
5. Devaux Y, Mueller M, Haaf P, et al. Diagnostic and prognostic value of circulating microRNAs in patients with acute chest pain. *J Intern Med* 2015;277:260-271.
6. Wang KJ, Zhao X, Liu YZ, et al. Circulating MiR-19b-3p, MiR-134-5p and MiR-186-5p are Promising Novel Biomarkers for Early Diagnosis of Acute Myocardial

- Infarction. *Cell Physiol Biochem* 2016;38:1015-1029.
7. Talwar S, Squire IB, Downie PF, et al. Profile of plasma N-terminal proBNP following acute myocardial infarction; correlation with left ventricular systolic dysfunction. *Eur Heart J* 2000;21:1514-1521.
  8. Devaux Y, Vausort M, McCann GP, et al. MicroRNA-150: a novel marker of left ventricular remodeling after acute myocardial infarction. *Circ Cardiovasc Genet* 2013;6:290-298.
  9. Devaux Y, Vausort M, McCann GP, et al. A panel of 4 microRNAs facilitates the prediction of left ventricular contractility after acute myocardial infarction. *PLoS One* 2013;8:e70644.
  10. Gupta SK, Foinquinos A, Thum S, et al. Preclinical Development of a MicroRNA-Based Therapy for Elderly Patients With Myocardial Infarction. *J Am Coll Cardiol* 2016;68:1557-1571.

**Cite this article as:** Goretti E, Devaux Y. Which future for circulating microRNAs as biomarkers of acute myocardial infarction? *Ann Transl Med* 2016;4(21):440. doi: 10.21037/atm.2016.11.21



## Micro-RNAs as promising biomarkers in cardiac diseases

Mariama Akodad<sup>1,2</sup>, Mathias Mericskay<sup>3</sup>, François Roubille<sup>1,2</sup>

<sup>1</sup>Cardiology Department, Hôpital Arnaud de Villeneuve, CHU de Montpellier, UFR de Médecine, Université Montpellier 1, 371, avenue du Doyen Gaston Giraud, 34295 Montpellier cedex 05, France; <sup>2</sup>PhyMedExp, University of Montpellier, INSERM U1046, CNRS UMR 9214, 34295 Montpellier cedex 5, France; <sup>3</sup>Université Paris-Saclay, Université Paris-Sud, Signalisation et Physiopathologie Cardiovasculaire, Inserm UMR-S 1180, LabEx LERMIT, DHU TORINO, Faculty of Pharmacy, F-92296 Chatenay-Malabry, France

*Correspondence to:* François Roubille, MD, PhD. Cardiology Department, Hôpital Arnaud de Villeneuve, CHU de Montpellier, UFR de Médecine, Université Montpellier 1, 371, avenue du Doyen Gaston Giraud, 34295 Montpellier cedex 05, France. Email: Francois.roubille@gmail.com.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, Jiangsu, China).

*Comment on:* Coskunpinar E, Cakmak HA, Kalkan AK, *et al.* Circulating miR-221-3p as a novel marker for early prediction of acute myocardial infarction. *Gene* 2016;591:90-6.

Submitted Nov 01, 2016. Accepted for publication Nov 07, 2016.

doi: 10.21037/atm.2016.12.38

View this article at: <http://dx.doi.org/10.21037/atm.2016.12.38>

### miRNA, emerging and promising biomarkers in cardiology

miRNAs are endogenous, small (21-22 nucleotides), single-stranded, non-coding RNAs recently discovered (1). miRNAs complexed with Ago proteins (RISC complex) regulate gene expression by binding to reverse complementary sequences in their target mRNAs leading to mRNA degradation and/or repression of protein translation (2). Circulating miRNAs can be detected in serum or plasma and have been proposed as potential biomarkers for cardiovascular diseases, with high sensitivity (3).

Indeed, they are involved in multiple cellular functions (proliferation, migration, differentiation...) (4) and are therefore involved in cardiac and vascular development. A dysregulation in their expression has been suggested to be responsible for cardiovascular disease. Major implications were demonstrated in several pathologies as congenital cardiac disease (5), hypertrophy, fibrosis, arrhythmias and atherosclerosis (6).

Expression of miR-378 dramatically decreased as well as expression of miR-133 and miR-1 in mouse models of heart failure and in human tissues (7,8). Concerning cardiac fibrosis, several miRNAs were incriminated as miR-133, miR-21 and miR-29, involved in fibroblasts proliferation, collagen synthesis and Connective tissue growth factor (CTGF) signaling (9). In a rat model, miR-433 was

consistently elevated in three various models of heart disease with prominent cardiac fibrosis (10).

The implication of miRNAs in heart failure was also demonstrated, particularly miR-25 whose increased expression can depress cardiac function (11). Interestingly, miRNAs secreted by cardiac fibroblasts act as paracrine mediators of cardiomyocyte hypertrophy (12).

Thereby, miRNAs appear as attractive biomarkers in cardiovascular field, easily assessed, with a robust stability in the plasma and an excellent sensitivity (13). A potential combination of several miRNAs can be considered to improve diagnostic performance.

However, few miRNA are tissue-specific and their plasma level can change depending on the physiological or the pathological situation, independently from the initial disease. Above all, there is a very large number of detectable miRNAs and only a few is likely to provide additional information compared to current validated biomarkers. Their specific interest in clinical settings both for diagnostic, prognostic and even therapeutic approaches remain largely under investigation.

### miRNA, promising in myocardial infarction as prognostic, diagnostic and therapeutic tools

In acute myocardial infarction, a rapid diagnostic is necessary to allow an immediate management of patients

**Table 1** Opened studies ongoing regarding diagnostic, prognostic or therapeutic effects of miRNA in myocardial infarction

Country	Objectives	Primary endpoint	Number of patient	Estimated primary completion date	NCT
France	(I) To study how the macrophage functions after MI are changed by diabetes  (II) To determine the potential role of miRNAs contained in secreted MVs in the transition M1/M2 after MI	Level of expression markers	20	2018	NCT02768935
China	To test the expression of microRNAs related to the syndromes after the intervention of Tongguan capsule	miRNAs spectrum	100	2017	NCT02850627
Spain	To evaluate the prognostic value of circulating miRNAs in patients admitted for STEMI complicated with cardiogenic shock	Mortality	142	2017	NCT02691286
Germany	To develop a biomarker protocol that combines the high sensitivity of cardiac Troponin T and the high specificity of mi-RNA profiles for early and safe identification of non-STEMI in ED patients	Mortality	Unknown	Unknown	NCT02116153
China	To determine whether the expression level of miR-320a are effective as biomarker in evaluating the diagnosis, prognosis and treatment effects of coronary heart disease	Plasma expression of miR-320a in coronary heart disease patients	400	2018	NCT02751060

and ensure a better prognosis, limiting long-term consequences as remodelling and fibrosis leading to chronic heart failure.

Several miRNAs were identified as promising candidates to early detect patients with MI. Among them, miR-1, miR-133a, miR-133b, miR-208a, miR-499, miR-499-5p were advocated but further validation is required (14). A large study evaluated the level of six miRNAs in 1155 patients admitted for acute chest pain. In the 224 patients diagnosed with MI, the levels of miR-208b, miR-499 and miR-320a were significantly higher (15). Thereby, miRNAs as early diagnostic biomarkers in MI seems promising but have to be identified more precisely.

In the context of MI, a potential role of miRNAs as prognostic biomarkers was also raised.

Devaux *et al.* (15) reported that miR-208b predicted survival at 30 days but none of the miRNAs could predict long-term mortality. In a study including 407 patients with a suspected MI, miR-208b and miR-499-5p were identified as potential diagnostic biomarkers of MI with an area under the curve (AUC) around 0.8 and were also predictive factors for outcomes with a prognostic value comparable to cTnT (16).

Taken all these considerations, miRNAs provide promising therapeutic targets. In a pig model of reperfused MI, an intracoronary injection of an anti-miR-

92a stimulates angiogenesis and could prevent cardiac remodelling (17). Recently, Gupta *et al.* demonstrated that miR-22, a key regulator of cardiac autophagy, could be an interesting target after myocardial infarction (18). Indeed, they show that miR-22 inhibition post-infarction improved cardiac function and inhibited cardiac remodelling in older mice but not young mice. Thereby, pharmacological inhibition of miR-22 could be promising, especially in older myocardium.

Opened ongoing studies regarding diagnostic, prognostic or therapeutic effects of miRNA in myocardial infarction are presented *Table 1*.

In this paper entitled “*Circulating miR-221-3p as a novel marker for early prediction of acute myocardial infarction*”, Coskunpinar *et al.* aimed to identify potential miRNAs to predict early myocardial infarction (MI).

They set 3 objectives to answer:

- (I) To compare the serum expression levels of miRNAs 1/in patients with AMI and control subjects with an acute atypical chest pain/dyspnoea and 2/in patients with STEMI and non- STEMI;
- (II) To evaluate the potential of these miRNAs to be used as novel diagnostic biomarkers for AMI in patients admitted to emergency department for acute chest pain and/or dyspnoea;

- (III) To investigate the relations between the serum levels of miRNAs with the serum levels of previously validated biomarkers, namely troponin I, cardiac risk scores and post-MI left ventricular functions.

In this study, 43 consecutive patients were included, all of them presented acute chest pain and/or dyspnoea, 27 were diagnosed acute myocardial infarction (AMI) and 16 were control subjects. The control subjects presented the same symptoms but without any diagnostic criteria for AMI, particularly no ECG modification and no rise of cardiac enzymes. The blood assessment was performed in all subjects within 4 h of onset of clinical symptoms and miRNAs expression levels were evaluated. Patients with AMI were compared to control patients and within the AMI group, STEMI patients were compared to NSTEMI patients.

The AMI group and control were comparable for baseline characteristics except for left ventricular function. Body mass index (BMI) was the only different baseline characteristics between the STEMI group and NSTEMI group, higher in the NSTEMI group ( $P=0.026$ ).

The study highlighted three major points:

- (I) Nine miRNAs were expressed differently between the control group and the AMI group, without any difference between STEMI and NSTEMI subgroups. Six of these miRNAs were upregulated while the other three were downregulated in patients with AMI. Moreover, the authors highlighted 2 miRNAs which were the most upregulated in the AMI group: miR-4290 and miR-221-3p with a fold regulation of 7.39 and 3.89 respectively. The most downregulated miRNA in this group was the miR-19b-1-5p with a fold regulation of -3.15;
- (II) miR221 was significantly positively correlated with Troponin, GRACE and SYNTAX Score while significantly inversely correlated with left ventricular ejection fraction;
- (III) miR-221-3p had the better discriminative value for the diagnosis of AMI with a ROC area under curve (AUC) of the level of 0.881 (95% CI: 0.774–0.987;  $P=0.002$ ), close to AUC for Troponin (AUC: 0.954; 95% CI: 0.892–1.000;  $P=0.001$ ).

These results are consistent with the upregulation of miR-221-3p in patients with atherosclerosis.

Here, the authors add a practical prognostic information of miRNAs demonstrating an association between the expression levels of mi-RNAs (miR-648, miR-4290, miR-

3914, miR-221-3p, miR-127-5p) and cardiac scores as well as cardiac function assessed by echocardiography, in patients with AMI. Among these miRNAs, miR-221-3p had a high discriminative value and significant relations with Troponin, GRACE and SYNTAX score and left ventricular systolic function. Likewise, this biomarker may be useful in daily practice for early prediction of AMI and could provide a prognosis value in this context.

From a pathophysiological point-of-view, miR-221-3p is suggested to facilitate the development of vulnerable coronary plaques, coronary artery atherosclerosis and severe endothelial dysfunction by using molecular pathway such as Netrin/DCC induced pathway.

This study presents three main limitations: first, a very small number of patients were included. Secondly, miRNAs expression levels were measured only once, no time-course or assessment after treatment to evaluate its effect are available. Importantly, the quantification of miRNA expression in different independent study cohorts was not performed and this will be a crucial step for further clinical development. Finally, it is striking that altogether the studies performed worldwide on plasma miRNA identification post AMI present a relatively poor overlap in the subset of differentially regulated miRNAs. This suggests potential methodological biases in the timing and handling of samples and quantification methods that will require to be solved in the future for efficient and reliable use of miRNAs detection in clinics. Alternatively, but not exclusively to the potential methodological biases, one can hypothesize that these differences in candidate miRNAs for AMI arise from different genetic backgrounds and environmental causes, which will generate a very interesting line of research for personalized medicine, a much awaited promise for the future in clinics.

Nevertheless, results presented in this study are innovative. The authors highlighted the miR-221-3p as a potential biomarker not only for early diagnostic of AMI but also for prognostic evaluation after AMI (because of its association with post-MI left ventricular systolic function). Further investigations are necessary to make clear the links between miRNAs products and pathophysiological mechanisms. Furthermore, larger clinical studies have to confirm the early predictive and the prognostic values of miR-221-3p in the context of MI in the aim to develop adapted therapeutic strategies. Finally, a key point for clinical development in this field of research will be to determine whether any of the identified miRNA can present at least an as high discriminating ROC value than

the validated Troponin I biomarker for AMI diagnostic.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281-297.
2. Baek D, Villén J, Shin C, et al. The impact of microRNAs on protein output. *Nature* 2008;455:64-71.
3. Gilad S, Meiri E, Yogev Y, et al. Serum microRNAs are promising novel biomarkers. *PLoS One* 2008;3:e3148.
4. Latronico MV, Catalucci D, Condorelli G. MicroRNA and cardiac pathologies. *Physiol Genomics* 2008;34:239-242.
5. Samanta S, Balasubramanian S, Rajasingh S, et al. MicroRNA: A new therapeutic strategy for cardiovascular diseases. *Trends Cardiovasc Med* 2016;26:407-419.
6. Chistiakov DA, Orekhov AN, Bobryshev YV. Cardiac-specific miRNA in cardiogenesis, heart function, and cardiac pathology (with focus on myocardial infarction). *J Mol Cell Cardiol* 2016;94:107-121.
7. Carè A, Catalucci D, Felicetti F, et al. MicroRNA-133 controls cardiac hypertrophy. *Nat Med* 2007;13:613-618.
8. Tritsch E, Mallat Y, Lefebvre F, et al. An SRF/miR-1 axis regulates NCX1 and annexin A5 protein levels in the normal and failing heart. *Cardiovasc Res* 2013;98:372-380.
9. Angelini A, Li Z, Mericskay M, et al. Regulation of Connective Tissue Growth Factor and Cardiac Fibrosis by an SRF/MicroRNA-133a Axis. *PLoS One* 2015;10:e0139858.
10. Tao L, Bei Y, Chen P, et al. Crucial Role of miR-433 in Regulating Cardiac Fibrosis. *Theranostics* 2016;6:2068-2083.
11. Wahlquist C, Jeong D, Rojas-Muñoz A, et al. Inhibition of miR-25 improves cardiac contractility in the failing heart. *Nature* 2014;508:531-535.
12. Bang C, Batkai S, Dangwal S, et al. Cardiac fibroblast-derived microRNA passenger strand-enriched exosomes mediate cardiomyocyte hypertrophy. *J Clin Invest* 2014;124:2136-2146.
13. Mitchell PS, Parkin RK, Kroh EM, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 2008;105:10513-10518.
14. Romaine SP, Tomaszewski M, Condorelli G, et al. MicroRNAs in cardiovascular disease: an introduction for clinicians. *Heart* 2015;101:921-928.
15. Devaux Y, Mueller M, Haaf P, et al. Diagnostic and prognostic value of circulating microRNAs in patients with acute chest pain. *J Intern Med* 2015;277:260-271.
16. Gidlöf O, Smith JG, Miyazu K, et al. Circulating cardio-enriched microRNAs are associated with long-term prognosis following myocardial infarction. *BMC Cardiovasc Disord* 2013;13:12.
17. Bellera N, Barba I, Rodriguez-Sinovas A, et al. Single intracoronary injection of encapsulated antagomir-92a promotes angiogenesis and prevents adverse infarct remodeling. *J Am Heart Assoc* 2014;3:e000946.
18. Gupta SK, Foinquinos A, Thum S, et al. Preclinical Development of a MicroRNA-Based Therapy for Elderly Patients With Myocardial Infarction. *J Am Coll Cardiol* 2016;68:1557-1571.

**Cite this article as:** Akodad M, Mericskay M, Roubille F. Micro-RNAs as promising biomarkers in cardiac diseases. *Ann Transl Med* 2016;4(24):551. doi: 10.21037/atm.2016.12.38

## Circulating micro-RNAs as biomarkers of coronary artery disease: is it ready for primetime or still a work in progress?

Surovi Hazarika, Brian H. Annex

Division of Cardiovascular Medicine, Robert M. Berne Cardiovascular Research Center, University of Virginia, Charlottesville, USA

*Correspondence to:* Brian H. Annex, MD. Division of Cardiovascular Medicine, Robert M. Berne Cardiovascular Research Center, University of Virginia, PO Box 800158, Charlottesville, VA 22908, USA. Email: Bha4n@virginia.edu.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* O Sullivan JF, Neylon A, McGorrian C, *et al.* miRNA-93-5p and other miRNAs as predictors of coronary artery disease and STEMI. *Int J Cardiol* 2016;224:310-6.

Submitted Nov 24, 2016. Accepted for publication Nov 30, 2016.

doi: 10.21037/atm.2017.01.05

View this article at: <http://dx.doi.org/10.21037/atm.2017.01.05>

MicroRNAs (miRs) are small non-coding RNAs that play a significant role in epigenetic regulation of gene expression (1,2). Several miRs have been established to play crucial roles in a spectrum of different cardiovascular diseases (3,4). Since the initial identification of detectable levels of circulating miRs in plasma (5-7), circulating miRs have become the spotlight of investigation as potential biomarkers for several diseases. Micro-RNAs in circulation have two important features of ideal biomarkers: they are remarkably stable and safe from degradation by RNases, and they are easily measured in the circulation using sensitive and specific quantitative PCR techniques. Thus, circulating miRs can be linked to specific disease processes or to specific tissue and/or cell types.

In a recent study in *International Journal of Cardiology*, O Sullivan *et al.* did microRNA profiling from plasma samples from a cohort of control subjects, patients with stable coronary artery disease (CAD) and patients presenting with ST-elevation myocardial infarction (STEMI). The authors used a set of selected 22 miRs based on miRs with known role in cardiovascular biology, and identified four miRs (miR15a-5p, miR16-5p, miR93-5p and miR146a-5p) that were differentially expressed between controls subjects versus patients with stable CAD. Out of these miRs, miR 146a-5p was significantly decreased in stable CAD *vs.* control, while the rest of the miRs were found to be elevated in stable CAD compared to controls. The authors also found one miR (miR499a-5p) that was significantly elevated

in patients with STEMI compared to control subjects. To adjust for traditional risk factors, the authors used a stepwise logistic regression model using all Framingham Heart Study (FHS) risk factors, and miR-93-5p remained significantly different between controls *vs.* stable CAD groups. Similarly, using a stepwise logistic regression incorporation using all FHS risk factors, the authors found miR-499a-5p was significantly elevated in patients with STEMI compared to controls. Further analysis using ROC curves showed that all four miRs that differed between control *vs.* stable CAD groups were significant predictors of stable CAD (AUCs of 0.67, 0.65, 0.68), while the miR-93-5p was found to be a better predictor based on the AUC curve of 0.75. In addition, addition of miR-93-5p to the FHS risk factors enhanced the discriminatory ability of FHS risk factors model to detect stable CAD. Similarly, the discriminatory ability of the FHS risk factors to detect STEMI was significantly enhanced with the addition of miR-499a -5p to the model.

This study by O Sullivan *et al.* provides an exciting step towards identification of potential circulating biomarkers for CAD. However, several notes of caution need to be considered. In the comparisons made by O Sullivan *et al.*, the miRs were quantitated using “normalized expression” levels, adjusted to the average Cp of all expressed miRs, and therefore do not reflect absolute copy numbers. Given the prediction models were done using relative levels of miRs in plasma, it is possible that the prediction models may differ

based on the normalization method used.

An interesting finding was the value of miR93-5p expression to the traditional FHS risk factors to improve ability to detect stable CAD. This can be clinically useful and warrants further evaluation. It is interesting that the finding did not hold in control *vs.* STEMI subjects, and the significance of this remains unknown. STEMI is a diagnosis made based on clinical presentation and EKG-based criteria, and given the time constraints of definitive treatment for STEMI (8,9); the utility of a blood test is very limited, except for unusual cases where diagnosis can be confounded. Even in these situations, given the very high sensitivity and specificity of the currently available biomarkers for myocardial injury such as cardiac troponins (10), the potential clinical utility of miR-499a-5p is questionable. An ideal biomarker should provide diagnostic and prognostic information that is specific and incremental to existing clinical and demographic data, and in the context of STEMI, the findings from miR-499a-5p falls short of existing cardiac biomarkers. miR-499 is encoded by an intronic region of the myosin heavy chain gene (11). Therefore, circulating levels of miR-499 likely reflect the pathogenic process of myocardial damage. Given that the comparisons in this study were specifically made between controls and patients with STEMI, it is unclear if miR-499 is specific to STEMI, or a reflection of myocyte injury in response to any form of acute myocardial infarctions. Given similar findings of miR-499 from other studies in patients with acute myocardial infarction (12-15), miR-499 may have some clinical utility in detecting any form of myocardial infarctions at a time frame before cardiac troponins are detectable as shown by Wang *et al.* (15), or if a troponin negative, but miR-499 positive group is established to have true myocardial damage.

In selecting a panel of limited miRs with known role in cardiovascular disease, O Sullivan *et al.* used a candidate biomarker approach in this study. This approach allows for stronger statistical analysis of the selected miRs, but this limits the identification of potential unknown or novel biomarkers. In addition, the possibility that combined changes in a panel of miRs may have a better predictive ability for diagnosis of CAD over a single miR was not explored in the current study.

Finally, miRs in circulation can exist as free micro-RNAs, in exosomes or micro-particles, in protein-bound complexes, or in lipid complexes (16). In this study by O Sullivan *et al.*, the authors examined the total circulating miRs, but it is prudent to consider that micro-RNAs

from each of these circulating fractions may give different information compared to miRs from total circulating fraction. miRs from these fractions may provide alternate approaches to identify circulating miRs as biomarkers, and give crucial information regarding pathophysiology of a disease process.

A biomarker is a defined characteristic that is measured as an indicator of normal biological processes, pathogenic processes, or responses to an exposure or intervention, including therapeutic interventions (17). Circulating miRs hold promise as potential biomarkers that can be indicative of these processes, but as knowledge of different circulating forms of miRs are still evolving, the identification of an ideal miR as a biomarker of CAD remains a work in progress.

### Acknowledgements

*Funding:* BH Annex is supported by 1R01HL116455, 1R01HL121635 and 2R01HL101200. S Hazarika is supported by 1K08HL130573-01.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. He L, Hannon GJ. MicroRNAs: small RNAs with a big role in gene regulation. *Nat Rev Genet* 2004;5:522-531.
2. Jonas S, Izaurralde E. Towards a molecular understanding of microRNA-mediated gene silencing. *Nat Rev Genet* 2015;16:421-433.
3. Gurha P. MicroRNAs in cardiovascular disease. *Curr Opin Cardiol* 2016;31:249-254.
4. Quiat D, Olson EN. MicroRNAs in cardiovascular disease: from pathogenesis to prevention and treatment. *J Clin Invest* 2013;123:11-18.
5. Chen X, Ba Y, Ma L, et al. Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res* 2008;18:997-1006.
6. Lawrie CH, Gal S, Dunlop HM, et al. Detection of elevated levels of tumour-associated microRNAs in serum of patients with diffuse large B-cell lymphoma. *Br J Haematol* 2008;141:672-675.
7. Mitchell PS, Parkin RK, Kroh EM, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 2008;105:10513-

- 10518.
8. O'Gara PT, Kushner FG, Ascheim DD, et al. 2013 ACCF/AHA guideline for the management of ST-elevation myocardial infarction: executive summary: a report of the American College of Cardiology Foundation/American Heart Association Task Force on Practice Guidelines: developed in collaboration with the American College of Emergency Physicians and Society for Cardiovascular Angiography and Interventions. *Catheter Cardiovasc Interv* 2013;82:E1-27.
  9. Task Force on the management of ST-segment elevation acute myocardial infarction of the European Society of Cardiology (ESC), Steg PG, James SK, et al. ESC Guidelines for the management of acute myocardial infarction in patients presenting with ST-segment elevation. *Eur Heart J* 2012;33:2569-619.
  10. Hammerer-Lercher A, Ploner T, Neururer S, et al. High-sensitivity cardiac troponin T compared with standard troponin T testing on emergency department admission: how much does it add in everyday clinical practice? *J Am Heart Assoc* 2013;2:e000204.
  11. Bhuiyan SS, Kinoshita S, Wongwarangkana C, et al. Evolution of the myosin heavy chain gene MYH14 and its intronic microRNA miR-499: muscle-specific miR-499 expression persists in the absence of the ancestral host gene. *BMC Evol Biol* 2013;13:142.
  12. Adachi T, Nakanishi M, Otsuka Y, et al. Plasma microRNA 499 as a biomarker of acute myocardial infarction. *Clin Chem* 2010;56:1183-1185.
  13. Corsten MF, Dennert R, Jochems S, et al. Circulating MicroRNA-208b and MicroRNA-499 reflect myocardial damage in cardiovascular disease. *Circ Cardiovasc Genet* 2010;3:499-506.
  14. D'Alessandra Y, Devanna P, Limana F, et al. Circulating microRNAs are new and sensitive biomarkers of myocardial infarction. *Eur Heart J* 2010;31:2765-2773.
  15. Wang GK, Zhu JQ, Zhang JT, et al. Circulating microRNA: a novel potential biomarker for early diagnosis of acute myocardial infarction in humans. *Eur Heart J* 2010;31:659-666.
  16. Creemers EE, Tijssen AJ, Pinto YM. Circulating microRNAs: novel biomarkers and extracellular communicators in cardiovascular disease? *Circ Res* 2012;110:483-495.
  17. FDA-NIH Biomarker Working Group. BEST (Biomarkers, EndpointS, and other Tools) Resource. Silver Spring (MD): Food and Drug Administration (US); Bethesda (MD): National Institutes of Health (US), 2016.

**Cite this article as:** Hazarika S, Annex BH. Circulating micro-RNAs as biomarkers of coronary artery disease: is it ready for primetime or still a work in progress? *Ann Transl Med* 2017;5(1):10. doi: 10.21037/atm.2017.01.05



# Sarco“MiR” friend or foe: a perspective on the mechanisms of doxorubicin-induced cardiomyopathy

Louis A. Saddic, Jochen D. Muehlschlegel

Department of Anesthesiology, Perioperative, and Pain Medicine, Brigham and Women's Hospital, Harvard Medical School, Boston, USA

*Correspondence to:* Jochen D. Muehlschlegel, MD, MMSc. Department of Anesthesiology, Perioperative, and Pain Medicine, Brigham and Women's Hospital, 75 Francis St., CWNL1, Boston, MA 02115, USA. Email: jmuehlschlegel@partners.org.

*Provenance:* This is a Guest Perspective commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

**Abstract:** Anthracyclines are a class of chemotherapeutics used to treat a variety of human cancers including both solid tumors such as breast, ovarian, and lung, as well as malignancies of the blood including leukemia and lymphoma. Despite being extremely effective anti-cancer agents, the application of these drugs is offset by side effects, most notably cardiotoxicity. Many patients treated with doxorubicin (DOX), one of the most common anthracyclines used in oncology, will develop radiographic signs and/or symptoms of cardiomyopathy. Since more and more patients treated with these drugs are surviving their malignancies and manifesting with heart disease, there is particular interest in understanding the mechanisms of anthracycline-induced injury and developing ways to prevent and treat its most feared complication, heart failure. MicroRNAs (miRNAs) are small noncoding RNAs that regulate the expression of mRNAs. Since miRNAs can regulate many mRNAs in a single network they tend to play a crucial role in the pathogenesis of several diseases, including heart failure. Here we present a perspective on a recent work by Roca-Alonso and colleagues who demonstrate a cardioprotective function of the miR-30 family members following DOX-induced cardiac injury. They provide evidence for direct targeting of these miRNAs on key elements of the  $\beta$ -adrenergic pathway and further show that this interaction regulates cardiac function and apoptosis. These experiments deliver fresh insights into the biology of toxin-induced cardiomyopathy and suggest the potential for novel therapeutic targets.

**Keywords:** Doxorubicin (DOX); heart failure; microRNA (miRNAs); beta-adrenergic pathway

Submitted Apr 20, 2016. Accepted for publication Apr 25, 2016.

doi: 10.21037/atm.2016.05.30

View this article at: <http://dx.doi.org/10.21037/atm.2016.05.30>

## Introduction to the biology of anthracycline toxicity in the heart

Anthracyclines, including doxorubicin (DOX), are among the most effective chemotherapeutic agents used to treat many human cancers. Unfortunately, their use is limited by adverse side effects, most notably dose-dependent cardiotoxicity. These events can manifest acutely following administration of the drug, or more commonly, months to years following cumulative exposure (1). In adults, early and late toxicity typically presents with dilated cardiomyopathy (2). Up to 5–10% of patients exposed to these drugs can have

symptomatic or radiographic evidence of heart failure (3). Even more, some studies have demonstrated an increased risk of cardiotoxicity in those patients receiving low doses of DOX ( $<300$  mg/m<sup>2</sup>) which were once thought to be considered safe (4). Children, on the other hand, tend to present with dilated cardiomyopathy that progresses to a restrictive pattern (2). Childhood cancer survivors exposed to anthracyclines can have up to a 2–5 fold increased risk of heart failure compared to those patients not exposed to these drugs (5). Given the increasing number of cancer survivors amongst adults and children, anthracycline-induced cardiotoxicity is becoming a prevalent disease despite efforts dedicated to surveillance and prophylactic



management.

DOX is thought to deliver its anti-tumor effects primarily through inhibition of the alpha isoform of topoisomerase II (Top2 $\alpha$ ). The mechanisms of toxicity in the heart seem to be much more complex especially since cardiomyocytes lack expression of Top2 $\alpha$ . These cells do, however, express the beta isoform (Top2 $\beta$ ), and cardiomyocyte specific deletion of the gene encoding this protein confers protection against DOX-induced toxicity through inhibition of apoptosis and DNA-damage (6). Another mechanism of DOX-induced cardiotoxicity involves the generation of reactive oxygen species (ROS). This was originally a very attractive mechanism to explain cardiomyocyte injury as these cells have high metabolic rates, are chock full of mitochondria, and express low levels of anti-oxidant enzymes (7). Despite experimental evidence to support this theory, clinical efforts to scavenge free radicals have not been promising (8). Newer mechanisms have focused on the role of DOX in the inhibition of pro-survival pathways such as NRG-1/ErbB (9,10), and the stimulation of inflammation through Toll-like receptors (TLRs) (11).

MicroRNAs (miRNAs) are small noncoding RNA molecules that bind to the 3' un-translated region (UTR) of mRNAs and regulate their expression (12). Recently, miRNAs have been shown to play a critical role in many elements of cardiovascular disease including ischemia (13-15) and heart failure (16-18). Naturally, this led many groups to investigate the potential role of miRNAs in the development of DOX-induced cardiomyopathy (10,19-23). Roca-Alonso and colleagues continued this mission with a comparison of the global changes in miRNA expression in adult rat ventricular cardiomyocytes (ARVCMs) through two models. The first was an acute *in vitro* model of cultured ARVCMs harvested 6 hours after a single dose of DOX. The second was a chronic *in vivo* model where rats were exposed to repeated doses of DOX (cumulative dose of 15 mg/kg) over a two week period followed by harvesting ARVCMs 3 weeks later. A reference model of cardiomyopathy generated from rats with proximal left anterior descending (LAD) artery ligation was also included in the comparison. Three members of the miR-30 family (miR-30a, miR-30d, and miR-30e) were down-regulated in at least two of three models (21). Down-regulation of miR-30b has also been documented in H9C2 rat cardiomyocytes following hypoxia/re-oxygenation (24). Interestingly, other groups that generated cardiac miRNA profiles in animal models exposed to DOX had unique signatures that did not uncover miR-30 family members

as being significantly dis-regulated (25,26). Nonetheless, there is quite a bit of variability in the timing of DOX exposure, the genetic background, and the technology used to generate these profiles.

### **The contribution of miR-30 and the $\beta$ -adrenergic pathway towards the pathogenesis of heart failure**

In cancer biology, miR-30 has been implicated as both an oncogene and a tumor suppressor. Its specific role tends to be cancer type specific (27-29). In the heart, overexpression of miR-30 directly regulated key pro-fibrotic proteins and thus may be associated with preventing the fibrosis characteristic of failing hearts (30). Another group demonstrated that up-regulation of miR-30 in cardiomyocytes blocked the up-regulation of angiotensin II-induced hypertrophy related genes and showed that increasing circulating levels of miR-30 may be used to diagnose myocardial hypertrophy (31).

Roca-Alonso and colleagues attempt to further promote miR-30 as a cardioprotective miRNA through a unique mechanism. Among the list of computationally derived predictive targets of miR-30, this group focused on three proteins in the  $\beta$ -adrenergic pathway ( $\beta_1$ AR,  $\beta_2$ AR, and  $G_{1\alpha-2}$ ) (21). Modulation of contractile function in the heart via the  $\beta$ -adrenergic pathway involves the interaction of  $\beta_1$ AR and  $\beta_2$ AR with stimulatory guanylyl nucleotide binding proteins, Gs. This leads to the activation of adenylyl cyclase, an increase in cyclic AMP (cAMP), activation of protein kinase A (PKA), and the phosphorylation of direct components of the contractile apparatus and elements of the excitation contraction coupling system.  $\beta_2$ AR is also able to interact with inhibitory G proteins,  $G_i$ , which block adenylyl cyclase function as well as having the potential to activate the pro-survival phosphoinositide 3 kinase (PI3K)/Akt pathway (32-34). The role of the  $\beta$ -adrenergic pathway in the pathogenesis of heart failure has been well studied (32,34,35) and thus the choice to focus on  $\beta_1$ AR,  $\beta_2$ AR, and  $G_{1\alpha-2}$  for further analysis was well conceived.

Chronic adrenergic stimulation in the heart has been shown to elicit cardiotoxicity (36). This effect is thought to be mediated primarily through aberrant activity of  $\beta_1$ AR resulting in calcium overload and cell death (37,38). In failing hearts, there is a down-regulation of  $\beta_1$ AR along with desensitization, whereas the density of  $\beta_2$ AR remains relatively unchanged (39). Transgenic mice overexpressing  $\beta_1$ AR specifically in cardiomyocytes developed fibrosis,

hypertrophy, and reduced fractional shortening at least in part due to increased apoptosis (37). Furthermore, administration of beta-blockers is the hallmark of heart failure treatment and prevention including heart failure from DOX-induced cardiomyopathy (8,40,41). On the other hand,  $\beta_2$ AR is thought to confer cardioprotection (38,42). This pro-survival phenotype is thought to be at least in part associated with its interaction with  $G_i$  proteins (43). In human end stage heart failure patients,  $G_i$  proteins, particularly the  $\alpha$ -2 subunit ( $G_{i\alpha-2}$ ), are up-regulated (44). Down-regulation of  $G_{i\alpha-2}$  is associated with apoptosis and worsening heart failure (45-47). Activation of  $G_{i\alpha-2}$  is slightly more controversial as one group demonstrated that constitutive activation of  $G_{i\alpha-2}$  in a dilated cardiomyopathy and an isoproterenol-induced heart failure mouse model led to worsening hypertrophy and fibrosis, respectively. The authors postulate that the role of  $G_{i\alpha-2}$  in fibroblasts as opposed to cardiomyocytes may be the driving force behind these phenotypes (43). To complicate the story even more, recent models have demonstrated reverse phenotypes for the role of  $\beta_1$ AR and  $\beta_2$ AR in heart failure whereby  $\beta_1$ AR is cardioprotective and  $\beta_2$ AR promotes cardiotoxicity (48). As a result, in addition to cell and disease specific contexts, the balance between  $\beta_1$ AR and  $\beta_2$ AR expression may contribute to the relative role of these receptors in the pathogenesis of heart failure.

### The cardioprotective potential of miR-30

Since miRNAs typically have multiple mRNA targets, these molecules are poised to regulate the delicate balance of multiple effectors in a single influential pathway such as  $\beta$ -adrenergic signaling. Roca-Alonso and colleagues utilized luciferase assays in H9C2 cells to demonstrate direct binding of a miR-30e mimic to the wild-type 3' UTR of four predicted targets ( $\beta_1$ AR,  $\beta_2$ AR,  $G_{i\alpha-2}$ , and the pro-apoptotic protein E1B-interacting protein 3-like or BNIP3L) but not to mutant constructs. In addition, these miR-30e mimics were sufficient to attenuate the up-regulation of these targets upon administration of DOX (21). Appropriate controls with random sequence molecules were used throughout their experiments. These results support the author's claim that DOX-induced repression of miR-30 is at least partially responsible for increased expression of  $\beta_1$ AR,  $\beta_2$ AR,  $G_{i\alpha-2}$ , and BNIP3L.

In order to demonstrate that miR-30 is able to augment the downstream effects of the  $\beta$ -adrenergic pathway, Roca-Alonso and colleagues showed that DOX treatment or

administration of a miR-30 sponge vector led to an increase in cAMP, although the magnitude of up-regulation was more prominent in DOX treated cells. This could be due to miR-30 independent mechanisms of cAMP signaling or incomplete miR-30 targeting, which is consistent with the fact that the authors demonstrate close to a 50% knockdown of miR-30e with sponge vectors. Overexpression of miR-30, on the other hand, resulted in decreased cAMP levels (21).

The phenotypic response to miR-30 was assayed through its effects on contractile function and DOX toxicity. Cells overexpressing miR-30 had an attenuation of contractile amplitude in response to increasing concentrations of isoproterenol compared to control cells. With regards to DOX toxicity, over-expression of miR-30 attenuated the increase in caspase activity triggered by DOX, although this attenuation was also incomplete. In addition, down-regulation of miR-30 led to an increased level of caspase activity compared to controls. Furthermore, in the presence of DOX, the intensity of ROS in cardiac cells was decreased with miR-30 overexpression and increased with miR-30 inhibition compared to cells exposed to DOX alone (21). While these findings are intriguing, they do not provide a direct link between miR-30 activity and damage through the  $\beta$ -adrenergic pathway. Future studies manipulating targets of miR-30 will be necessary to draw such conclusions.

Finally, Roca-Alonso and colleagues also uncovered the presence of GATA-6 binding to miR-30 cluster promoters in publicly available data from chromatin immunoprecipitation sequencing experiments. They hypothesized that GATA-6 binding to these regions following DOX may mediate down-regulation of miR-30 (21). GATA-6 has already been implicated in cardiac pathology where it seems to be required to mount the cardiac hypertrophic response and prevent heart failure in animal models (49). To substantiate their hypothesis, these authors demonstrated that *GATA-6* is up-regulated acutely following exposure of DOX to cardiac cells in culture and siRNA constructs against *GATA-6* resulted in increased expression of miR-30 family members and decreased expression of miR-30 targets. Even more, in the setting of DOX treatment, knockdown of *GATA-6* resulted in decreased signaling through the apoptotic pathways (21). Nevertheless, experiments demonstrating the attenuation of miR-30 down-regulation after DOX in the absence of *GATA-6* are lacking. As a result, other transcription factors could be more important than *GATA-6* on the effect of DOX on miR-30. These experiments also beg the question of how *GATA-6* is activated following DOX treatment and whether additional targets of *GATA-6* could contribute to

the damage response. Moreover, if GATA-6 does indeed prove cardioprotective against failing hearts as demonstrated by others, its negative regulation of miR-30 would appear to promote cardiotoxicity. Further studies are needed to untangle the intricacies of these networks.

Roca-Alonso and colleagues provide an interesting model whereby down-regulation of  $\beta_1$ AR,  $\beta_2$ AR, and  $G_{1\alpha-2}$  leads to a cardioprotective phenotype. While this certainly aligns with studies that demonstrate cardiotoxicity of excessive  $\beta_1$ AR, it appears to contradict the cardioprotective role of  $\beta_2$ AR and  $G_{1\alpha-2}$ . However, the authors offer an explanation of this apparent contradiction by arguing that the fine tuning of expression characteristic of miRNAs can regulate the feedback loop involving  $\beta_1$ AR,  $\beta_2$ AR, and  $G_{1\alpha-2}$  in such a way to prevent the cardiotoxicity of  $\beta_1$ AR signaling, while still maintaining some of the cardioprotective benefits of  $\beta_2$ AR and  $G_{1\alpha-2}$ . In a similar fashion, the beta-blocker Carvedilol has been shown in patients and animal models to be protective against DOX-induced cardiomyopathy despite being characterized as nonselective towards  $\beta_1$ AR and  $\beta_2$ AR (40,41). Most of the experiments for these conclusions were based on *in vitro* cell cultures. Future studies *in vivo* are necessary to determine the long-term phenotypic consequences of miR-30 knockdown and overexpression on DOX-induced cardiomyopathy in order to fully test the therapeutic potential of these miRNAs. Such models can also be used to determine potential additive or synergistic benefit of miR-30 overexpression with beta-blockade.

## Conclusions

In a recent report by Roca-Alonso and colleagues, a novel mechanism for DOX-induced injury involving the regulation of the  $\beta$ -adrenergic pathway through miRNAs is uncovered. These findings provide new insight into the complex pathways that govern the damage response and the delicate balance between the expression of key elements in a single pathway. Even more, these studies offer exciting potential for future therapeutic targets of chemotherapy-induced cardiomyopathy.

## Acknowledgements

**Funding:** This work was supported by NIH R01HL118266 to JDM.

## Footnote

**Conflicts of Interest:** The authors have no conflicts of interest to declare.

## References

1. Volkova M, Russell R 3rd. Anthracycline cardiotoxicity: prevalence, pathogenesis and treatment. *Curr Cardiol Rev* 2011;7:214-220.
2. Raj S, Franco VI, Lipshultz SE. Anthracycline-induced cardiotoxicity: a review of pathophysiology, diagnosis, and treatment. *Curr Treat Options Cardiovasc Med* 2014;16:315.
3. Salazar-Mendiguchía J, González-Costello J, Roca J, et al. Anthracycline-mediated cardiomyopathy: basic molecular knowledge for the cardiologist. *Arch Cardiol Mex* 2014;84:218-223.
4. Swain SM, Whaley FS, Ewer MS. Congestive heart failure in patients treated with doxorubicin: a retrospective analysis of three trials. *Cancer* 2003;97:2869-2879.
5. Mulrooney DA, Yeazel MW, Kawashima T, et al. Cardiac outcomes in a cohort of adult survivors of childhood and adolescent cancer: retrospective analysis of the Childhood Cancer Survivor Study cohort. *BMJ* 2009;339:b4606.
6. Zhang S, Liu X, Bawa-Khalfe T, et al. Identification of the molecular basis of doxorubicin-induced cardiotoxicity. *Nat Med* 2012;18:1639-1642.
7. Ghigo A, Li M, Hirsch E. New signal transduction paradigms in anthracycline-induced cardiotoxicity. *Biochim Biophys Acta* 2016. [Epub ahead of print].
8. van Dalen EC, Caron HN, Dickinson HO, et al. Cardioprotective interventions for cancer patients receiving anthracyclines. *Cochrane Database Syst Rev* 2011.CD003917.
9. Vasti C, Hertig CM. Neuregulin-1/erbB activities with focus on the susceptibility of the heart to anthracyclines. *World J Cardiol* 2014;6:653-662.
10. Horie T, Ono K, Nishi H, et al. Acute doxorubicin cardiotoxicity is associated with miR-146a-induced inhibition of the neuregulin-ErbB pathway. *Cardiovasc Res* 2010;87:656-664.
11. Nozaki N, Shishido T, Takeishi Y, et al. Modulation of doxorubicin-induced cardiac dysfunction in toll-like receptor-2-knockout mice. *Circulation* 2004;110:2869-2874.
12. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281-297.

13. Wang Y, Pan X, Fan Y, et al. Dysregulated expression of microRNAs and mRNAs in myocardial infarction. *Am J Transl Res* 2015;7:2291-2304.
14. Boon RA, Dimmeler S. MicroRNAs in myocardial infarction. *Nat Rev Cardiol* 2015;12:135-142.
15. Saddic LA, Chang TW, Sigurdsson MI, et al. Integrated microRNA and mRNA responses to acute human left ventricular ischemia. *Physiol Genomics* 2015;47:455-462.
16. Gurha P. MicroRNAs in cardiovascular disease. *Curr Opin Cardiol* 2016;31:249-254.
17. Wong LL, Wang J, Liew OW, et al. MicroRNA and Heart Failure. *Int J Mol Sci* 2016.17.
18. Greco S, Gorospe M, Martelli F. Noncoding RNA in age-related cardiovascular diseases. *J Mol Cell Cardiol* 2015;83:142-155.
19. Holmgren G, Synnergren J, Andersson CX, et al. MicroRNAs as potential biomarkers for doxorubicin-induced cardiotoxicity. *Toxicol In Vitro* 2016;34:26-34.
20. Yin Z, Zhao Y, Li H, et al. miR-320a mediates doxorubicin-induced cardiotoxicity by targeting VEGF signal pathway. *Aging (Albany NY)* 2016;8:192-207.
21. Roca-Alonso L, Castellano L, Mills A, et al. Myocardial MiR-30 downregulation triggered by doxorubicin drives alterations in  $\beta$ -adrenergic signaling and enhances apoptosis. *Cell Death Dis* 2015;6:e1754.
22. Tong Z, Jiang B, Wu Y, et al. MiR-21 Protected Cardiomyocytes against Doxorubicin-Induced Apoptosis by Targeting BTG2. *Int J Mol Sci* 2015;16:14511-14525.
23. Tony H, Yu K, Qiutang Z. MicroRNA-208a Silencing Attenuates Doxorubicin Induced Myocyte Apoptosis and Cardiac Dysfunction. *Oxid Med Cell Longev* 2015;2015:597032.
24. Li T, Sun ZL, Xie QY. Protective effect of microRNA-30b on hypoxia/reoxygenation-induced apoptosis in H9C2 cardiomyocytes. *Gene* 2015;561:268-275.
25. Vacchi-Suzzi C, Bauer Y, Berridge BR, et al. Perturbation of microRNAs in rat heart during chronic doxorubicin treatment. *PLoS One* 2012;7:e40395.
26. Desai VG, C, Kwekel J, Vijay V, et al. Early biomarkers of doxorubicin-induced heart injury in a mouse model. *Toxicol Appl Pharmacol* 2014;281:221-229.
27. Che S, Sun T, Wang J, et al. miR-30 overexpression promotes glioma stem cells by regulating Jak/STAT3 signaling pathway. *Tumour Biol* 2015;36:6805-6811.
28. Kao CJ, Martiniez A, Shi XB, et al. miR-30 as a tumor suppressor connects EGF/Src signal to ERG and EMT. *Oncogene* 2014;33:2495-2503.
29. Zhong K, Chen K, Han L, Li B. MicroRNA-30b/c inhibits non-small cell lung cancer cell proliferation by targeting Rab18. *BMC Cancer* 2014;14:703.
30. Duisters RF, Tijssen AJ, Schroen B, et al. miR-133 and miR-30 regulate connective tissue growth factor: implications for a role of microRNAs in myocardial matrix remodeling. *Circ Res* 2009;104:170-8, 6p following 178.
31. Pan W, Zhong Y, Cheng C, et al. MiR-30-regulated autophagy mediates angiotensin II-induced myocardial hypertrophy. *PLoS One* 2013;8:e53950.
32. Najafi A, Sequeira V, Kuster DW, et al.  $\beta$ -adrenergic receptor signalling and its functional consequences in the diseased heart. *Eur J Clin Invest* 2016;46:362-374.
33. Xiao RP. Beta-adrenergic signaling in the heart: dual coupling of the beta2-adrenergic receptor to G(s) and G(i) proteins. *Sci STKE* 2001;2001:re15.
34. Yang J, Liu Y, Fan X, et al. A pathway and network review on beta-adrenoceptor signaling and beta blockers in cardiac remodeling. *Heart Fail Rev* 2014;19:799-814.
35. Santulli G, Iaccarino G. Adrenergic signaling in heart failure and cardiovascular aging. *Maturitas* 2016. [Epub ahead of print].
36. Communal C, Singh K, Pimentel DR, et al. Norepinephrine stimulates apoptosis in adult rat ventricular myocytes by activation of the beta-adrenergic pathway. *Circulation* 1998;98:1329-1334.
37. Bisognano JD, Weinberger HD, Bohlmeyer TJ, et al. Myocardial-directed overexpression of the human beta(1)-adrenergic receptor in transgenic mice. *J Mol Cell Cardiol* 2000;32:817-830.
38. Baker AJ. Adrenergic signaling in heart failure: a balance of toxic and protective effects. *Pflugers Arch* 2014;466:1139-1150.
39. Bristow MR, Ginsburg R, Umans V, et al. Beta 1- and beta 2-adrenergic-receptor subpopulations in nonfailing and failing human ventricular myocardium: coupling of both receptor subtypes to muscle contraction and selective beta 1-receptor down-regulation in heart failure. *Circ Res* 1986;59:297-309.
40. Tashakori Beheshti A, Mostafavi Toroghi H, Hosseini G, et al. Carvedilol Administration Can Prevent Doxorubicin-Induced Cardiotoxicity: A Double-Blind Randomized Trial. *Cardiology* 2016;134:47-53.
41. Kalay N, Basar E, Ozdogru I, et al. Protective effects of carvedilol against anthracycline-induced cardiomyopathy. *J Am Coll Cardiol* 2006;48:2258-2262.
42. Patterson AJ, Zhu W, Chow A, et al. Protecting the

- myocardium: a role for the beta2 adrenergic receptor in the heart. *Crit Care Med* 2004;32:1041-1048.
43. Kaur K, Parra S, Chen R, et al. G $\alpha$ i2 signaling: friend or foe in cardiac injury and heart failure? *Naunyn Schmiedebergs Arch Pharmacol* 2012;385:443-453.
  44. Eschenhagen T, Mende U, Nose M, et al. Increased messenger RNA level of the inhibitory G protein alpha subunit Gi alpha-2 in human end-stage heart failure. *Circ Res* 1992;70:688-696.
  45. DeGeorge BR Jr, Gao E, Boucher M, et al. Targeted inhibition of cardiomyocyte Gi signaling enhances susceptibility to apoptotic cell death in response to ischemic stress. *Circulation* 2008;117:1378-1387.
  46. Foerster K, Groner F, Matthes J, et al. Cardioprotection specific for the G protein Gi2 in chronic adrenergic signaling through beta 2-adrenoceptors. *Proc Natl Acad Sci U S A* 2003;100:14475-14480.
  47. Chesley A, Lundberg MS, Asai T, et al. The beta(2)-adrenergic receptor delivers an antiapoptotic signal to cardiac myocytes through G(i)-dependent coupling to phosphatidylinositol 3'-kinase. *Circ Res* 2000;87:1172-1179.
  48. Fajardo G, Zhao M, Urashima T, et al. Deletion of the  $\beta$ 2-adrenergic receptor prevents the development of cardiomyopathy in mice. *J Mol Cell Cardiol* 2013;63:155-164.
  49. van Berlo JH, Aronow BJ, Molkentin JD. Parsing the roles of the transcription factors GATA-4 and GATA-6 in the adult cardiac hypertrophic response. *PLoS One* 2013;8:e84591.

**Cite this article as:** Saddic LA, Muehlschlegel JD. Sarco“MiR” friend or foe: a perspective on the mechanisms of doxorubicin-induced cardiomyopathy. *Ann Transl Med* 2016;4(10):203. doi: 10.21037/atm.2016.05.30

# Molecular evidence that exercise training has beneficial effects on cardiac performance

Marek Kiliszek<sup>1</sup>, Urszula Mackiewicz<sup>2</sup>, Michal Maczewski<sup>2</sup>, Beata Burzynska<sup>3</sup>

<sup>1</sup>Department of Cardiology and Internal Diseases, Military Institute of Medicine, Warsaw, Poland; <sup>2</sup>Department of Clinical Physiology, Medical Centre of Postgraduate Education, Warsaw, Poland; <sup>3</sup>Institute of Biochemistry and Biophysics, Polish Academy of Sciences, Warsaw, Poland

Correspondence to: Beata Burzynska. Institute of Biochemistry and Biophysics, Polish Academy of Sciences, Pawinskiego 5A, 02-106 Warsaw, Poland. Email: atka@ibb.waw.pl.

Provenance: This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

Comment on: Melo SF, Barauna VG, Neves VJ, *et al.* Exercise training restores the cardiac microRNA-1 and -214 levels regulating Ca<sup>2+</sup> handling after myocardial infarction. *BMC Cardiovasc Disord* 2015;15:166.

Submitted May 12, 2016. Accepted for publication May 18, 2016.

doi: 10.21037/atm.2016.05.53

View this article at: <http://dx.doi.org/10.21037/atm.2016.05.53>

Numerous epidemiological and observational studies demonstrate that there is an inverse relationship between physical activity and risk of cardiovascular disease (1). Recently a few randomized controlled trials revealed that exercise training is not only effective as primary prevention, but also in the secondary prevention and thus can be viewed as a “medication” that should be taken on a regular basis by a patient with cardiovascular disease. Exercise in Left Ventricular Dysfunction (ELVD) trial in a small group of 77 patients with <40% ejection fraction after a first Q-wave myocardial infarction showed that a 6-month exercise training program prevented deleterious LV remodeling (2). The Heart Failure: A Controlled Trial Investigating Outcomes of Exercise Training (HF-ACTION) trial, enrolling 2,331 outpatients with stable systolic heart failure demonstrated that exercise training was associated with an 11% reduction in combined all-cause death or hospitalization (P=0.03) (3). A meta-analysis of exercise training trials in patients with chronic heart failure, majority of whom had a history of myocardial infarction, revealed that exercise training significantly reduced deaths and hospital admissions (4). Thus exercise training is recommended as part of a comprehensive approach to the patient with stable chronic heart failure (1).

Presumable mechanisms of this beneficial effect of exercise in the post-myocardial infarction and chronic heart failure setting include: improvement of VO<sub>2</sub>max, reduction

of neurohormonal imbalance, antiarrhythmic effects, resolution of ventilatory abnormalities, improvement of endothelial function, improved both systolic and diastolic myocardial performance through improvement of cardiomyocyte contraction-relaxation cycle.

Cardiomyocyte contractile function has been shown to be impaired in post-MI heart failure. Decreased amplitude of myocyte contraction as well as slower kinetics of contraction-relaxation cycle has been demonstrated in many experimental models of post-MI heart failure and in humans (5,6). Cardiomyocyte contractile function is strictly controlled by beat-to-beat transient increase of intracellular Ca<sup>2+</sup> concentration (i.e., calcium transient). After electrical activation, rising of the membrane potential opens the voltage-gated sarcolemmal L-type Ca<sup>2+</sup> channels. This results in influx of small amount of Ca<sup>2+</sup> to the myocyte, which activates the calcium-dependent sarcoplasmic reticulum (SR) Ca<sup>2+</sup> release channels [ryanodine receptors (RyRs)]. This process is commonly called calcium-induced calcium release. Rapid release of considerable amount of SR Ca<sup>2+</sup> results in increase of intracellular Ca<sup>2+</sup> concentration and promotes Ca<sup>2+</sup> binding to troponin C, a contractile apparatus regulatory protein. The change of troponin C conformation upon Ca<sup>2+</sup> binding enables actin-myosin interaction and thus myocyte contraction. Relaxation is initiated by termination of the Ca<sup>2+</sup> release from the SR and by rapid Ca<sup>2+</sup> removal from the cytosol. Two main



transporting proteins are involved in this process: SR  $\text{Ca}^{2+}$ -ATP-ase (SERCA) which uses ATP to pump calcium back into the SR and the  $\text{Na}^+/\text{Ca}^{2+}$  exchanger (NCX) which transports 1  $\text{Ca}^{2+}$  ion out of the cell and 3  $\text{Na}^+$  ions into the cell. SERCA accounts for approximately 80% of removal of systolic calcium in humans and even more (about 90%) in rodents. Thus, the transporting function of SERCA is the main determinant of the rate of cardiomyocyte relaxation. Additionally the SERCA transporting ability determines the SR  $\text{Ca}^{2+}$  content and thus amplitude of  $\text{Ca}^{2+}$  transient and amplitude of myocyte contraction. The transporting function of SERCA depends on its expression, intrinsic activity of enzyme (ability to utilize ATP) and the phosphorylation level of phospholamban, endogenous SERCA inhibitor. Phospholamban is phosphorylated at Ser-16 and Thr-17 by adrenergic stimulation dependent kinase (PKA) and calmodulin and  $\text{Ca}^{2+}$  dependent kinase (CAMKII), respectively and increase of phosphorylation level relieves SERCA inhibition (7).

In post-MI heart failure detrimental changes in  $\text{Ca}^{2+}$  handling have been described. Decreased SERCA transporting function has been demonstrated consistently, due to both decreases of SERCA expression as well as decreased transporting ability, mainly due to reduced level of phospholamban phosphorylation. Moreover, in many animal models and in humans increased NCX expression or/and function has also been described. These changes are additionally accompanied by an increased  $\text{Ca}^{2+}$  sensitivity of RyRs due to their hyperphosphorylation which results in  $\text{Ca}^{2+}$ -leak from the SR independently from  $\text{Ca}^{2+}$  influx through the L-type  $\text{Ca}^{2+}$  channels (diastolic  $\text{Ca}^{2+}$ -leak) (8,9).

Decreased SERCA expression accompanied by increased NCX function results in increased proportion of intracellular  $\text{Ca}^{2+}$  removed from the cytoplasm by NCX as compared with SERCA. This together with increased  $\text{Ca}^{2+}$ -leak results in decreased  $\text{Ca}^{2+}$  SR content, amplitude of  $\text{Ca}^{2+}$  transient and myocyte shortening as well as the decreased rate of  $\text{Ca}^{2+}$  transient decay and slower relaxation. Moreover, increased NCX contribution to the relaxation increased inward current (1  $\text{Ca}^{2+}$  ion is exchanged with 3  $\text{Na}^+$  ions) and may promote afterdepolarization, premature beats and increased susceptibility to ventricular arrhythmias. Indeed the post-MI animals as well as patients with ischemic heart failure die from progressive pump failure or sudden arrhythmic events (10).

Many elegant papers have shown that regular, intensive aerobic exercise training influences  $\text{Ca}^{2+}$  handling and thus myocyte contraction and relaxation process in

cardiomyocytes from both healthy and post-MI hearts (11).

In healthy animals exercise training resulted in approximately 30% increase of transporting activity of SERCA measured in the intact SR membranes or permeabilized cardiomyocytes (12). It was due to increased SERCA expression at mRNA and protein level as well as increased phospholamban phosphorylation. Additionally in some studies increased level of NCX and increased sensitivity of contractile apparatus were observed. Consequently, amplitude of the cell shortening and the rate of relaxation were increased (13).

In post-MI heart failure, exercise training seems to be especially beneficial. The restoration of the normal amplitude and rates of contraction and relaxation has been observed. It was associated with normalization of the expression of SERCA and NCX proteins. It supports the cardiomyocyte function and decrease propensity to  $\text{Ca}^{2+}$ -dependent ventricular arrhythmias (14). There is growing body of evidence indicating that exercise training is able to restore of the proper expression of the protein involved in  $\text{Ca}^{2+}$  handling in failing hearts the mechanisms of this restoration is still poorly understood.

The discovery of microRNAs (miRNAs), abundant single-stranded small (roughly 22 nucleotide long) nonprotein-coding RNAs, has made important contribution to the better understanding of mechanisms that regulate of gene expression. MicroRNAs have been shown to be involved in most biological processes, both physiological and pathophysiological, including cardiovascular diseases. MicroRNAs are transcribed as individual or in clusters, often as part of longer transcripts, and are expressed in a tissue and cell-specific manner. The miRNA system is generally regarded as a negative regulator of specific mRNA targets. They can inhibit translation and/or promote mRNA degradation by sequence-specific base pairing (15).

Many well documented studies revealed that microRNAs were frequently downregulated in various types of cardiac diseases, including pathogenesis of MI (16). Because miRNAs are important in many cardiac pathologies, they may play a functional role in exercise-induced cardiac phenotypes.

Melo *et al.* in their work titled “*Exercise training restores the cardiac microRNA-1 and -214 levels regulating  $\text{Ca}^{2+}$  handling after myocardial infarction*”, published in *BMC Cardiovascular Disorders* (17) demonstrated that myocardial infarction in the rat resulted in reduced expression of SERCA and increased expression of NCX. Expression of microRNA-214 that targets SERCA, was increased, while

that of microRNA-1 that targets NCX, was reduced. Ten weeks of exercise training resulted in restoration of both microRNA levels and prevents changes of expression of both calcium transporters induced by myocardial infarction. These results suggest that changes in microRNA are responsible for restoration of SERCA and NCX expression, through this conclusion is only based on the above mentioned correlations.

This article provides new data on possible mechanisms behind effects of exercise training on cardiac performance in infarcted heart. microRNA-1 is cardiac specific miRNA and plays a role in heart hypertrophy, myocardial infarction, and arrhythmias, by promoting apoptosis. Recently, studies have revealed that miR-1 was frequently downregulated in various types of cardiac disease but when overexpressed, played a protective role against cardiac hypertrophy or heart failure by regulating several hypertrophy-associated genes, including transcription factors, receptor ligands, apoptosis regulators and ion channels (18). Overexpression of microRNA-1 and other miRs is implicated in regulation of G-PCR and calcium handling (19). Second miRNA investigated by authors, microRNA-214, improved LV remodeling and decreased apoptosis of myocardial cell and had a protective effect on heart function (20).

In summary, the study by Melo *et al.* highlights new potential mechanisms of beneficial effects of exercise on the post-MI heart, providing new areas for future research.

### Acknowledgements

*Funding:* Research on role of mRNAs and miRNAs in heart failure conducted in our Lab is supported by The National Centre for Research and Development, Poland (TANGO1/266456/NCBR/2015).

### Footnote

*Conflict of Interest:* The authors have no conflict of interest to declare.

### References

1. Fletcher GF, Ades PA, Kligfield P, et al. Exercise standards for testing and training: a scientific statement from the American Heart Association. *Circulation* 2013;128:873-934.
2. Giannuzzi P, Temporelli PL, Corrà U, et al. Attenuation of unfavorable remodeling by exercise training in postinfarction patients with left ventricular dysfunction: results of the Exercise in Left Ventricular Dysfunction (ELVD) trial. *Circulation* 1997;96:1790-1797.
3. O'Connor CM, Whellan DJ, Lee KL, et al. Efficacy and safety of exercise training in patients with chronic heart failure: HF-ACTION randomized controlled trial. *JAMA* 2009;301:1439-1450.
4. Piepoli MF, Davos C, Francis DP, et al. Exercise training meta-analysis of trials in patients with chronic heart failure (ExTraMATCH). *BMJ* 2004;328:189.
5. Hasenfuss G, Pieske B. Calcium cycling in congestive heart failure. *J Mol Cell Cardiol* 2002;34:951-969.
6. Yano M, Ikeda Y, Matsuzaki M. Altered intracellular Ca<sup>2+</sup> handling in heart failure. *J Clin Invest* 2005;115:556-564.
7. Leszek P, Szperl M, Klisiewicz A, et al. Alteration of myocardial sarcoplasmic reticulum Ca<sup>2+</sup>-ATPase and Na<sup>+</sup>-Ca<sup>2+</sup> exchanger expression in human left ventricular volume overload. *Eur J Heart Fail* 2007;9:579-586.
8. Marx SO, Marks AR. Dysfunctional ryanodine receptors in the heart: new insights into complex cardiovascular diseases. *J Mol Cell Cardiol* 2013;58:225-231.
9. Mackiewicz U, Maczewski M, Konior A, et al. Sarcolemmal Ca<sup>2+</sup>-ATPase ability to transport Ca<sup>2+</sup> gradually diminishes after myocardial infarction in the rat. *Cardiovasc Res* 2009;81:546-554.
10. Myles RC, Burton FL, Cobbe SM, et al. The link between repolarisation alternans and ventricular arrhythmia: does the cellular phenomenon extend to the clinical problem? *J Mol Cell Cardiol* 2008;45:1-10.
11. Kemi OJ, Wisløff U. Mechanisms of exercise-induced improvements in the contractile apparatus of the mammalian myocardium. *Acta Physiol (Oxf)* 2010;199:425-439.
12. Kemi OJ, Ceci M, Condorelli G, et al. Myocardial sarcoplasmic reticulum Ca<sup>2+</sup> ATPase function is increased by aerobic interval training. *Eur J Cardiovasc Prev Rehabil* 2008;15:145-148.
13. Wisløff U, Loennechen JP, Currie S, et al. Aerobic exercise reduces cardiomyocyte hypertrophy and increases contractility, Ca<sup>2+</sup> sensitivity and SERCA-2 in rat after myocardial infarction. *Cardiovasc Res* 2002;54:162-174.
14. Venetucci LA, Trafford AW, O'Neill SC, et al. The sarcoplasmic reticulum and arrhythmogenic calcium release. *Cardiovasc Res* 2008;77:285-292.
15. Small EM, Olson EN. Pervasive roles of microRNAs in cardiovascular biology. *Nature* 2011;469:336-342.
16. Chistiakov DA, Orekhov AN, Bobryshev YV. Cardiac-specific miRNA in cardiogenesis, heart function, and cardiac pathology (with focus on myocardial infarction). *J*



- Mol Cell Cardiol 2016;94:107-121.
17. Melo SF, Barauna VG, Neves VJ, et al. Exercise training restores the cardiac microRNA-1 and -214 levels regulating Ca<sup>2+</sup> handling after myocardial infarction. *BMC Cardiovasc Disord* 2015;15:166.
  18. Sygitowicz G, Tomaniak M, Błaszczuk O, et al. Circulating microribonucleic acids miR-1, miR-21 and miR-208a in patients with symptomatic heart failure: Preliminary results. *Arch Cardiovasc Dis* 2015;108:634-642.
  19. Yang X, Qin Y, Shao S, et al. MicroRNA-214 Inhibits Left Ventricular Remodeling in an Acute Myocardial Infarction Rat Model by Suppressing Cellular Apoptosis via the Phosphatase and Tensin Homolog (PTEN). *Int Heart J* 2016;57:247-250.
  20. Fernandes T, Baraúna VG, Negrão CE, et al. Aerobic exercise training promotes physiological cardiac remodeling involving a set of microRNAs. *Am J Physiol Heart Circ Physiol* 2015;309:H543-H552.

**Cite this article as:** Kiliszek M, Mackiewicz U, Maczewski M, Burzynska B. Molecular evidence that exercise training has beneficial effects on cardiac performance. *Ann Transl Med* 2016;4(11):228. doi: 10.21037/atm.2016.05.53

## How to be young at heart? miR-22 as a potential therapeutic target to boost autophagy and protect the old myocardium

Sebastiano Sciarretta<sup>1,2</sup>, Elena De Falco<sup>1</sup>, Giacomo Frati<sup>1,2</sup>, Junichi Sadoshima<sup>3</sup>

<sup>1</sup>Department of Medico-Surgical Sciences and Biotechnologies, Sapienza University of Rome, Rome, Italy; <sup>2</sup>Department of AngioCardioNeurology, IRCCS Neuromed, Pozzilli, Italy; <sup>3</sup>Department of Cell Biology and Molecular Medicine, Rutgers New Jersey Medical School, Newark, NJ, USA

*Correspondence to:* Junichi Sadoshima, MD, PhD. Department of Cell Biology and Molecular Medicine, Rutgers New Jersey Medical School, 185 South Orange Avenue, MSB-609, Newark, NJ, USA. Email: sadoshju@njms.rutgers.edu.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Gupta SK, Foinquinos A, Thum S, *et al.* Preclinical Development of a MicroRNA-Based Therapy for Elderly Patients With Myocardial Infarction. *J Am Coll Cardiol* 2016;68:1557-71.

Submitted Nov 29, 2016. Accepted for publication Dec 05, 2016.

doi: 10.21037/atm.2017.01.52

**View this article at:** <http://dx.doi.org/10.21037/atm.2017.01.52>

Cardiovascular diseases remain the most common cause of death in Western countries and their prevalence in the general population increases progressively with age, reaching more than 60% in subjects older than 60 years (1). This is at least in part due to a progressive impairment of the cellular processes regulating cardiac and vascular homeostasis, finally leading to the development of cardiovascular abnormalities (2). In addition, the molecular mechanisms that protect the heart against stress are downregulated by aging, making the myocardium more susceptible to injury (2). It is therefore important to find new therapeutic targets to reduce cardiovascular aging and protect the aged myocardium from stress.

Autophagy is a mechanism of intracellular degradation through lysosomes characterized by the presence of double membrane vesicles called autophagosomes (2,3). Increasing lines of evidence suggest that autophagy plays an important role in maintaining cardiac homeostasis. Autophagy is downregulated in the heart during aging, and downregulation of autophagy plays a causative role in the progression of aging in the heart (2,4). Furthermore, we and others have shown that downregulation of autophagy during pressure overload (5,6), post-myocardial infarction cardiac remodeling (7) and metabolic syndrome (8,9) contributes to the progression of cardiac dysfunction and ischemic injury. On the other hand, excessive activation of autophagy during the acute phase of myocardial infarction

or ischemia/reperfusion may be detrimental (10). Thus, identifying molecular interventions to control the activity of autophagy is important and may lead to the development of a novel treatment for heart disease, especially in elderly patients.

Using high-throughput screening based on the fluorescence-activated cell sorting-based detection of GFP-LC3 protein and library screening, Gupta *et al.* identified miR-22 as a strong inhibitor of autophagy in cardiomyocytes (11). The authors showed that miR-22 inhibits the reduction in GFP-LC3 signal in response to starvation, an established stimulus for autophagy. This screening method is based on the fact that the GFP-LC3 signal is attenuated when GFP-LC3 autophagosomes move from the cytosol to lysosomes (12). Thus, it is a convenient and reliable method to evaluate the level of autophagic flux from autophagosome formation to autolysosome formation. A cautionary note, however, is that the results of this method are easily affected by conditions that alter the intensity of the GFP signal through autophagy-independent mechanisms. Thus, the authors could have conducted more extensive validation regarding how miR-22 affects autophagic flux in cardiomyocytes throughout the study. Nevertheless, this is a fascinating way to identify effective modifiers of autophagy in cardiomyocytes and, by extending this approach to small molecules, it may be possible to identify novel interventions for heart failure patients.

Gupta *et al.* demonstrated that miR-22 expression progressively increases in the mouse heart during aging (11). miR-22 inhibition by locked nucleic acid-based anti-miR-22 (LNA-anti-miR-22) significantly attenuated cardiac remodeling in old mice subjected to permanent coronary artery ligation, reducing left ventricular dilation and improving systolic function. In contrast, the beneficial effects of miR-22 inhibition were not obvious in young mice with chronic myocardial infarction. Thus, miR-22 upregulation in the aged heart appears to impair adaptation to chronic myocardial infarction. miR-22 inhibition limits senescence-induced cardiomyocyte hypertrophy *in vitro*. Therefore, the cardioprotective effects of pharmacological miR-22 inhibition in the mouse heart may be dependent on its protective actions in cardiomyocytes. However, since miR-22 also induces senescence in cardiac fibroblasts, increasing their migratory capacity (13), it is possible that miR-22 inhibition may also elicit beneficial functions through the other cell types in the heart.

miR-22 appeared to be responsible for the reduction of autophagy in the old myocardium. The increase in cardiac miR-22 levels during aging was paralleled by a progressive decline in myocardial autophagy. Importantly, miR-22 inhibition appears to rescue autophagy in aging hearts (11). miR-22 may have autophagy-independent actions, and thus, the causative involvement of autophagy in mediating the salutary actions of miR-22 inhibition remains to be demonstrated. Nevertheless, the fact that miR-22 inhibition can stimulate autophagy in the aging heart is attractive. Restoring the level of autophagy has been shown to be salutary in some cardiovascular conditions. For example, genetic inhibition of the serine-threonine kinase MST1, a stress activated kinase known to inhibit autophagy, reduces remodeling caused by chronic ischemia through the activation of autophagy (7). In addition, administration of spermidine, a natural polyamine compound, extended life span in mice and reduced cardiac hypertrophy and diastolic dysfunction induced by aging. These effects were found to be dependent on the capacity of spermidine to activate autophagy through the inhibition of the histone acetyltransferase p300. In fact, the cardioprotective effects of spermidine were lost in mice with genetic disruption of autophagy (4).

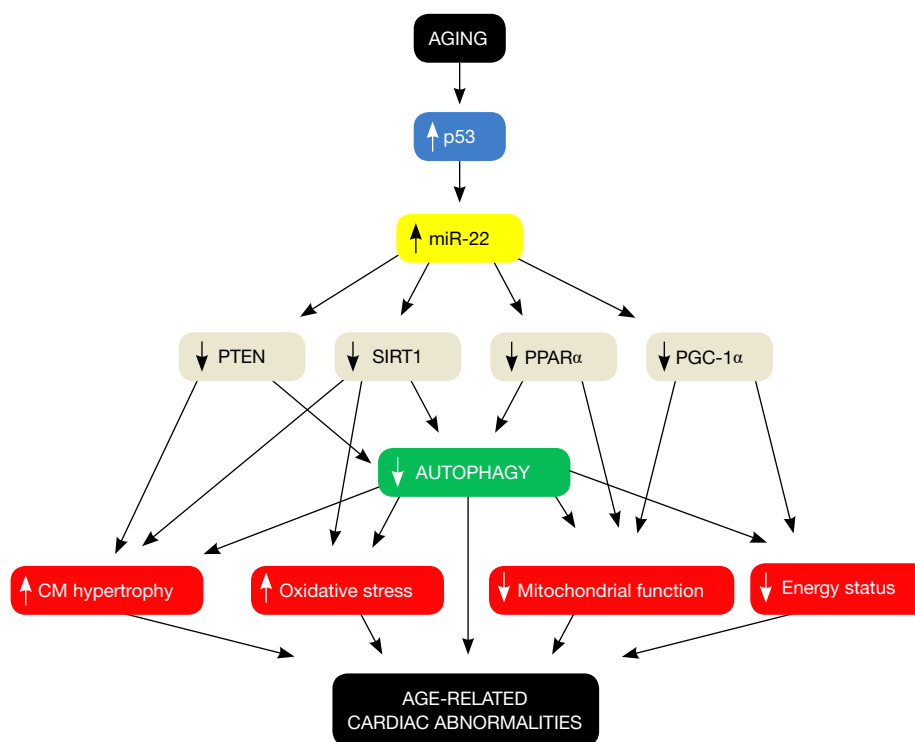
Other miRs also regulate autophagy in the heart. Thum's group elegantly showed that miRNA-212/132 inhibits autophagy in cardiomyocytes by targeting FoxO3a. Pharmacological inhibition of miR-132 reduced

cardiac hypertrophy and heart failure induced by pressure overload (14). In addition, exosome-mediated miR-145 administration was found to reduce ischemic injury by activating autophagy (15). Thus, targeting these miRNAs could represent a new avenue of exploration in the development of novel interventions to treat heart failure in elderly patients.

miR-22a plays a dual role in the heart, performing both physiological and maladaptive functions. Mice with either systemic or cardiac-specific miR-22 gene deletion do not develop compensatory hypertrophy in response to isoproterenol treatment and display increased cardiac dilation and dysfunction with respect to control mice (16). Similarly, systemic miR-22 knockout mice were found to develop cardiac dilation and dysfunction in response to pressure overload (17). These effects were associated with a downregulation of Serca2a expression and with sarcomere disarray. Interestingly, however, miR-22 gain of function also appears to be maladaptive. Mice with cardiac-specific miR-22 overexpression develop cardiac hypertrophy and dysfunction, and these deleterious effects are associated with impaired calcium handling and reduced expression of PPAR $\alpha$ , SIRT1 and PGC-1 $\alpha$  (18). Thus, it is likely that proper cardiac function requires that miR-22 expression levels remain within a relatively narrow physiological range.

The molecular mechanism by which miR-22 inhibits autophagy has not yet been fully elucidated. miR-22 was found to inhibit autophagy in cardiomyocytes by reducing the expression of PPAR $\alpha$  (11). However, other mechanisms may also be involved in the inhibitory effects of miR-22 on autophagy. It was previously demonstrated that miR-22 reduces the expression of PTEN, a negative regulator of the AKT pathway, in cardiomyocytes (19). This suggests that miR-22 may also suppress autophagy through activation of AKT, which may, in turn, activate the kinase mTOR, a negative regulator of the autophagic process (20). Since miR-22 affects cardiac function through multiple mechanisms, it is also possible that the inhibition of autophagy is indirectly mediated through other functional targets of miR-22, such as the mitochondrial dysfunction and metabolic remodeling observed in hearts following myocardial infarction. Future studies are warranted to test this possibility.

Aside from autophagy, other mechanisms may also underlie the beneficial cardiac effects of pharmacological miR-22 inhibition (*Figure 1*). Inhibition of the AKT/mTOR pathway may be one of these mechanisms, since



**Figure 1** miR-22 promotes age-related cardiac abnormalities. Schematic representation of the molecular mechanisms through which miR-22 promotes age-related cardiac abnormalities. Autophagy inhibition appears to play a major role in the development of these abnormalities. CM, cardiomyocyte; PTEN, phosphatase and tensin homolog; PPAR $\alpha$ , peroxisome proliferator-activated receptor  $\alpha$ ; PGC-1 $\alpha$ , peroxisome proliferator-activated receptor gamma coactivator-related protein 1 $\alpha$ .

there is a large body of evidence demonstrating that chronic and deregulated activation of mTOR is detrimental during cardiac stress, whereas its pharmacological inhibition is protective (20). SIRT1 was also found to be a target of miR-22 (18), so that activation of SIRT1 may represent another mechanism through which miR-22 inhibition may reduce age-related cardiac diseases. We previously found that SIRT1 reduces cardiac aging and confers resistance to oxidative stress (21), and SIRT1 has also been shown to limit ischemic injury (22). Finally, miR-22 inhibition may upregulate PGC-1 $\alpha$ , another miR-22 target. PGC-1 $\alpha$  controls mitochondrial biogenesis and its upregulation may favor mitochondrial turnover and proper mitochondrial function, which are usually impaired in the old heart (2). In this regard, mice with PGC-1 $\alpha$  gene deletion display an impaired energy state and develop contractile dysfunction during aging (23).

One of the most remarkable results of the study by Gupta *et al.* is the demonstration that miR-22 dysregulation is relevant to human disease. In fact, the authors found

that high circulating levels of miR-22 are associated with a higher cardiovascular mortality in patients affected by heart failure (11). Future studies are warranted to investigate whether miR-22 can be used as a biomarker to monitor the efficacy of pharmacological therapy in heart failure patients. It will be important to evaluate whether the pharmacological inhibition of miR-22 ameliorates symptoms and outcomes in these subjects. Of note, atorvastatin was previously seen to reduce the expression of miR-22 in cardiomyocytes (24) and was also shown to ameliorate heart failure in patients (25).

In conclusion, Gupta *et al.* provided compelling results indicating that miR-22 upregulation in the heart is responsible for the reduction of autophagy and the increased susceptibility to stress during aging. miR-22 may be considered as a potential therapeutic target for the treatment of age-related diseases (*Figure 1*).

### Acknowledgements

We thank Daniela Zablocki for critical reading of the

manuscript.

**Funding:** This work was supported in part by U.S. Public Health Service Grants HL67724, HL91469, HL102738, HL112330 and AG23039 (J Sadoshima) and by the Leducq Foundation Transatlantic Network of Excellence (J Sadoshima). This work was also supported by a grant from the Italian Ministry of Health (GR-201302355401) to S Sciarretta.

## Footnote

**Conflicts of Interest:** The authors have no conflicts of interest to declare.

## References

1. Writing Group Members., Mozaffarian D, Benjamin EJ, et al. Heart Disease and Stroke Statistics-2016 Update: A Report From the American Heart Association. *Circulation* 2016;133:e38-360.
2. Shirakabe A, Ikeda Y, Sciarretta S, et al. Aging and Autophagy in the Heart. *Circ Res* 2016;118:1563-1576.
3. Levine B, Kroemer G. Autophagy in the pathogenesis of disease. *Cell* 2008;132:27-42.
4. Eisenberg T, Abdellatif M, Schroeder S, et al. Cardioprotection and lifespan extension by the natural polyamine spermidine. *Nat Med* 2016;22:1428-1438.
5. Nakai A, Yamaguchi O, Takeda T, et al. The role of autophagy in cardiomyocytes in the basal state and in response to hemodynamic stress. *Nat Med* 2007;13:619-624.
6. Shirakabe A, Zhai P, Ikeda Y, et al. Drp1-Dependent Mitochondrial Autophagy Plays a Protective Role Against Pressure Overload-Induced Mitochondrial Dysfunction and Heart Failure. *Circulation* 2016;133:1249-1263.
7. Maejima Y, Kyo S, Zhai P, et al. Mst1 inhibits autophagy by promoting the interaction between Beclin1 and Bcl-2. *Nat Med* 2013;19:1478-1488.
8. Sciarretta S, Boppana VS, Umaphathi M, et al. Boosting autophagy in the diabetic heart: a translational perspective. *Cardiovasc Diagn Ther* 2015;5:394-402.
9. Sciarretta S, Zhai P, Shao D, et al. Rheb is a critical regulator of autophagy during myocardial ischemia: pathophysiological implications in obesity and metabolic syndrome. *Circulation* 2012;125:1134-1146.
10. Matsui Y, Takagi H, Qu X, et al. Distinct roles of autophagy in the heart during ischemia and reperfusion: roles of AMP-activated protein kinase and Beclin 1 in mediating autophagy. *Circ Res* 2007;100:914-922.
11. Gupta SK, Foinquinos A, Thum S, et al. Preclinical Development of a MicroRNA-Based Therapy for Elderly Patients With Myocardial Infarction. *J Am Coll Cardiol* 2016;68:1557-1571.
12. Shvets E, Fass E, Elazar Z. Utilizing flow cytometry to monitor autophagy in living mammalian cells. *Autophagy* 2008;4:621-628.
13. Jazbutyte V, Fiedler J, Kneitz S, et al. MicroRNA-22 increases senescence and activates cardiac fibroblasts in the aging heart. *Age (Dordr)* 2013;35:747-762.
14. Ucar A, Gupta SK, Fiedler J, et al. The miRNA-212/132 family regulates both cardiac hypertrophy and cardiomyocyte autophagy. *Nat Commun* 2012;3:1078.
15. Higashi K, Yamada Y, Minatoguchi S, et al. MicroRNA-145 repairs infarcted myocardium by accelerating cardiomyocyte autophagy. *Am J Physiol Heart Circ Physiol* 2015;309:H1813-H1826.
16. Huang ZP, Chen J, Seok HY, et al. MicroRNA-22 regulates cardiac hypertrophy and remodeling in response to stress. *Circ Res* 2013;112:1234-1243.
17. Gurha P, Abreu-Goodger C, Wang T, et al. Targeted deletion of microRNA-22 promotes stress-induced cardiac dilation and contractile dysfunction. *Circulation* 2012;125:2751-2761.
18. Gurha P, Wang T, Larimore AH, et al. microRNA-22 promotes heart failure through coordinate suppression of PPAR/ERR-nuclear hormone receptor transcription. *PLoS One* 2013;8:e75882.
19. Xu XD, Song XW, Li Q, et al. Attenuation of microRNA-22 derepressed PTEN to effectively protect rat cardiomyocytes from hypertrophy. *J Cell Physiol* 2012;227:1391-1398.
20. Sciarretta S, Volpe M, Sadoshima J. Mammalian target of rapamycin signaling in cardiac physiology and disease. *Circ Res* 2014;114:549-564.
21. Alcendor RR, Gao S, Zhai P, et al. Sirt1 regulates aging and resistance to oxidative stress in the heart. *Circ Res* 2007;100:1512-1521.
22. Hsu CP, Zhai P, Yamamoto T, et al. Silent information regulator 1 protects the heart from ischemia/reperfusion. *Circulation* 2010;122:2170-2182.
23. Arany Z, He H, Lin J, et al. Transcriptional coactivator PGC-1 alpha controls the energy state and contractile function of cardiac muscle. *Cell Metab* 2005;1:259-271.
24. Tu Y, Wan L, Bu L, et al. MicroRNA-22 downregulation by atorvastatin in a mouse model of cardiac hypertrophy:

a new mechanism for antihypertrophic intervention. *Cell Physiol Biochem* 2013;31:997-1008.

25. Sola S, Mir MQ, Lerakis S, et al. Atorvastatin improves

left ventricular systolic function and serum markers of inflammation in nonischemic heart failure. *J Am Coll Cardiol* 2006;47:332-337.

**Cite this article as:** Sciarretta S, De Falco E, Frati G, Sadoshima J. How to be young at heart? miR-22 as a potential therapeutic target to boost autophagy and protect the old myocardium. *Ann Transl Med* 2017;5(3):52. doi: 10.21037/atm.2017.01.52

## Another promise against ischemia reperfusion injury: every success raises new questions

Dennis V. Cokkinos

Heart and Vessel Department, Biomedical Research Foundation Academy of Athens, 115 27 Athens, Greece

*Correspondence to:* Dennis V. Cokkinos, MD. Heart and Vessel Department, Biomedical Research Foundation Academy of Athens, 4 Soranou Ephessiou St., 115 27 Athens, Greece. Email: dcokkinos@bioacademy.gr.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Bian C, Xu T, Zhu H, *et al.* Luteolin Inhibits Ischemia/Reperfusion-Induced Myocardial Injury in Rats via Downregulation of microRNA-208b-3p. *PLoS One* 2015;10:e0144877.

Submitted Jul 28, 2016. Accepted for publication Jul 30, 2016.

doi: 10.21037/atm.2016.08.33

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.08.33>

The study by Bian *et al.* (1) raises many important deliberations. Briefly, they found that luteolin (LUT), a flavonoid found in many vegetables, fruits and seeds, inhibits ischemia/reperfusion induced myocardial injury (IRI) in rats.

The first question is: how relevant to the clinical situation are results on IRI alleviation in the experimental setting. The number of successful interventions in animals is legion. However, against these promising results, the very important position paper of the Working Group of Cellular Biology of the Heart of the European Society of Cardiology (2) should be remembered: the experts producing this paper concluded only 3 years ago that there is no effective proven therapy against IRI. It is widely recognized that it is not always possible to translate animal experiments into clinical therapy.

According to Bian *et al.* (1), LUT joins a long list of herbal medicines proposed to have wonder qualities, such as quercetin, curcumin, resveratrol, and many others. LUT is followed by an impressive list of references, supporting it as an antineoplastic and cardioprotective agent. In its latter role LUT has been given only in rodents, mostly on isolated cardiomyocytes (3,4) or hearts under Langendorff perfusion as in the present study (3,5) or both (4,6). Sun *et al.* (7) used the drug in diabetic rats undergoing coronary artery occlusion for 30 minutes followed by 3 hours of reperfusion. The drug was administered via tail vein injection, which makes it a feasible agent for employment in the clinical arena as an adjunct to primary percutaneous

coronary intervention.

Many mechanisms have been proposed for the cardioprotective effects of LUT against IRI. Thus in previous studies, Qi *et al.* (8) from the same group found that LUT decreased both necrosis and apoptosis. As regards apoptosis, they found that LUT upregulated Bcl-2, decreased the ratio of Bax to Bcl-2 and inhibited the activation of caspase-3. These findings are important since IRI engenders both necrosis and apoptosis (9). Xu *et al.* (3) again from the same group in a review recapitulate the action of LUT on the following processes, involved in apoptosis: upregulation of phosphorylated Akt, suppression of NF- $\kappa$ B activation, increase of Bcl-2, inhibition of caspase-8 and -3.

Fang *et al.* (4) additionally reported that LUT increased phosphorylated SERCA-2 and phospholamban as opposed to control, through the p13K/Akt pathway. Also, Wu *et al.* (6) found that it activates pERK and inhibits the JNK pathway. Reduction of JNK, and p38 MAPK are also reported by Cheng *et al.* (10) in rat cortical necrosis.

Sun *et al.* (7) also showed that LUT inhibits LDH release (suggestive of necrosis inhibition) and in addition to its decrease of the Bax to Bcl-2 ratio, it upregulated the anti-apoptotic proteins FGFR2 and LIF and increased BAD phosphorylation. It also inhibited MPO expression and inflammatory cytokine production, including IL-6, IL-1 $\alpha$  and TNF $\alpha$ . They also found that LUT decreased

infarct size as measured by Evans Blue/TTC staining, and myocardial apoptosis, as assessed by TUNEL, while improving cardiac function as assessed by echocardiography, and the incidence of arrhythmia.

In the index study, Bian *et al.* (1) studied Langendorff perfused rat hearts, subjected to ischemia for 30 min and reperfused for 120 min, in which cardiac function was studied with a pressure sensor Millar catheter. They found that LUT pretreatment in the perfusate improved cardiac function and also had the following effects on microRNAs (miRs): it downregulated the expression of 4 miRs, including miR 208b-3p and upregulated the expression of 26 miRs.

Additionally, they transfected the H9c2 cells with a specific miR-208b-3p mimic or a duplex RNA inhibitor to effectively overexpress or knock down miR 208b-3p.

Then after the cells were transfected with a siRNA to knock down Ets-1, they underwent an anoxia/reoxygenation protocol (A/R simulation of I/R).

They showed that in the cell culture LUT-pretreatment protected the cells against A/R injury, reversing the appearance of dead cells, while significantly downregulating miR 208b-3p. They verified that by administering the miR 208b-3p mimic; this miR was very robustly overexpressed with increased cell apoptosis, while it was greatly underexpressed when incubated with its inhibitor, with resulting decreased apoptosis.

Also, with the most effective Ets-1-SiRNA sequence, Ets-1 was strongly underexpressed, with an increase of H9c2 cells in the early phase of apoptosis.

The administration of LUT was accompanied by a corresponding decrease of pro-(caspase-3 and Bax) and increase of anti-(Bcl-2) apoptotic agents. The miR 208b-3p mimic decreased Ets-1 protein levels. These levels were however increased with LUT addition and miR-208b-3p inhibition.

These results give some important messages:

First, the decrease of miR-208b-3p expression by LUT is potentially of significance. This miR has been found to be increased after an acute myocardial infarction (11). The novel finding that its inhibition decreases apoptosis can have important clinical consequences. Equally importantly, miR-208b-3p has been associated with post infarct myocardial remodeling (REM), being one of the main miRs associated with this unfavorable course (12).

Thus, an agent, such as LUT could potentially affect a diminution of myocardial death both acutely, signifying cardioprotection, and chronically, signifying

cardiopreservation: since LUT is easy to administer orally, a chronic experiment to evaluate its action against REM is in order.

When a miR is found effective in the living organisms, theories about its potential clinical value immediately arise. However, up to now the effectiveness of antagomirs given systematically is less than satisfactory (13); direct local infusion is needed. The same holds true for miR-mimetics, which need to be attached to lipoparticles or to viral vectors (5). Nanoparticle therapy clinically still belongs to the future.

Thus, the use of a simple substance which can readily and robustly manipulate miR expression is very promising.

A second finding attributed to LUT should not be overlooked. Through its diverse reported actions it has been found to decrease the incidence of myocardial infarction in the Zutphen elderly study (14). If this—as logically expected—could also include re-infarctions, LUT could protect patient populations with coronary artery disease against REM by decreasing re-infarctions, such as Kjekshus (15) has advocated to happen with the use of statins.

The study of Bian *et al.* (1) brings into focus another problem. The authors state that LUT increases Ets1 protein levels. The same result was seen with transfection with a miR-208b-3p inhibitor. Thus they suggest that Ets1 is a target gene of miR-208b-3p. They pertinently state that there exist relatively few reports on the role of Ets-1 on cardiomyocyte apoptosis. However, Wang *et al.* (16) found that in hyperglycemia, HMGB1 induces apoptosis via an ERK/Ets-1 pathway; moreover, caspase is its direct target gene (17).

Ets-1 can induce inflammation and apoptosis in endothelial cells (18,19) but reduce apoptosis in vascular smooth muscle cells (20) which share many properties with cardiomyocytes. Thus, the role of Ets1 on cardiomyocytes needs further study.

Lastly, I must point out another source of perplexity: LUT is advanced as being both cardioprotective through its inhibition of apoptosis of cardiac cells and anti-neoplastic through its promotion of apoptosis and inhibition of angiogenesis (21).

Cai *et al.* (21) showed that in human pancreatic carcinoma cells it increased Bax and caspase-3 and decreased Bcl-2 expression. These are exactly the opposite to what Qi *et al.* (8) and Fang *et al.* (4) have found in the rat heart.

Is the Yin and Yang of Tao philosophy taken too far? Invariably, transcriptional pathways follow similar courses in biological processes, such as cardiac, neoplastic and diabetic



system perturbations. Could the very intense proliferation rate of neoplastic cells and the marked quiescence of cardiomyocytes explain this difference? It is interesting that many cancer cells have higher than normal Akt (22) and Bcl-2 levels (23). Obviously these perplexing biological oddities should be studied further. They also often manifest downexpression of the apoptotic phosphatase PTEN (22), an antagonist of pro-survival PI3K which is highly expressed in cardiomyocytes, and inactivation of caspases (24). These differences may play a role.

Here, the author must confess that he is skeptical towards oriental herbal medications, which are advanced as a panacea (an all-curing medicine in ancient and modern Greek). Still one should never dismiss ideas which are not to his preference. It should not be overlooked that very recently, Eggebeen *et al.* (24) in JACC Heart Failure reported that beetroot juice has beneficial effects in older patients with heart failure and preserved ejection fraction. This is American herbal medicine at its best.

Thus, I would conclude that the elegant study of Bian *et al.* (1), a group consistently studying LUT over the years, preempts a host of clinical and theoretical considerations and definitely warrants further study, both acutely and also due to its ease of use against chronic cardiac REM, a subject on which this author has been working for over 20 years (25).

## Acknowledgements

None.

## Footnote

*Conflicts of Interest:* The author has no conflicts of interest to declare.

## References

- Bian C, Xu T, Zhu H, et al. Luteolin Inhibits Ischemia/Reperfusion-Induced Myocardial Injury in Rats via Downregulation of microRNA-208b-3p. *PLoS One* 2015;10:e0144877.
- Hausenloy DJ, Erik Bøtker H, Condorelli G, et al. Translating cardioprotection for patient benefit: position paper from the Working Group of Cellular Biology of the Heart of the European Society of Cardiology. *Cardiovasc Res* 2013;98:7-27.
- Xu T, Li D, Jiang D. Targeting cell signaling and apoptotic pathways by luteolin: cardioprotective role in rat cardiomyocytes following ischemia/reperfusion. *Nutrients* 2012;4:2008-2019.
- Fang F, Li D, Pan H, et al. Luteolin inhibits apoptosis and improves cardiomyocyte contractile function through the PI3K/Akt pathway in simulated ischemia/reperfusion. *Pharmacology* 2011;88:149-158.
- Mericskay M. MicroRNAs: what cardiologists should know about them? *Presse Med* 2015;44:761-771.
- Wu X, Xu T, Li D, et al. ERK/PP1a/PLB/SERCA2a and JNK pathways are involved in luteolin-mediated protection of rat hearts and cardiomyocytes following ischemia/reperfusion. *PLoS One* 2013;8:e82957.
- Sun D, Huang J, Zhang Z, et al. Luteolin limits infarct size and improves cardiac function after myocardium ischemia/reperfusion injury in diabetic rats. *PLoS One* 2012;7:e33491.
- Qi L, Pan H, Li D, et al. Luteolin improves contractile function and attenuates apoptosis following ischemia-reperfusion in adult rat cardiomyocytes. *Eur J Pharmacol* 2011;668:201-207.
- Hausenloy DJ, Yellon DM. Ischaemic conditioning and reperfusion injury. *Nat Rev Cardiol* 2016;13:193-209.
- Cheng HY, Hsieh MT, Tsai FS, et al. Neuroprotective effect of luteolin on amyloid beta protein (25-35)-induced toxicity in cultured rat cortical neurons. *Phytother Res* 2010;24 Suppl 1:S102-S108.
- Ji X, Takahashi R, Hiura Y, et al. Plasma miR-208 as a biomarker of myocardial injury. *Clin Chem* 2009;55:1944-1949.
- Lv P, Zhou M, He J, et al. Circulating miR-208b and miR-34a are associated with left ventricular remodeling after acute myocardial infarction. *Int J Mol Sci* 2014;15:5774-5788.
- Li P. MicroRNAs in cardiac apoptosis. *J Cardiovasc Transl Res* 2010;3:219-224.
- Hertog MG, Feskens EJ, Hollman PC, et al. Dietary antioxidant flavonoids and risk of coronary heart disease: the Zutphen Elderly Study. *Lancet* 1993;342:1007-1011.
- Kjekshus J. Are statins failing in heart failure? *Eur Heart J* 2015;36:1502-1504.
- Wang WK, Lu QH, Zhang JN, et al. HMGB1 mediates hyperglycaemia-induced cardiomyocyte apoptosis via ERK/Ets-1 signalling pathway. *J Cell Mol Med* 2014;18:2311-2320.
- Pei H, Li C, Adereth Y, et al. Caspase-1 is a direct target gene of ETS1 and plays a role in ETS1-induced apoptosis. *Cancer Res* 2005;65:7205-7213.
- Zhu N, Zhang D, Chen S, et al. Endothelial enriched

- microRNAs regulate angiotensin II-induced endothelial inflammation and migration. *Atherosclerosis* 2011;215:286-293.
19. Teruyama K, Abe M, Nakano T, et al. Role of transcription factor Ets-1 in the apoptosis of human vascular endothelial cells. *J Cell Physiol* 2001;188:243-252.
  20. Zhang C, Kavurma MM, Lai A, et al. Ets-1 protects vascular smooth muscle cells from undergoing apoptosis by activating p21WAF1/Cip1: ETS-1 regulates basal and inducible p21WAF1/Cip1 transcription via distinct cis-acting elements in the p21WAF/Cip1 promoter. *J Biol Chem* 2003;278:27903-27909.
  21. Cai X, Lu W, Ye T, et al. The molecular mechanism of luteolin-induced apoptosis is potentially related to inhibition of angiogenesis in human pancreatic carcinoma cells. *Oncol Rep* 2012;28:1353-1361.
  22. Soria JC, Lee HY, Lee JI, et al. Lack of PTEN expression in non-small cell lung cancer could be related to promoter methylation. *Clin Cancer Res* 2002;8:1178-1184.
  23. Martin B, Paesmans M, Berghmans T, et al. Role of Bcl-2 as a prognostic factor for survival in lung cancer: a systematic review of the literature with meta-analysis. *Br J Cancer* 2003;89:55-64.
  24. Eggebeen J, Kim-Shapiro DB, Haykowsky M, et al. One Week of Daily Dosing With Beetroot Juice Improves Submaximal Endurance and Blood Pressure in Older Patients With Heart Failure and Preserved Ejection Fraction. *JACC Heart Fail* 2016;4:428-437.
  25. Cokkinos DV, Chrysanthopoulos S. Thyroid hormones and cardiac remodeling. *Heart Fail Rev* 2016;21:365-372.

**Cite this article as:** Cokkinos DV. Another promise against ischemia reperfusion injury: every success raises new questions. *Ann Transl Med* 2016;4(Suppl 1):S3. doi: 10.21037/atm.2016.08.33

## RNAs that make a heart beat

Mithun Mitra<sup>1,2</sup>, Hilary A. Collier<sup>1,2</sup>

<sup>1</sup>Department of Molecular, Cell and Developmental Biology, University of California, Los Angeles, CA, USA; <sup>2</sup>Department of Biological Chemistry, David Geffen School of Medicine, Los Angeles, CA, USA

Correspondence to: Hilary A. Collier. Department of Molecular, Cell and Developmental Biology, UCLA, Los Angeles, CA 90095, USA. Email: hcoller@ucla.edu.

Provenance: This is a Guest Perspective commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

Comment on: Liu X, Zhang Y, Du W, *et al.* MiR-223-3p as a Novel MicroRNA Regulator of Expression of Voltage-Gated K<sup>+</sup> Channel Kv4.2 in Acute Myocardial Infarction. *Cell Physiol Biochem* 2016;39:102-14.

**Abstract:** An increase in stress-associated microRNAs has been observed in the heart after an induced myocardial infarction. Liu and colleagues now demonstrate that one of these stress-associated microRNAs, *miR-223-3p*, can regulate a component of the voltage-gated channel that mediates rapid outward efflux of potassium during an action potential. Aberrations in the potassium current have been associated with ventricular arrhythmia and heart disease. Strikingly, introducing a small RNA antagonist directed against *miR-223-3p* into rat hearts, while also inducing a myocardial infarction, resulted in a reduction in arrhythmias. We place these studies in the larger context of the field and discuss the potential of anti-*miR-223-3p* molecules as new therapeutics for myocardial infarction.

**Keywords:** Action potential; potassium channel; voltage-gated channel; microRNA; arrhythmia; myocardial infarction

Submitted Sep 29, 2016. Accepted for publication Oct 06, 2016.

doi: 10.21037/atm.2016.11.39

View this article at: <http://dx.doi.org/10.21037/atm.2016.11.39>

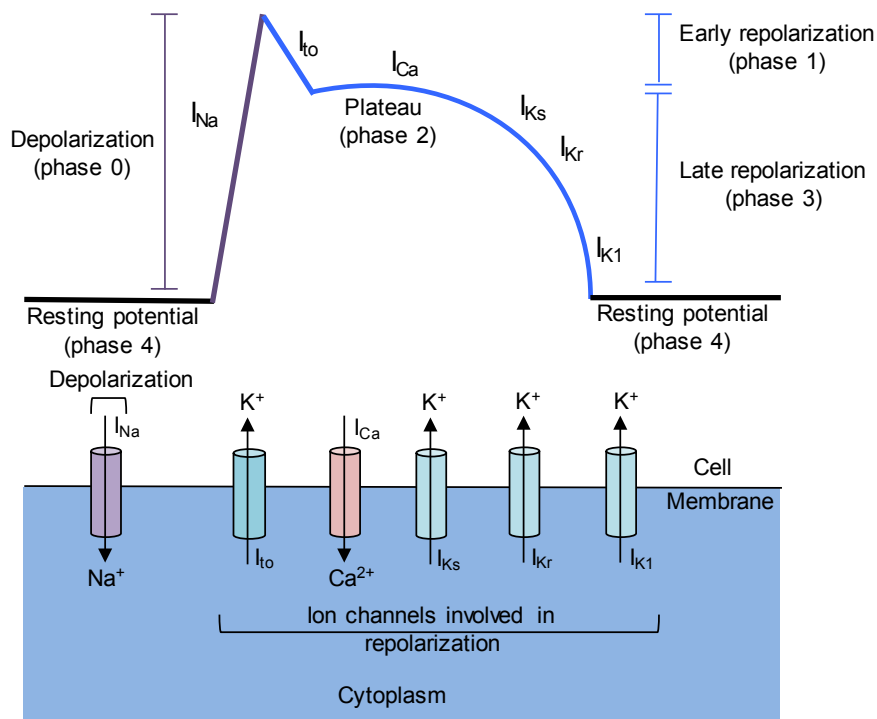
### Action potentials drive heart beat

The beating of a heart is a symbol of life and vitality. The ability of the heart to pump blood throughout the body is essential for bringing oxygen and nutrients to peripheral tissues and removing carbon dioxide and other wastes. Heart beats power this flow of blood. Each time the heart beats, arteries expand and fill with blood; the pauses between the beats allow for constriction followed by a re-expansion with the next heart beat.

The ability of the heart to beat relies on the conversion of electrical to mechanical energy (1). The electrical basis for heart beats are action potentials that start in the specialized pacemaker cells of the heart, and are then transmitted to the atrial and ventricular heart muscle cells, cardiac myocytes, via the passage of ions between cells through gap junctions. Action potentials involve the flow of ions into and out of cardiac myocytes through voltage-gated channels. These channels open and close depending on the

membrane potential, the difference in electrical potential on the inside of the cell compared with the outside (*Figure 1*). During an action potential, the opening and closing of one set of channels leads to a change in membrane potential, thereby allowing a different set of channels to open. These changes lead to entrance or exit of different ions, and subsequent changes in transmembrane potential (3-7).

During cardiac action potentials, there is a flux of sodium (Na<sup>+</sup>) into the cell, along with an inward flux of calcium (Ca<sup>+</sup>), that leads to the depolarization, followed by outward potassium (K<sup>+</sup>) currents that repolarize the cell (1,8-11). An action potential is divided into several phases (1,9-12). In phase 4, the resting phase, the membrane potential of a ventricular cardiomyocyte is about -90 mV because, like most cells, ventricular cardiomyocytes have a more negative charge inside than outside of the cell. The membrane potential is established largely by potassium channels that allow for a loss of positively charged potassium ions from the inside of the cell resulting in a negative membrane



**Figure 1** Example of a ventricular action potential. The membrane potential is plotted over time on the y axis. Different currents active at different phases of the action potential are labeled. Below, the different ion channels that open and close to produce an action potential are shown. Adapted with permission from Kim 2013 (2).

potential. In addition, sodium-potassium pumps actively transport three sodium ions out of the cell and two potassium ions into the cell, both against a concentration and electrical gradient, thus maintaining the concentrations of both ions and preserving the voltage polarization.

Phase 0 is the depolarization phase that initiates the action potential. An action potential can result from an electrical stimulation, or, in the case of the pacemaker cells of the heart, from spontaneous automaticity of the pacemaker cells due to a slowly depolarizing Na channel that generates a pacemaker current called the funny current (10). In ventricular myocytes, rapidly activating, phase 0, voltage-gated channels specific for sodium ions open. Sodium ions are more abundant outside than inside the cell, and opening these channels results in a rush of positively charged sodium ions into the cell ( $I_{Na}$ ). The flooding of sodium ions changes the transmembrane potential from negative to positive, resulting in depolarization of the membrane to about +20 mV.

The subsequent phases involve a repolarization of the membrane as positive ions flow outward from the cell. In the next phase, phase 1, the sodium channels close and

voltage-gated potassium ( $K_v$ ) channels open, creating an outward potassium current  $I_{to}$ . The rapid efflux of positively charged potassium ions results in a sharp decrease in the membrane potential from  $\sim +20$  mV to  $\sim +10$  mV at the end of phase 1. Phase 1 of an action potential can influence the height and duration of the ensuing phases (12,13).

The membrane repolarization of phase 1 activates voltage-gated calcium currents ( $I_{Ca}$ ). In phase 2, a plateau phase, there is a balance between a slow inward flux of calcium ions and a reduced outward flux of potassium ions through a different set of potassium channels, the slow delayed rectifier potassium channels ( $I_{Ks}$ ). The membrane potential in phase 2 ends at  $\sim -20$  mV. The calcium ions that enter the cell in phase 2 act as triggers for contraction of the myofilaments of the heart muscle.

In phase 3, the calcium channels close. The rapid delayed rectifier potassium current ( $I_{Kr}$ ) contributes to the rapid potassium efflux that creates phase 3 of repolarization. Finally, an inward delayed rectifier potassium current ( $I_{K1}$ ) helps establish and maintain the cell transmembrane potential (phase 4). The  $I_{K1}$  current represents the current that must be overcome to establish the next action potential.

A cardiac action potential lasts ~200 msec. After an action potential, there is a refractory period of ~250 msec in which another action potential is unable or less likely to occur. After the refractory period, the cell returns to the resting phase and can respond to signals that evoke a subsequent action potential.

### Voltage-gated potassium channels are tightly regulated to control myocyte function and arrhythmia

Voltage-gated channels allow fast and selective ion permeation that is regulated by opening and closing of a pore through a mechanism that senses transmembrane voltage (14). Voltage-gated channels are composed of alpha pore-forming subunits and accessory subunits (1,15). The pore-forming alpha subunits contain multiple transmembrane domains. The channels can be homotetrameric or heterotetrameric with four voltage-sensitive alpha subunits arranged surrounding a common center that, when open, serves as a pore for ions (15).

The  $K_v$  channel has six transmembrane domains, with both amino and carboxy termini localized within the cell (14). The two transmembrane domains at the carboxy terminus of the protein form the pore for potassium ion flux. The transmembrane domain adjacent to the pore-forming domains contains positively charged arginine and lysine residues and moves in response to membrane depolarization, thereby mediating the voltage-dependent opening of the channel (15).

Through extensive gene duplication events, the human genome encodes 40 voltage-gated potassium channels (12). The expression and regulation of these potassium channels, for instance, in different positions within the heart, can lead to changes in action potential amplitude, duration, waveforms, and rhythmicity (8,12). The transient voltage-gated potassium current ( $I_{to}$ ) in phase one is sometimes divided into  $I_{to,fast}$  and  $I_{to,slow}$  based on the rate of recovery (20–100 ms for the fast current and seconds for the slow current) (11). The pore-forming subunits of  $I_{to,fast}$  are  $K_v4.2$  and  $K_v4.3$ , which are encoded by the *KCND2* and *KCND3* genes. These proteins can likely form pores as homomers or heteromers (16,17).

Heart beats that deviate from the normal rhythm are called arrhythmias (11). Heart failure, a common cause of death worldwide, is frequently associated with arrhythmias and electrical instability. Abnormalities in the repolarization phase of action potentials due to heart failure can contribute

to arrhythmias (11,18). Dramatic changes in the levels and properties of myocardial potassium currents have been observed with cardiac disease (8), including myocardial infarction, the death and destruction of heart muscle as a result of lack of oxygen. In canine models of myocardial infarction, potassium currents are down-regulated in cells in the infarcted zone (19–21), that is, the portion of tissue that is dying or dead due to lack of blood supply. The down-regulation is most pronounced within days following the infarct and returns to normal over the course of two months. These findings highlight the physiological importance of the regulation of potassium channels in the heart.

### miRNAs regulate cardiac function

The regulation of potassium channels in normal physiology or heart disease can occur through multiple mechanisms. Transcriptional regulation plays an important role in the proper expression of different potassium channel components (8). Other mechanisms that regulate potassium channel activity include splicing, RNA editing, and post-translational modifications such as phosphorylation (8,11). In addition, microRNAs have been demonstrated as post-transcriptional regulators of potassium channel expression (2).

Mature microRNAs are small 22–26 nucleotide single-stranded endogenously encoded RNAs (22,23). Originally transcribed as pri-miRNAs, they are processed by the RNase III *Drosha*, *Dgcr8*, and other factors to form a hairpin of ~70 nucleotides in the nucleus. The hairpins are then processed in the cytoplasm by *Dicer* to give rise to mature miRNAs (24–26). These miRNAs can associate with Argonaute proteins in complexes called RNA-induced silencing complexes (RISC). miRNAs in the RISC complex can anneal to mRNA transcripts with similar base pair sequences in their 3'UTRs or coding regions (27,28). In most cases, miRNA targeting leads to the degradation of transcripts or inhibition of their translation (29,30).

Families of miRNAs including *miR-1*, *miR-29*, *miR-15* and *miR-208* have been demonstrated to respond to cardiac stress and play a role in controlling heart function (2,8,11). Van Rooij and colleagues found that microRNAs are up- and down-regulated in cardiac tissue from mice undergoing cardiac stress (31), and in response to myocardial infarction (32). Downregulation of *miR-29*, a microRNA that targets extracellular matrix proteins (33–35), was found to contribute to the fibrotic response post myocardial infarction (32). Ikeda and colleagues extended these studies to humans, and reported reproducible changes in miRNA levels

in human patients with heart failure (36). The functional importance of miRNAs for maintaining healthy hearts has been demonstrated by cardiomyocyte-specific deletion of either *Dicer* or *Dgcr8*. Deletion of *Dicer* in 3-week-old mouse cardiomyocytes resulted in arrhythmias and lethality, while deletion of *Dicer* in the hearts of adult mice led to severe heart failure (37). Similarly, perinatal deletion of *Dgcr8* in mice resulted in severe and lethal heart failure (38).

Overexpression and loss-of function studies have revealed roles for individual miRNAs in heart disease (31,39,40). For example, *miR-208* is expressed from the intron of myosin heavy chain 6 and is expressed specifically in cardiac and slow skeletal muscle (41). *miR-208*-knockout mice respond to cardiac stress with reduced fibrosis and hypertrophy (heart cell enlargement) (42). Inhibition of *miR-208a* in rats fed a high salt diet resulted in improved survival and reduced cardiac fibrosis (43). As another example, *miR-15* is induced in the infarcted region of the heart in response to ischemia-reperfusion injury in mice and pigs (39). Systemic delivery of inhibitors of *miR-15* family members in mice with myocardial infarctions reduced the size of the infarcted region and enhanced cardiac function (39).

In addition to miRNAs that affect the survival of myocardial cells, there are also examples in which miRNAs specifically target mRNAs encoding ion channels. *miR-1* is expressed in heart and skeletal muscle and is overexpressed in individuals with coronary artery disease (44). Homozygous deletion of one of two *miR-1* genes (*miR-1-2*) in mice resulted in mortality of 50% of the offspring by developmental abnormalities (45). Most of the survivors died from sudden cardiac death caused by arrhythmias (45). Several targets of *miR-1* could explain these findings. In addition to the notch ligand delta (46) and the Rho GTPase *Cdc42* (47), *miR-1* also regulates Iroquois related homeobox 5 (*Irx5*), a transcriptional repressor of the  $I_{to}$  component potassium voltage-gated channel, Shal-related family, member 2 (*Kcnd2*) (45). *miR-1* also post-transcriptionally represses potassium voltage-gated channel subfamily J member 2 (*Kcnj2*), which encodes the potassium channel subunit Kir2.1, a component of the inwardly rectifying potassium channel (44), and gap junction protein alpha 1 (*Gja*), which encodes connexin 43, a connexin important for cardiac gap junctions (44). Consistent with an important role for *miR-1* in regulating action potentials, introduction of *miR-1* was found to exacerbate arrhythmias, while introduction of *miR-1* antisense inhibitors in rat hearts undergoing myocardial infarction reduced arrhythmias (44).

### **miR-223-3p is induced with myocardial infarction, targets Kv4.2 in its coding region, and promotes arrhythmia**

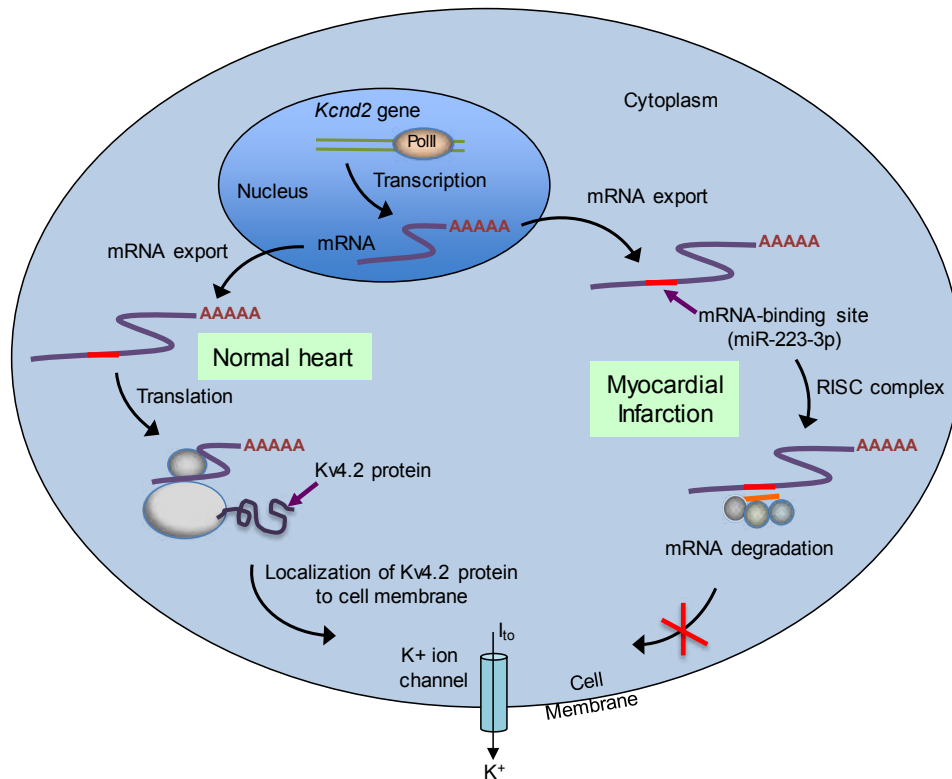
In a recent issue of *Annals of Translational Medicine: Cellular Physiology and Biochemistry*, Liu and colleagues now report that *miR-223-3p* regulates cardiac function (48). Inspired by previous studies showing that *miR-223-3p* is upregulated in hearts after myocardial infarction (32), Liu and colleagues induced myocardial infarction in rats by tying off the artery that supplies the heart with blood (a method called left anterior descending artery ligation). This procedure resulted in arrhythmias in the rats and mortality in 40% of animals. Heart tissue surrounding the infarct area contained strongly elevated levels of *miR-223-3p* and *miR-1*.

Liu and co-authors recognized that *miR-223-3p* is complementary to, and therefore has the potential to bind to, the coding sequence of the *Kcnd2* transcript. *Kcnd2* encodes  $K_v4.2$ , the alpha subunit of the voltage-gated transient outward potassium channel that carries  $I_{to}$  in the rat during phase 1 of an action potential (48) (Figure 2). Consistent with this hypothesis, Liu and colleagues found that  $K_v4.2$  protein levels were lower in the area around the rat's infarcted heart tissue. Further, down-regulation of  $K_v4.2$  was found to be associated with reduced  $I_{to}$  flux. To determine whether  $K_v4.2$  is a bonafide target of the *miR-223-3p* miRNA, the authors subcloned the coding sequence of  $K_v4.2$  into a luciferase-expressing plasmid to generate a luciferase-*Kcnd2* chimeric vector, and transfected the vector into neonatal rat ventricular cardiomyocytes. Co-transfection of *miR-223-3p*, but not a negative control, substantially suppressed luciferase activity. Finally, Liu and colleagues transfected an inhibitor of *miR-223-3p* into the ventricular cardiomyocytes of rats and found that the inhibitor significantly reduced the incidence of arrhythmias after acute myocardial infarction. The findings, taken together, support an important role for *miR-223-3p* as a regulator of cardiac action potentials after a myocardial infarction in rats.

### **miR-223-3p inhibition as a therapy for myocardial infarction-induced arrhythmia**

The findings suggest that inhibiting the induction of *miR-223-3p* during a myocardial infarction could normalize action potentials and benefit patients. Indeed, potassium channels are being recognized as important therapeutic targets and strategies to inhibit potassium channels are being





**Figure 2** Schematic of the research findings of Liu *et al.* (48). In the normal heart, the *Kcnd2* gene encodes expression of the  $K_v4.2$  protein that localizes to the cell membrane and forms voltage-gated potassium channels that contribute to the  $I_{to}$  current. Under conditions of myocardial infarction, *miR-223-3p* is induced and targets  $K_v4.2$  for degradation through a recognition site in the coding region. Reduced levels of the voltage-gated potassium channels contribute to altered action potentials and arrhythmia after a myocardial infarction.

explored as treatment for arrhythmias and other conditions including seizures, pain, and Alzheimer's disease (49). Among the strategies to inhibit potassium channels, miRNA inhibitors are particularly appealing because they recognize specific mRNA sequence (50). Indeed, preclinical models are being developed by miRagen/Servier and other companies for the inhibition of miRNAs associated with cardiovascular disease (50,51).

The possibility of a new therapeutic strategy for myocardial infarction raises several issues. First, upregulation of *miR-223-3p* in human hearts undergoing myocardial infarction and *miR-223-3p* targeting of human *KCND2* would need to be established.

Next, *miR-223*, the same miRNA discovered to be associated with arrhythmia, has also been associated with cardioprotection. In one study, both arms of *miR-223*, 3p and 5p, were discovered to be induced after ischemia reperfusion in mouse hearts (52). But in this study, as opposed to Liu *et al.*, overexpressing a precursor miRNA

containing both strands (5' and 3') of *miR-223* was associated with better contractility and reduced necrosis after myocardial ischemia (52). Transgenic mice with a knockout of the *miR-223* locus exhibited aggravated ischemia-reperfusion-induced cardiac dysfunction and more cell death (52). This study identified two cell death receptors, *Tnfr1* and *Dr6*, as *miR-223* targets in mouse hearts (52). In another study, downregulation of *miR-223* occurred in the hearts of mice with severe sepsis (53). In this study, sepsis-induced mortality, inflammation and cardiac dysfunction were exacerbated in mice with knockout of *miR-223-3p*. Semaphorin 3A was identified as a candidate gene mediating this effect. These studies raise concerns that inhibition of *miR-223-3p* in hearts would have negative consequences.

Another issue is that Liu and colleagues focus on *Kcnd2*, which is well-established as a mediator of outward potassium current in rodent heart left ventricle apex cells. However, heart rates and action potential duration differ

between species. While the phases of a heart beat are similar in all mammals, human hearts beat ~60 times per minute while rodent hearts beat ~600 times each minute (11). Until recently, the prevailing paradigm was that larger mammals such as dogs and humans rely on *KCND3*-encoded channels, while rodents use both *Kcnd2* and *Kcnd3*-encoded channels and thereby achieve more rapid depolarization required for their fast heart beats (11,54,55). However, a recent discovery of a gain-of-function point mutation in *KCND2* in a human patient with sudden cardiac arrest suggests *KCND2*-encoded proteins may be important for proper action potentials in humans as well (54). Thus, additional studies will be needed to determine the importance of  $K_{v4.2}$  in human action potentials.

In addition, achieving the correct level of  $K_{v4.2}$  may be challenging. Adding inhibitors of *miR-223-3p* would be expected to increase  $K_{v4.2}$  and thus  $I_{to}$ . Established anti-arrhythmia drugs inhibit  $I_{to}$  (56), supporting the importance of  $I_{to}$  in establishing proper action potentials. However, inhibition of *miR-223-3p* could result in excess  $K_{v4.2}$ , elevated  $I_{to}$ , and arrhythmias, raising concern that miRNAs targeting  $K_{v4.2}$  may increase rather than reduce arrhythmia.

Other concerns relate to the characteristics of miRNAs. miRNAs tend to have modest effects on the target gene, off-target effects, and more pleiotropic effects than expected. For example, an anti-miR against the cardiac-specific miR-208a was unexpectedly discovered to prevent systemic phenotypes of obesity and metabolic syndrome (57). In addition, because miRNA and miRNA inhibitors are not expected to affect proteins that exist, but rather affect the levels and translation of mRNA transcripts that code for new protein, determining the pharmacokinetics of any proposed therapy will be important. Finally, developing anti-miRNAs that are not rapidly degraded and will accumulate to therapeutic levels in the heart will also be necessary (43).

In summary, proper regulation of the ion channels that control action potentials is critical for normal heart beat. Dysregulation of these channels can contribute to arrhythmia-induced mortality. miRNAs represent an emerging mechanism for regulating the expression levels of ion channel components, and are being developed as novel emerging therapeutic targets for heart disease.

### Acknowledgements

The authors would like to thank Kalyanam Shivkumar and Marmar Vaseghi for helpful comments.

**Funding:** This work was funded by grants to HAC National Institute of General Medical Sciences R01 GM081686 and National Institute of General Medical Sciences R01 GM0866465. HAC is a member of the Eli & Edythe Broad Center of Regenerative Medicine & Stem Cell Research, the Jonsson Comprehensive Cancer Center, the UCLA Molecular Biology Institute, the UCLA Institute for Quantitative and Computational Biology, and the UCLA Bioinformatics Interdepartmental Program.

### Footnote

**Conflicts of Interest:** The authors have no conflicts of interest to declare.

### References

1. Grant AO. Cardiac ion channels. *Circ Arrhythm Electrophysiol* 2009;2:185-194.
2. Kim GH. MicroRNA regulation of cardiac conduction and arrhythmias. *Transl Res* 2013;161:381-392.
3. Hodgkin AL, Huxley AF, Katz B. Measurements of current-voltage relations in the membrane of the giant axon of Loligo. *J Physiol (Lond)* 1952;116:424-448.
4. Hodgkin AL, Huxley AF. A quantitative description of membrane current and its application to conduction and excitation in nerve. *J Physiol (Lond)* 1952;117:500-544.
5. Woodbury LA, Woddbury JW, Hecht HH. Membrane resting and action potentials from single cardiac muscle fibers. *Circulation* 1950;1:264-266.
6. Weidmann S. Effect of current flow on the membrane potential of cardiac muscle. *J Physiol (Lond)* 1951;115:227-236.
7. Hutter OF, Trautwein W. Vagal and sympathetic effects on the pacemaker fibers in the sinus venosus of the heart. *J Gen Physiol* 1956;39:715-733.
8. Yang KC, Nerbonne JM. Mechanisms contributing to myocardial potassium channel diversity, regulation and remodeling. *Trends Cardiovasc Med* 2016;26:209-218.
9. Qu Z, Hu G, Garfinkel A, et al. Nonlinear and Stochastic Dynamics in the Heart. *Phys Rep* 2014;543:61-162.
10. Bartos DC, Grandi E, Ripplinger CM. Ion Channels in the Heart. *Compr Physiol* 2015;5:1423-1464.
11. Schmitt N, Grunnet M, Olesen SP. Cardiac potassium channel subtypes: new roles in repolarization and arrhythmia. *Physiol Rev* 2014;94:609-653.
12. Nerbonne JM, Kass RS. Molecular physiology of cardiac repolarization. *Physiol Rev* 2005;85:1205-1253.



13. Delmar M. Role of potassium currents on cell excitability in cardiac ventricular myocytes. *J Cardiovasc Electrophysiol* 1992;3:474-486.
14. Yellen G. The voltage-gated potassium channels and their relatives. *Nature* 2002;419:35-42.
15. Birnbaum SG, Varga AW, Yuan LL, et al. Structure and function of Kv4-family transient potassium channels. *Physiol Rev* 2004;84:803-833.
16. Liu J, Kim KH, Morales MJ, et al. Kv4.3-Encoded Fast Transient Outward Current Is Presented in Kv4.2 Knockout Mouse Cardiomyocytes. *PLoS One* 2015;10:e0133274.
17. Guo W, Jung WE, Marionneau C, et al. Targeted deletion of Kv4.2 eliminates I(to,f) and results in electrical and molecular remodeling, with no evidence of ventricular hypertrophy or myocardial dysfunction. *Circ Res* 2005;97:1342-1350.
18. Näbauer M, Kääh S. Potassium channel down-regulation in heart failure. *Cardiovasc Res* 1998;37:324-334.
19. Dun W, Baba S, Yagi T, et al. Dynamic remodeling of K<sup>+</sup> and Ca<sup>2+</sup> currents in cells that survived in the epicardial border zone of canine healed infarcted heart. *Am J Physiol Heart Circ Physiol* 2004;287:H1046-H1054.
20. Jiang M, Cabo C, Yao J, et al. Delayed rectifier K currents have reduced amplitudes and altered kinetics in myocytes from infarcted canine ventricle. *Cardiovasc Res* 2000;48:34-43.
21. Pinto JM, Boyden PA. Reduced inward rectifying and increased E-4031-sensitive K<sup>+</sup> current density in arrhythmogenic subendocardial purkinje myocytes from the infarcted heart. *J Cardiovasc Electrophysiol* 1998;9:299-311.
22. Ambros V. The functions of animal microRNAs. *Nature* 2004;431:350-355.
23. Kloosterman WP, Plasterk RH. The diverse functions of microRNAs in animal development and disease. *Dev Cell* 2006;11:441-450.
24. Zamore PD, Tuschl T, Sharp PA, et al. RNAi: double-stranded RNA directs the ATP-dependent cleavage of mRNA at 21 to 23 nucleotide intervals. *Cell* 2000;101:25-33.
25. Grishok A, Pasquinelli AE, Conte D, et al. Genes and mechanisms related to RNA interference regulate expression of the small temporal RNAs that control *C. elegans* developmental timing. *Cell* 2001;106:23-34.
26. Hutvagner G, McLachlan J, Pasquinelli AE, et al. A cellular function for the RNA-interference enzyme Dicer in the maturation of the let-7 small temporal RNA. *Science* 2001;293:834-838.
27. Grimson A, Farh KK, Johnston WK, et al. MicroRNA targeting specificity in mammals: determinants beyond seed pairing. *Mol Cell* 2007;27:91-105.
28. Forman JJ, Legesse-Miller A, Collier HA. A search for conserved sequences in coding regions reveals that the let-7 microRNA targets Dicer within its coding sequence. *Proc Natl Acad Sci U S A* 2008;105:14879-14884.
29. Filipowicz W, Bhattacharyya SN, Sonenberg N. Mechanisms of post-transcriptional regulation by microRNAs: are the answers in sight? *Nat Rev Genet* 2008;9:102-114.
30. Guo H, Ingolia NT, Weissman JS, et al. Mammalian microRNAs predominantly act to decrease target mRNA levels. *Nature* 2010;466:835-840.
31. van Rooij E, Sutherland LB, Liu N, et al. A signature pattern of stress-responsive microRNAs that can evoke cardiac hypertrophy and heart failure. *Proc Natl Acad Sci U S A* 2006;103:18255-18260.
32. van Rooij E, Sutherland LB, Thatcher JE, et al. Dysregulation of microRNAs after myocardial infarction reveals a role of miR-29 in cardiac fibrosis. *Proc Natl Acad Sci U S A* 2008;105:13027-13032.
33. Suh EJ, Remillard MY, Legesse-Miller A, et al. A microRNA network regulates proliferative timing and extracellular matrix synthesis during cellular quiescence in fibroblasts. *Genome Biol* 2012;13:R121.
34. Cushing L, Kuang P, Lu J. The role of miR-29 in pulmonary fibrosis. *Biochem Cell Biol* 2015;93:109-118.
35. Maurer B, Stanczyk J, Jungel A, et al. MicroRNA-29, a key regulator of collagen expression in systemic sclerosis. *Arthritis Rheum* 2010;62:1733-1743.
36. Ikeda S, Kong SW, Lu J, et al. Altered microRNA expression in human heart disease. *Physiol Genomics* 2007;31:367-373.
37. da Costa Martins PA, Bourajaj M, Gladka M, et al. Conditional dicer gene deletion in the postnatal myocardium provokes spontaneous cardiac remodeling. *Circulation* 2008;118:1567-1576.
38. Rao PK, Toyama Y, Chiang HR, et al. Loss of cardiac microRNA-mediated regulation leads to dilated cardiomyopathy and heart failure. *Circ Res* 2009;105:585-594.
39. Hullinger TG, Montgomery RL, Seto AG, et al. Inhibition of miR-15 protects against cardiac ischemic injury. *Circ Res* 2012;110:71-81.
40. Mutharasan RK, Nagpal V, Ichikawa Y, et al. microRNA-210 is upregulated in hypoxic cardiomyocytes

- through Akt- and p53-dependent pathways and exerts cytoprotective effects. *Am J Physiol Heart Circ Physiol* 2011;301:H1519-H1530.
41. van Rooij E, Quiat D, Johnson BA, et al. A family of microRNAs encoded by myosin genes governs myosin expression and muscle performance. *Dev Cell* 2009;17:662-673.
  42. van Rooij E, Sutherland LB, Qi X, et al. Control of stress-dependent cardiac growth and gene expression by a microRNA. *Science* 2007;316:575-579.
  43. Montgomery RL, Hullinger TG, Semus HM, et al. Therapeutic inhibition of miR-208a improves cardiac function and survival during heart failure. *Circulation* 2011;124:1537-1547.
  44. Yang B, Lin H, Xiao J, et al. The muscle-specific microRNA miR-1 regulates cardiac arrhythmogenic potential by targeting GJA1 and KCNJ2. *Nat Med* 2007;13:486-491.
  45. Zhao Y, Ransom JF, Li A, et al. Dysregulation of cardiogenesis, cardiac conduction, and cell cycle in mice lacking miRNA-1-2. *Cell* 2007;129:303-317.
  46. Kwon C, Han Z, Olson EN, et al. MicroRNA1 influences cardiac differentiation in *Drosophila* and regulates Notch signaling. *Proc Natl Acad Sci U S A* 2005;102:18986-18991.
  47. Qian L, Wythe JD, Liu J, et al. Tinman/Nkx2-5 acts via miR-1 and upstream of Cdc42 to regulate heart function across species. *J Cell Biol* 2011;193:1181-1196.
  48. Liu X, Zhang Y, Du W, et al. MiR-223-3p as a Novel MicroRNA Regulator of Expression of Voltage-Gated K<sup>+</sup> Channel Kv4.2 in Acute Myocardial Infarction. *Cell Physiol Biochem* 2016;39:102-114.
  49. Wulff H, Castle NA, Pardo LA. Voltage-gated potassium channels as therapeutic targets. *Nat Rev Drug Discov* 2009;8:982-1001.
  50. Li Z, Rana TM. Therapeutic targeting of microRNAs: current status and future challenges. *Nat Rev Drug Discov* 2014;13:622-638.
  51. Dangwal S, Thum T. microRNA therapeutics in cardiovascular disease models. *Annu Rev Pharmacol Toxicol* 2014;54:185-203.
  52. Qin D, Wang X, Li Y, et al. MicroRNA-223-5p and -3p Cooperatively Suppress Necroptosis in Ischemic/Reperfused Hearts. *J Biol Chem* 2016;291:20247-20259.
  53. Wang X, Huang W, Yang Y, et al. Loss of duplexmiR-223 (5p and 3p) aggravates myocardial depression and mortality in polymicrobial sepsis. *Biochim Biophys Acta* 2014;1842:701-11.
  54. Perrin MJ, Adler A, Green S, et al. Evaluation of genes encoding for the transient outward current (I<sub>to</sub>) identifies the KCND2 gene as a cause of J-wave syndrome associated with sudden cardiac death. *Circ Cardiovasc Genet* 2014;7:782-789.
  55. Dixon JE, Shi W, Wang HS, et al. Role of the Kv4.3 K<sup>+</sup> channel in ventricular muscle. A molecular correlate for the transient outward current. *Circ Res* 1996;79:659-668.
  56. Feng J, Wang Z, Li GR, et al. Effects of class III antiarrhythmic drugs on transient outward and ultra-rapid delayed rectifier currents in human atrial myocytes. *J Pharmacol Exp Ther* 1997;281:384-392.
  57. Grueter CE, van Rooij E, Johnson BA, et al. A cardiac microRNA governs systemic energy homeostasis by regulation of MED13. *Cell* 2012;149:671-683.

**Cite this article as:** Mitra M, Coller HA. RNAs that make a heart beat. *Ann Transl Med* 2016;4(23):469. doi: 10.21037/atm.2016.11.39

## Circulating fibrocytes serve as a marker for clinical diagnosis

Thuy Cao<sup>1</sup>, Sheeja Rajasingh<sup>1</sup>, Johnson Rajasingh<sup>1,2</sup>

<sup>1</sup>Department of Internal Medicine, Cardiovascular Research Institute, <sup>2</sup>Department of Biochemistry and Molecular Biology, University of Kansas Medical Center, Kansas City, KS 66160, USA

*Correspondence to:* Johnson Rajasingh, PhD. Assistant Professor of Internal Medicine, Cardiovascular Research Institute, Division of Cardiovascular Diseases, University of Kansas Medical Center, Kansas City, KS 66160, USA. Email: rjohnson9@kumc.edu.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

*Comment on:* Keeley EC, Schutt RC, Marinescu MA, *et al.* Circulating fibrocytes as predictors of adverse events in unstable angina. *Transl Res* 2016;172:73-83.e1.

Submitted Aug 30, 2016. Accepted for publication Sep 03, 2016.

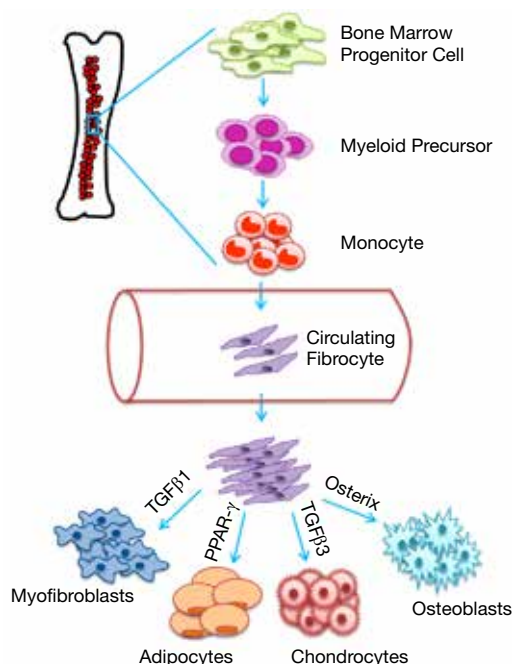
doi: 10.21037/atm.2016.10.26

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.10.26>

Cardiovascular disease continues to be a major health problem in the United States and the leading cause of death (1). According to the American Heart Association, 86.5 million or more than one in three Americans have one or more cardiovascular diseases. Of the 86.5 million, 8.2 million suffer from angina pectoris. The American Heart Association predicts 43.9% of Americans will be afflicted with cardiovascular disease by 2030 (1). Despite marked progress towards the understanding of cardiovascular pathophysiology and rapid improvement in medical and surgical options, the number is still increasing. Therefore, discovery of an early diagnostic tool is important to prevent disease. Fibrocytes are progenitor cells which primarily function in response to inflammation. A recent study by Keeley *et al.* aimed to identify markers in unstable angina that may be used to predict future adverse outcomes (2). They demonstrated that the total number of fibrocytes strongly correlates with recurrent angina and unfavorable clinical events independent of risk factors. There is also evidence of expansion of circulating fibrocytes which express an activated phenotype and myofibroblast differentiation (2). These findings further support the authors' reasoning that fibrocytes have a role in vascular remodeling and their usefulness as markers. However, fibrocytes play an extensive part in immunity and utility as markers for specific pathologies may be difficult.

### Identification of fibrocytes

Circulating fibrocytes are reported for first time in 1994 and are characterized as a distinct population of spindle-shaped cells with the phenotype of CD45+, collagen+, and CD34+ that are present within the blood (3). Fibrosis contributes to the pathology of a variety of diseases (4), particularly inflammatory. Due to the importance of the role of fibrocytes in tissue remodeling, much work is done to investigate the significance of fibrocyte participation in different diseases and to establish markers to detect, determine prognosis, and prevent adverse clinical outcomes. In general, mature fibrocytes have the markers CD34, CD43, CD45, LSP-1, and major histocompatibility complex (MHC) class II, which contributes to their hematopoietic nature and collagen type I and III, which explains their stromal behavior (5). Their ability to migrate to sites of injury is because they contain the markers CCR2, CCR7, and CXCR4. When fibrocytes home to sites of injury and differentiate, they change the expression of their markers. For instance, some may lose CD34 and CD45 and some may express markers to mimic the cells they specialize (5). As a result, their dynamic expression presents an obstacle to track their activity (6). Additionally, fibrocytes are derived from monocytes, thus have characteristics of hematopoietic cells and macrophages along with features of



**Figure 1** Generation of fibroblast and other related cells from bone marrow. Fibrocytes circulated within the blood stream and exit to migrate towards the sites of injury. These cells then differentiate into other cell types depending on the signals they receive.

fibroblasts. Therefore, finding specific markers of fibrocytes is especially arduous. Despite the challenge, one study has found that they can be distinguished from other cells because of the unique combination of CD45RO, 25F9, and S100A8/A9 expression (7). However, the discovery of more specific markers is yet to be determined.

### Function of fibrocytes

Circulating fibrocytes are progenitor cells that originate from bone marrow, which circulate within the bloodstream and principally function to generate components of the extracellular matrix such as vimentin, collagen type I, and collagen type II (8,9). They are derived from monocyte precursors and have characteristics of both macrophages and fibroblasts (10). Under inflammatory conditions, these cells participate in tissue healing and repair. In response to injury, fibrocytes migrate to the inflammatory site via induction by stromal cell-derived factor 1 alpha (SDF-1 $\alpha$ ) (6). Once there, fibrocytes enhance leukocyte

trafficking via increased expression of leukocyte adhesion molecules and recruitment of inflammatory cells through production of interleukin 6 (IL-6), IL-8, CC-chemokine ligand 3 (CCL3), and CCL4 (10). Repair function is initiated in fibrocytes by IL-10 and the presence of apoptotic cells (10). Additionally, neovascularization is promoted by a pro-angiogenic factor, vascular endothelial growth factor (VEGF), released by fibrocytes to aid in the repair process (6). Thus, these cells regulate immune responses via secretion of cytokines and growth factors and stimulate repair through activation of fibroblasts (11). Similar to macrophages, fibrocytes are also involved in antigen presentation to CD8+ T cells and lipid metabolism (10).

Due to the mesenchymal properties of fibrocytes, they are capable of forming myofibroblasts, osteoblasts, and adipocytes (7). The differentiation and activity of fibrocytes are primarily determined by signals in the microenvironment, which activate intracellular pathways (7). For example, a study found that transforming growth factor beta 1 (TGF- $\beta$ 1) increases the expression of type I collagen, alpha-smooth muscle actin ( $\alpha$ SMA), and tissue inhibitor of metalloproteinase-1 (TIMP-1) in fibrocytes which indicates transformation into myofibroblasts (6). Whereas, another study found that fibrocytes are under the influence of TGF- $\beta$ 3, which leads to upregulation of col2A1 and aggrecan and differentiation into chondrocytes (12). Fibrocytes also have the ability to differentiate into osteoblasts when stimulated by Runx2/core binding factor alpha 1 (Cbfa1) and osterix, transcription factors essential for producing bone matrix (12). Exposure of fibrocytes to peroxisome proliferator-activated receptor gamma (PPAR- $\gamma$ ) stimulates transformation into adipocytes (12). Direct regulation of fibrocyte differentiation has been related to CD4+ lymphocytes, which supports differentiation (7). The schematic diagram depicts the origin and differentiation of circulating fibrocytes (Figure 1).

### Clinical implications

Studies have shown that circulating fibrocytes have a role in many cardiac diseases, particularly those involving fibrosis such as coronary heart disease, hypertensive heart disease, and cardiac ischemia (13-15). As such, markers of fibrocytes can provide valuable information regarding the extent of disease and course of treatment. In the study by Keely *et al.*, fibrocytes were examined under the condition

of unstable angina with a focus on their differentiation to myofibroblasts. The combinations of the markers CD45,  $\alpha$ SMA, and collagen 1 were used to identify the fibrocytes and the levels of TGF- $\beta$ 1 to determine the breadth of expansion to myofibroblasts (2). Since both fibrocytes and myofibroblasts have general functions in healing and repair, their markers may be found in several diseases. For example, a study done in neonates with bronchopulmonary dysplasia, increased fibrocytes with  $\alpha$ SMA has been demonstrated when compared to healthy subjects (9). However, the authors recognized that patients with known existing fibrotic diseases or other conditions may trigger fibrocyte activity and excluded them from the study.

Levels of circulating fibrocytes are relatively stable under normal conditions and rises with inflammation and hypoxic situations (13). This phenomenon can be taken advantage of in order to indirectly measure the extent of disease. Furthermore, the study by Keeley *et al.* has shown that the number of total fibrocytes correlates strongly with adverse outcomes in patients with unstable angina (2). However, the relationship is not causative and perhaps the total number of fibrocytes measures the extent of cardiac fibrosis and thus more likely to lead to unfavorable clinical events with higher numbers. Another investigation showed a positive correlation of CXCR4/procollagen-1 and CXCR4/ $\alpha$ SMA fibrocytes with SDF-1/CXCL12<sup>+</sup> expression by infarcted cells in coronary heart disease (16).

Fibrocyte markers can be applied in a clinical setting to help determine prognosis and clinical course. The ability to foresee adverse outcomes as a product of unstable angina can lead to prevention by more aggressive treatments to those at risk (2). In addition, circulating fibrocytes may be used to improve wound healing and perhaps prevent pathological fibrosis (8). More importantly, fibrocytes help determine the magnitude of fibrotic reactions (17) and may be a potential target to inhibit excessive fibrosis as seen in many inflammatory diseases. For example, class I histone deacetylase (HDAC) inhibitors regulate differentiation of fibrocytes and leads to a reduction of both fibrocytes in the heart and circulating fibrocytes when cardiac fibrosis is induced with angiotensin II (18). Moreover, fibrocytes may have the potential in regenerative medicine due to their ability to form other cell types. For instance, the capacity to form chondrocytes and osteoblasts can be used in repair, especially in damage to the articular cartilage (12). In the last few years, there have been many studies aimed to understand the importance of circulating fibrocytes in various diseases, including cardiovascular diseases. However,

the currently available data suggest that circulating fibroblast might be a novel and promising therapeutic target and a marker for treatment response and prognostic evaluation.

## Conclusions

Circulating fibrocytes have important biologic roles, but can also contribute to diseases related to fibrosis. Therefore, understanding these functions under normal conditions can help to prevent aberrant fibrotic processes and identification of more specific cell-surface markers may be used to predict the clinical course. The clinical importance of fibrocytes is not limited to cardiac disease, but can be virtually applied to all fibrotic diseases and may extend into regenerative medicine.

## Acknowledgements

*Funding:* This work was supported, in part, by American Heart Association Grant-in-Aid 16GRNT30950010 and National Institutes of Health COBRE grant P20GM104936 (to J Rajasingh).

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

1. Writing Group Members, Mozaffarian D, Benjamin EJ, et al. Executive Summary: Heart Disease and Stroke Statistics--2016 Update: A Report From the American Heart Association. *Circulation* 2016;133:447-454.
2. Keeley EC, Schutt RC, Marinescu MA, et al. Circulating fibrocytes as predictors of adverse events in unstable angina. *Transl Res* 2016;172:73-83.e1.
3. Bucala R, Spiegel LA, Chesney J, et al. Circulating fibrocytes define a new leukocyte subpopulation that mediates tissue repair. *Mol Med* 1994;1:71-81.
4. Alhamad EH, Shakoor Z, Al-Kassimi FA, et al. Rapid detection of circulating fibrocytes by flowcytometry in idiopathic pulmonary fibrosis. *Ann Thorac Med* 2015;10:279-283.
5. Pilling D, Fan T, Huang D, et al. Identification of markers that distinguish monocyte-derived fibrocytes from monocytes, macrophages, and fibroblasts. *PLoS One*

- 2009;4:e7475.
6. Suga H, Rennert RC, Rodrigues M, et al. Tracking the elusive fibrocyte: identification and characterization of collagen-producing hematopoietic lineage cells during murine wound healing. *Stem Cells* 2014;32:1347-1360.
  7. Smith TJ. Potential role for bone marrow-derived fibrocytes in the orbital fibroblast heterogeneity associated with thyroid-associated ophthalmopathy. *Clin Exp Immunol* 2010;162:24-31.
  8. Quan TE, Cowper SE, Bucala R. The role of circulating fibrocytes in fibrosis. *Curr Rheumatol Rep* 2006;8:145-150.
  9. Li C, Li X, Deng C, et al. Circulating Fibrocytes Are Increased in Neonates with Bronchopulmonary Dysplasia. *PLoS One* 2016;11:e0157181.
  10. Reilkoff RA, Bucala R, Herzog EL. Fibrocytes: emerging effector cells in chronic inflammation. *Nat Rev Immunol* 2011;11:427-435.
  11. Pilling D, Gomer RH. Differentiation of circulating monocytes into fibroblast-like cells. *Methods Mol Biol* 2012;904:191-206.
  12. Choi YH, Burdick MD, Strieter RM. Human circulating fibrocytes have the capacity to differentiate osteoblasts and chondrocytes. *Int J Biochem Cell Biol* 2010;42:662-671.
  13. Lin RJ, Su ZZ, Liang SM, et al. Role of Circulating Fibrocytes in Cardiac Fibrosis. *Chin Med J (Engl)* 2016;129:326-331.
  14. Chu PY, Mariani J, Finch S, et al. Bone marrow-derived cells contribute to fibrosis in the chronically failing heart. *Am J Pathol* 2010;176:1735-1742.
  15. van Amerongen MJ, Bou-Gharios G, Popa E, et al. Bone marrow-derived myofibroblasts contribute functionally to scar formation after myocardial infarction. *J Pathol* 2008;214:377-386.
  16. Lei PP, Qu YQ, Shuai Q, et al. Fibrocytes are associated with the fibrosis of coronary heart disease. *Pathol Res Pract* 2013;209:36-43.
  17. Baker DW, Tsai YT, Weng H, et al. Alternative strategies to manipulate fibrocyte involvement in the fibrotic tissue response: pharmacokinetic inhibition and the feasibility of directed-adipogenic differentiation. *Acta Biomater* 2014;10:3108-3116.
  18. Williams SM, Golden-Mason L, Ferguson BS, et al. Class I HDACs regulate angiotensin II-dependent cardiac fibrosis via fibroblasts and circulating fibrocytes. *J Mol Cell Cardiol* 2014;67:112-125.

**Cite this article as:** Cao T, Rajasingh S, Rajasingh J. Circulating fibrocytes serve as a marker for clinical diagnosis. *Ann Transl Med* 2016;4(Suppl 1):S38. doi: 10.21037/atm.2016.10.26



# Measuring soluble CD40 ligand: it is a fancy prognostic biomarker in STEMI-patients?

Alberto Dominguez-Rodriguez<sup>1,2</sup>

<sup>1</sup>Hospital Universitario de Canarias, Servicio de Cardiología, Tenerife, Spain; <sup>2</sup>Facultad de Ciencias de la Salud, Universidad Europea de Canarias, La Orotava, Santa Cruz de Tenerife, Spain

*Correspondence to:* Dr. Alberto Dominguez-Rodriguez, MD, PhD, FESC. Hospital Universitario de Canarias, Department of Cardiology, Ofra s/n La Cuesta E-38320, Tenerife, Spain. Email: adrvdg@hotmail.com.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Napoleão P, Cabral LB, Selas M, *et al.* Stratification of ST-elevation myocardial infarction patients based on soluble CD40L longitudinal changes. *Transl Res* 2016;176:95-104.

Submitted Sep 16, 2016. Accepted for publication Sep 22, 2016.

doi: 10.21037/atm.2016.10.53

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.10.53>

Acute coronary syndromes are accompanied by progressive mechanical obstruction, dynamic obstruction, and plaque inflammation, instability, and rupture, followed by superimposed thrombosis. Myocardial ischemia and necrosis are the sequelae, followed over time by remodeling ventricular. Thus, activation of select markers and enzymes during the different phases of the process can be detected in the peripheral circulation (1).

The coronary inflammation is a primary driving force for the development and progression of atherosclerosis and, thus, increased inflammation is also a common indicator of coronary atherosclerosis. An understanding of the pathobiology of atherosclerosis and the molecular events implicated in the progression from subclinical disease to overt disease has enabled the development of biomarkers to cardiovascular diseases (2,3). The vascular wall releases molecules into the bloodstream that can reflect the pathological processes taking place there. On the other hand, blood itself is clearly involved in thrombus formation. Thus, in theory, the concentrations of the molecules involved in the different pathological processes present in atherosclerosis could be biomarkers. However, not all of these molecules are suited to this aim and should fulfil certain conditions (4,5). The characteristics of an ideal biomarker are shown in *Table 1*. Although most of the biomarkers studied up to now have been based on the possibility of being useful from the diagnostic/prognostic

**Table 1** Characteristics of an ideal biomarker

Specific
Sensitive
Predictive
Fast, simple, and cheap analysis
Equal concentrations at any time of day
Samples easily acquired

standpoint, it is worth recalling that ideally they would also provide a therapeutic target. Finally, although some have no diagnostic or therapeutic value, they can provide us with information on the origin and formation of atheromatous plaque (6).

Atherosclerotic plaque instability leading to adverse events is the consequence of a complex inflammatory response of the vessel wall that involves the activation of macrophages and T cells and the production of inflammatory mediators (6). Likewise, increasing evidence suggests that CD40 ligand plays an important part in disease progression and plaque destabilization (7). The CD40-CD40 ligand system is widely distributed on a variety of leukocytic and non-leukocytic cells, including endothelial and smooth muscle cell (8), and on activated platelets (9). The CD40 ligand also occurs in a soluble form that is fully active biologically termed soluble CD40 ligand (sCD40L) (10),

which are shed from stimulated lymphocytes and are actively released after platelet stimulation

The study of Napoleão *et al.* (11) was designed to identify the groups of ST segment elevation myocardial infarction (STEMI) patients with different profiles of sCD40L concentrations and verify how medication, clinical evolution, biochemical data, and markers of regulation of endothelial function at genetic (endothelial nitric oxide synthase polymorphisms) and post-transcriptional levels (circulating microRNAs) affect sCD40L serum levels. The results of this study showed that low levels of sCD40L 1 month after percutaneous coronary intervention distinguish STEMI patients with worse prognosis, a compromised cardiac healing, and a persistent endothelial dysfunction, as given by the association between genetic and post-transcriptional markers (11).

Nonetheless, the suggestion by Napoleão *et al.* (11) to encourage further studies to evaluate the clinical role of sCD40L may be overenthusiastic. Certainly, further confirmatory data from large prospective studies are needed. In everyday clinical practice, we also need simple pragmatic and practical predictors of adverse outcomes. The cardiac troponins and brain natriuretic peptide are obvious examples that are already in use. The sceptic would therefore argue that it is not only the fancy biomarker (e.g., sCD40L) that can provide clear prognostic information for death or heart failure post-acute myocardial infarction, but even very simple biomarker, such as admission troponin levels can be of value not only in assessing prognosis for cardiac events post-acute myocardial infarction but also the response to treatment. Perhaps we need to concentrate on simple things in life rather than look for too many exotic things that may be complex and expensive to measure.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The author has no conflicts of interest to

declare.

### References

1. Voudris KV, Chanin J, Feldman DN, et al. Novel Inflammatory Biomarkers in Coronary Artery Disease: Potential Therapeutic Approaches. *Curr Med Chem* 2015;22:2680-2689.
2. Rader DJ. Inflammatory markers of coronary risk. *N Engl J Med* 2000;343:1179-1182.
3. Morrow DA, Braunwald E. Future of biomarkers in acute coronary syndromes: moving toward a multimarker strategy. *Circulation* 2003;108:250-252.
4. Domínguez-Rodríguez A, Abreu-González P. Inflammatory biomarkers in the acute coronary syndrome. *Med Clin (Barc)* 2011;136:461-462; author reply 462.
5. Martín-Ventura JL, Blanco-Colio LM, Tuñón J, et al. Biomarkers in cardiovascular medicine. *Rev Esp Cardiol* 2009;62:677-688.
6. Lubrano V, Balzan S. Consolidated and emerging inflammatory markers in coronary artery disease. *World J Exp Med* 2015;5:21-32.
7. Dominguez-Rodriguez A, Abreu-Gonzalez P, Garcia-Gonzalez MJ, et al. Diurnal variation of soluble CD40 ligand in patients with acute coronary syndrome. Soluble CD40 ligand and diurnal variation. *Thromb Res* 2009;123:617-621.
8. Schönbeck U, Libby P. The CD40/CD154 receptor/ligand dyad. *Cell Mol Life Sci* 2001;58:4-43.
9. Henn V, Slupsky JR, Gräfe M, et al. CD40 ligand on activated platelets triggers an inflammatory reaction of endothelial cells. *Nature* 1998;391:591-594.
10. Henn V, Steinbach S, Büchner K, et al. The inflammatory action of CD40 ligand (CD154) expressed on activated human platelets is temporally limited by coexpressed CD40. *Blood* 2001;98:1047-1054.
11. Napoleão P, Cabral LB, Selas M, et al. Stratification of ST-elevation myocardial infarction patients based on soluble CD40L longitudinal changes. *Transl Res* 2016;176:95-104.

**Cite this article as:** Dominguez-Rodriguez A. Measuring soluble CD40 ligand: it is a fancy prognostic biomarker in STEMI-patients? *Ann Transl Med* 2016;4(Suppl 1):S25. doi: 10.21037/atm.2016.10.53



## Corin as novel biomarker for myocardial infarction

Hans-Josef Feistritzer, Bernhard Metzler

University Clinic of Internal Medicine III, Cardiology and Angiology, Medical University of Innsbruck, A-6020 Innsbruck, Austria

*Correspondence to:* Bernhard Metzler. University Clinic of Internal Medicine III, Cardiology and Angiology, Medical University of Innsbruck, A-6020 Innsbruck, Austria. Email: Bernhard.Metzler@tirol-kliniken.at.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Zhou X, Chen J, Zhang Q, *et al.* Prognostic Value of Plasma Soluble Corin in Patients With Acute Myocardial Infarction. *J Am Coll Cardiol* 2016;67:2008-14.

Submitted Jul 06, 2016. Accepted for publication Jul 08, 2016.

doi: 10.21037/atm.2016.08.17

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.08.17>

The study by Zhou *et al.* recently published in the *J Am Coll Cardiol* provides new insights into the prognostic utility of the novel biomarker corin for risk stratification of patients with acute myocardial infarction (AMI) (1). In this prospective cohort study comprising more than 1,300 patients with ST-elevation myocardial infarction (STEMI) and non-STEMI (NSTEMI), low plasma corin concentrations were an independent predictor of major adverse cardiac events (MACE) defined as a composite endpoint of all-cause mortality, hospitalization for heart failure or recurrent AMI at a median follow-up duration of 634 days. This study impressively pointed out the incremental prognostic value of plasma corin concentrations in AMI patients beyond clinical characteristics and traditional cardiac biomarkers.

AMI is among the most frequent causes of morbidity and mortality in industrialized countries. Optimization of secondary prevention strategies is sought to be a cornerstone to improve outcome after AMI. Effective secondary prevention, again, is based on a comprehensive risk stratification of AMI patients, which preferably should be performed early after the index event. Multiple biomarkers, routinely available in clinical practice, have been linked to myocardial injury, left ventricular dysfunction and clinical outcome after AMI (2,3). However, during the last decade a number of novel biomarkers have been described, whose prognostic utility in AMI patients is either unknown or controversially discussed (4-6). As recently shown by O'Donoghue *et al.*, a multimarker model including novel biomarkers could

more precisely predict the occurrence of cardiovascular death or heart failure at 30 days after STEMI (7).

Corin, a transmembrane serine protease, is highly expressed in cardiomyocytes (8). Biologically, corin contributes to activation of natriuretic peptide precursor molecules and therefore plays a key role in the regulation of blood volume and blood pressure. Shedding of corin from the cardiomyocyte cell surface has been described and might reflect corin activity and cellular homeostasis (9). In patients after STEMI, plasma corin concentrations have been linked to cardiac troponin T levels and infarct size derived from cardiac magnetic resonance imaging (10). Although the impact of this study is limited due to the small sample size, these data suggest high plasma corin concentrations as an indicator of myocardial injury which is among the strongest predictors of poor outcome in AMI patients.

Now, in this imposing prospective study by Zhou *et al.* low plasma corin concentrations were related with poor clinical outcome at a median follow-up of approximately 600 days. These findings disagree with the hypothesis stated above, that intensified corin release might reflect myocardial injury (10). To a certain extent, the time point of corin measurement after AMI might explain these conflicting data. In the study performed by Zhou *et al.*, corin concentrations were measured out of plasma samples collected on 'admission', which represents a rather broad time frame. On the other hand, in the study showing an association between high plasma corin concentrations and infarct size, plasma samples were collected at a median of 2 days after symptom onset. Possibly, data of the latter study

reflect corin release in the setting of AMI and therefore the extent of myocardial damage. The data shown by Zhou *et al.*, however, might reflect the widely accepted long-term deleterious effect of corin deficiency on the cardiovascular system (11). For instance, low corin expression was related with myocardial fibrosis and contractile dysfunction in a murine model of dilated cardiomyopathy (12). Contrary, overexpression of corin could improve cardiac morphology and function. In terms of these data, the study by Zhou *et al.* reflects chronic adverse effects of corin deficiency on the cardiovascular system rather than an association between corin and the extent of myocardial injury. Respectively, in this study no significant association between corin and cardiac troponin levels was observed.

In sum, the association between corin and outcome following AMI appears controversial. On the one hand, myocardial injury results in increased plasma corin concentrations suggesting intensified corin release (10). The extent of myocardial necrosis, preferably assessed using cardiac magnetic resonance imaging, is among the strongest surrogate endpoints for poor outcome after AMI (13,14). On the other hand, corin overexpression goes ahead with favourable effects on the myocardium. Hence, both mechanisms, myocardial injury and cellular overexpression, might result in increased plasma corin levels. In both cases, however, the pathophysiological agent causing corin release is completely different. The study by Zhou *et al.* suggests a predominant adverse effect of corin deficiency on prognosis after AMI. However, this study might underestimate the prognostic role of myocardial injury, particularly as cardiac troponin levels were not related with poor outcome in this study. Necessarily, larger studies are needed to further elucidate the relation of myocardial injury and corin release in the setting of AMI.

Finally, although an independent prognostic role of plasma soluble corin concentrations for poor outcome has been demonstrated in the study by Zhou *et al.*, different pathophysiological mechanisms might determine plasma corin concentrations after AMI. Studies comprising corin concentrations together with established cardiac biomarkers and imaging parameters for myocardial injury, structure and function are needed to answer this open question.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Zhou X, Chen J, Zhang Q, et al. Prognostic Value of Plasma Soluble Corin in Patients With Acute Myocardial Infarction. *J Am Coll Cardiol* 2016;67:2008-2014.
2. Klug G, Mayr A, Mair J, et al. Role of biomarkers in assessment of early infarct size after successful p-PCI for STEMI. *Clin Res Cardiol* 2011;100:501-510.
3. Reinstadler SJ, Feistritz HJ, Klug G, et al. High-sensitivity troponin T for prediction of left ventricular function and infarct size one year following ST-elevation myocardial infarction. *Int J Cardiol* 2016;202:188-193.
4. Feistritz HJ, Klug G, Reinstadler SJ, et al. Fetuin-A is related to infarct size, left ventricular function and remodelling after acute STEMI. *Open Heart* 2015;2:e000244.
5. Feistritz HJ, Klug G, Reinstadler SJ, et al. Novel biomarkers predicting cardiac function after acute myocardial infarction. *Br Med Bull* 2016. [Epub ahead of print].
6. Mayr A, Klug G, Mair J, et al. Galectin-3: relation to infarct scar and left ventricular function after myocardial infarction. *Int J Cardiol* 2013;163:335-337.
7. O'Donoghue ML, Morrow DA, Cannon CP, et al. Multimarker Risk Stratification in Patients With Acute Myocardial Infarction. *J Am Heart Assoc* 2016;5:e002586.
8. Yan W, Sheng N, Seto M, et al. Corin, a mosaic transmembrane serine protease encoded by a novel cDNA from human heart. *J Biol Chem* 1999;274:14926-14935.
9. Jiang J, Wu S, Wang W, et al. Ectodomain shedding and autocleavage of the cardiac membrane protease corin. *J Biol Chem* 2011;286:10066-10072.
10. Feistritz HJ, Klug G, Reinstadler SJ, et al. Circulating corin concentrations are related to infarct size in patients after ST-segment elevation myocardial infarction. *Int J Cardiol* 2015;192:22-23.
11. Dong N, Chen S, Yang J, et al. Plasma soluble corin in patients with heart failure. *Circ Heart Fail* 2010;3:207-211.
12. Gladysheva IP, Wang D, McNamee RA, et al. Corin overexpression improves cardiac function, heart failure, and survival in mice with dilated cardiomyopathy. *Hypertension* 2013;61:327-332.

13. Eitel I, de Waha S, Wöhrle J, et al. Comprehensive prognosis assessment by CMR imaging after ST-segment elevation myocardial infarction. *J Am Coll Cardiol* 2014;64:1217-1226.
14. Klug G, Mayr A, Schenk S, et al. Prognostic value at 5 years of microvascular obstruction after acute myocardial infarction assessed by cardiovascular magnetic resonance. *J Cardiovasc Magn Reson* 2012;14:46.

**Cite this article as:** Feistritzer HJ, Metzler B. Corin as novel biomarker for myocardial infarction. *Ann Transl Med* 2016;4(20):405. doi: 10.21037/atm.2016.08.17

# miRNA-197 and miRNA-223 and cardiovascular death in coronary artery disease patients

Esteban Orenes-Piñero<sup>1</sup>, Francisco Marín<sup>2</sup>, Gregory Y. H. Lip<sup>3</sup>

<sup>1</sup>Proteomic Unit, Instituto Murciano de Investigación Biosanitaria Virgen de la Arrixaca (IMIB-Arrixaca), Universidad de Murcia, Murcia, Spain; <sup>2</sup>Department of Cardiology, Hospital Clínico Universitario Virgen de la Arrixaca (IMIB-Arrixaca), Universidad de Murcia, Murcia, Spain;

<sup>3</sup>University of Birmingham Institute of Cardiovascular Sciences, City Hospital, Birmingham, UK

*Correspondence to:* Prof. Gregory Y. H. Lip, MD. University of Birmingham Centre for Cardiovascular Sciences, City Hospital, Birmingham B18 7QH, UK. Email: g.y.h.lip@bham.ac.uk.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

Submitted Apr 03, 2016. Accepted for publication Apr 10, 2016.

doi: 10.21037/atm.2016.05.27

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.05.27>

Coronary artery disease (CAD) is commonly associated with the presence of atheromatous plaques in the coronary arteries. The growth of these plaques may cause arterial stenosis and blockage of the blood flow, leading to cardiac ischemia and clinical symptoms, such as acute coronary syndrome (ACS) or stable angina pectoris (SAP). Due to the high morbidity and mortality rate, CAD has probably the most serious cardiovascular disorder threatening people's health in Western countries (1).

It is widely accepted that the erosion of vulnerable plaques results in the formation of luminal thrombi secondary to platelet activation and the release of thrombogenic elements within the atherosclerotic lesions. Indeed, coagulation components and platelet activation play a major role in the development and outcome of coronary atherosclerosis.

MicroRNAs (miRNAs) are endogenous, conserved, single stranded, small (approximately 22 nucleotides in length), non-coding RNAs that repress gene expression at the post-transcriptional level by targeting mRNA (2). According to the miRNA database (miRBase), the human genome encodes 2,588 mature miRNA sequences, which may target more than 60% of human protein-coding genes. miRNA anneals to complementary sequences in the 3'-untranslated regions (3'UTR) of target mRNAs of protein-coding genes, causing mRNA to be cleaved or to repress the translational machinery needed for protein synthesis. Thus, miRNA can either inhibit translation or induce degradation of its

target mRNA or both, depending upon the overall degree of complementarity of the binding site, the number of binding sites, and the accessibility of those binding sites (3). The stronger its complementarity with the prospective target RNA, the more likely that the miRNA will degrade the target mRNA, and those miRNAs that display imperfect sequence complementarities with target mRNAs primarily, result in translational inhibition (4,5).

Accumulating studies reveal the importance of miRNAs in regulating key signaling and lipid homeostasis pathways that alter the balance of atherosclerotic plaque progression and regression. Several miRNAs have been associated with cholesterol homeostasis by production and clearance of lipoproteins that deliver [low-density lipoprotein (LDL)] and remove [high-density lipoprotein (HDL)] cholesterol from cells and tissues. Thus, miR-148a, miR-128-1, miR-130b and miR-301b have been identified as negative regulators of LDL receptor expression and activity, promoting the clearance of circulating LDL particles (6,7). On the other hand, miRNAs have also been identified to act as critical regulators of HDL biogenesis. Many miRNAs have been identified that target ATP-binding cassette transporter-A1 (ABCA1) to reduce cholesterol efflux to apolipoprotein-A1 *in vitro*, including miR-33, miR-758, miR-26, miR-106, miR-144, as well as the above-mentioned miR-128-1 and miR-148a (8-10).

Importantly, miRNAs are not only associated with lipoprotein metabolism but they are also implied in the

regulation of endothelial cell inflammation and plaque progression. For example, several studies highlight that miR-181b and miR-146a regulate distinct components of NF- $\kappa$ B signaling being atheroprotective (11). Moreover, miRNAs also regulate leukocyte recruitment and activation in atherosclerosis, one of the earliest pathogenic events in atherosclerosis. A growing list of miRNAs are implicated in regulating the activation of leukocytes, including miR-let7a, miR-19a, miR-21, miR-27a, miR-33, miR-124, miR-125a, miR-146a, miR-155, miR-214, and miR-223 (12).

All these observations point out the importance of miRNAs as potential biomarkers of atherosclerosis progression and consequently, with CAD. Indeed, a number of studies have analyzed the profiling of specific miRNAs as diagnostic markers and as predictors of future cardiovascular events in CAD patients. For example, Schulte *et al.* (13) reported the capacity of miRNA-197 and miRNA-223 in predicting cardiovascular death and burden of future cardiovascular events in a large cohort of CAD patients. In this study, 873 consecutive patients [38.9% (n=340) cases of ACS and 61.1% (n=533) cases of SAP] were included in the miRNA quantification analyses after RNA isolation, and cardiovascular death was observed in 2.1% (n=18) of the patients over a median follow-up time of 4 years (IQR, 2.78–5.04). Cox regression analysis adjusted for age and gender revealed relevant prognostic power of miR-197 and miR-223 with respect to the primary end point cardiovascular death in the overall group [miRNA-197: HR =1.77 per one SD increase (95% CI, 1.20–2.60), P=0.004, C-index =0.78; miRNA-223: HR =2.23 per one SD increase (95% CI, 1.20–4.14), P=0.011, C-index =0.80]. In addition, subgroup analysis for ACS patients revealed a stronger association between elevated levels of miR-197 and miR-223 and future cardiovascular death [miRNA-197: HR =2.24 per one SD increase (95% CI, 1.25–4.01), P=0.006, C-index =0.89; miRNA-223: HR =4.94 per one SD increase (95% CI, 1.42–17.20), P=0.012, C-index =0.89].

Nonetheless, the rather small number of cardiovascular death endpoints may limit the validation of the observed findings in this study. This may influence the statistical ability to detect small effects and contains a risk of statistically overfitting the results, especially with respect to the subgroups ACS and SAP. We also need to see more characteristics of the cardiovascular death patients, as this is the primary endpoint of the AtheroGene study. Such information, together with a multivariate analysis may help the readers to clarify the role of miR-197 and miR-223 for the prediction of cardiovascular deaths in this cohort. We

should stress the importance of new statistical approaches for giving us the additional information of these biomarkers in clinical practice (14).

Finally, some biomarkers for predicting cardiovascular events or deaths in community-based populations have not consistently added information to standard risk factors. Although the use of circulating biomarkers to aid risk prediction is attractive, prior studies have not consistently demonstrated the value of biomarkers for prognosis or diagnosis beyond standard risk factors in low to intermediate risk individuals in different cohorts.

### Acknowledgements

Dr. Orenes-Piñero is supported by a postdoctoral contract from Instituto Murciano de Investigación Biosanitaria Virgen de la Arrixaca (IMIB-Arrixaca), Murcia, Spain.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Piccolo R, Giustino G, Mehran R, et al. Stable coronary artery disease: revascularisation and invasive strategies. *Lancet* 2015;386:702-713.
2. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281-297.
3. Agarwal V, Bell GW, Nam JW, et al. Predicting effective microRNA target sites in mammalian mRNAs. *Elife* 2015.4.
4. Orenes-Piñero E, Montoro-García S, Patel JV, et al. Role of microRNAs in cardiac remodelling: new insights and future perspectives. *Int J Cardiol* 2013;167:1651-1659.
5. Navas-Carrillo D, Ríos A, Rodríguez JM, et al. Familial nonmedullary thyroid cancer: screening, clinical, molecular and genetic findings. *Biochim Biophys Acta* 2014;1846:468-76.
6. Goedeke L, Rotllan N, Canfrán-Duque A, et al. MicroRNA-148a regulates LDL receptor and ABCA1 expression to control circulating lipoprotein levels. *Nat Med* 2015;21:1280-1289.
7. Wagschal A, Najafi-Shoushtari SH, Wang L, et al. Genome-wide identification of microRNAs regulating cholesterol and triglyceride homeostasis. *Nat Med* 2015;21:1290-1297.

8. Rayner KJ, Esau CC, Hussain FN, et al. Inhibition of miR-33a/b in non-human primates raises plasma HDL and lowers VLDL triglycerides. *Nature* 2011;478:404-407.
9. Ramirez CM, Dávalos A, Goedeke L, et al. MicroRNA-758 regulates cholesterol efflux through posttranscriptional repression of ATP-binding cassette transporter A1. *Arterioscler Thromb Vasc Biol* 2011;31:2707-2714.
10. Sun D, Zhang J, Xie J, et al. MiR-26 controls LXR-dependent cholesterol efflux by targeting ABCA1 and ARL7. *FEBS Lett* 2012;586:1472-1479.
11. Sun X, Belkin N, Feinberg MW. Endothelial microRNAs and atherosclerosis. *Curr Atheroscler Rep* 2013;15:372.
12. Feinberg MW, Moore KJ. MicroRNA Regulation of Atherosclerosis. *Circ Res* 2016;118:703-720.
13. Schulte C, Molz S, Appelbaum S, et al. miRNA-197 and miRNA-223 predict cardiovascular death in a cohort of patients with symptomatic coronary artery disease. *PLoS One* 2015;10:e0145930.
14. Pencina MJ, D'Agostino RB Sr, D'Agostino RB Jr, et al. Evaluating the added predictive ability of a new marker: from area under the ROC curve to reclassification and beyond. *Stat Med* 2008;27:157-172; discussion 207-212.

**Cite this article as:** Orenes-Piñero E, Marín F, Lip GY. miRNA-197 and miRNA-223 and cardiovascular death in coronary artery disease patients. *Ann Transl Med* 2016;4(10):200. doi: 10.21037/atm.2016.05.27



## miR-126: a potential new key player in hypoxia and reperfusion?

Sabina P. W. Guenther<sup>1,2,3,4,5</sup>, Sonja Schrepfer<sup>1,2,3,4</sup>

<sup>1</sup>Transplant and Stem Cell Immunobiology Laboratory, Department of Surgery, University of California, San Francisco, CA, USA; <sup>2</sup>Transplant and Stem Cell Immunobiology Laboratory, University Heart Centre Hamburg, University of Hamburg, Cardiovascular Research, Hamburg, Germany; <sup>3</sup>Cardiovascular Research Center (CVRC), University Medical Center Hamburg-Eppendorf, Hamburg, Germany; <sup>4</sup>German Centre for Cardiovascular Research (DZHK) e.V., University Medical Center Hamburg-Eppendorf, Hamburg, Germany; <sup>5</sup>Department of Cardiac Surgery, University Hospital Munich, Ludwig-Maximilian-University, Munich, Germany

*Correspondence to:* Sonja Schrepfer, MD, PhD. Transplant and Stem Cell Immunobiology Laboratory, Department of Surgery, University of California San Francisco (UCSF), Medical Sciences S1207, 513 Parnassus Avenue, San Francisco, CA 94143-2205, USA. Email: sonja.schrepfer@ucsf.edu.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Li B, Tao Y, Huang Q. Effect and mechanism of miR-126 in myocardial ischemia reperfusion. *Genet Mol Res* 2015;14:18990-8.

Submitted Jun 28, 2016. Accepted for publication Jun 29, 2016.

doi: 10.21037/atm.2016.08.22

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.08.22>

It is well known that microRNAs (miRNAs) have recently emerged as multifaceted regulators of biological processes and thus of various diseases. miRNAs are small single-stranded non-coding RNAs that bind to their cognate messenger RNAs (mRNAs) via recognition of seed sequences, i.e., the 2<sup>nd</sup>–8<sup>th</sup> nucleotide of the miRNA. This binding can trigger mRNA degradation and translational repression leading to a decrease in the protein products encoded by the corresponding mRNA. In contrast to profound effects of conventional transcription factors, individual miRNAs are rather considered as nuancing regulators of particular mRNA expression. However, since a single miRNA may act on different mRNAs and vice versa single mRNAs can be regulated by various miRNAs and additionally since miRNAs may have multiple target sites within one mRNA, cumulating effects result and complex regulatory networks are created (1).

Besides regulating translational control, circulating miRNAs act as endocrine signalling molecules and may furthermore serve as diagnostic disease markers (2). Conversely, targeting miRNA pathways offers novel therapeutic options (1).

Cardiovascular diseases remain to be among the leading causes of morbidity and mortality worldwide and impose a relevant economic burden on the health care systems. Multiple recent studies suggested a relevant role of miRNAs in cardiovascular processes and diseases.

Initially, miR-126 was demonstrated to have significant impact on tumor development and metastasis. Amongst others, it can influence inflammation, proliferation and plays a role in tumor-angiogenesis (3,4). Specifically, it seems to serve as a tumor and metastasis suppressor (3,4). In colon cancer, its expression is significantly lower especially in highly metastatic colon cancer cells. It reduces cancer cell viability and migration as well as invasion via downregulation of CXCR4 expression (3). Comparable findings have also been reported for other malignant cell lines, for example in non-small cell lung cancer via Crk (5).

More recently, a growing body of scientific findings points out the role of miR-126 in cardiovascular diseases, paralleling the earlier findings in cancer research. Endothelial cells are essential for maintaining vascular integrity, play a major role in angiogenesis and in the response to ischemia or injury (6). Vascular remodelling can be both, beneficial with repair and adaption after injury and ischemia or deleterious as for example in atherogenesis. In this context, a single miRNA can regulate multiple processes and thus affect both, positive and negative vascular remodelling (7). Due to overlappings, therapeutic targeting of miRNAs may thus influence several mechanisms in remodelling and result in unexpected and unwanted additional effects (7).

The miR-126 gene is located within an intron on the epidermal growth factor-like-domain 7 (*EGFL7*) gene

on chromosome 9 and gives rise to two mature miRNAs: miR-126-3p and miR-126-5p (6,7). miR-126 is highly expressed in the vascular endothelium and exerts distinct yet dichotomous effects in the embryonic, healthy adult and diseased adult vascular system (8). Opposing to the induction of angiogenic signalling and promotion of endothelial cell differentiation and maturation in embryonic vasculogenesis, it preserves vascular homeostasis and integrity via inhibition of angiogenesis and endothelial cell proliferation maintaining a quiescent phenotype in the mature state (6,8-10). Besides, it is a key regulator of inflammation, which is a major contributor to vascular pathology including endothelial dysfunction, remodelling and atherosclerosis, and has effects on cells of the hematopoietic system (9,11,12). Confirmed targets include VCAM1, SPRED1 and DLK1 (7). Interestingly, the blood flow pattern and thus intravascular shear stress seem to contribute to the different effects exerted by miR-126 on endothelial cells and the vascular system (12). In the context of vessel injury and also in hypoxia, miR-126 does have differential effects. Van Solingen *et al.* showed that antagomir silencing of miR-126 did not have any effect on the proliferation and migration of HUVECs *in vitro*. Likewise, in a hindlimb ischemia model in mice induced by electrocoagulation of the femoral artery 24 h after injection of antagomir-126, no differences in blood flow recovery were seen (6). However, quantitative analysis of capillaries in the calf muscle showed a significantly lower density of capillaries in mice treated with high-dose antagomir-126. Even though miR-126 is supposed to target VCAM1 and downregulation of miR-126 might result in increased leukocyte adherence, the latter findings suggest that while not directly or at least not rate-limitingly affecting arteriogenesis, miR-126 exhibits a distinct effect on the ischemia-induced angiogenic response. In line with these findings, silencing of miR-126 impaired endothelial cell outgrowth in aortic explant cultures. A suggested mechanism for promoting angiogenesis in this setting includes a reduced expression of repressors of VEGF signalling by overexpressed miR-126 (6).

In humans, miR-126-3p was significantly downregulated in patients with acute myocardial infarction and it was suggested that administration of miR-126 could rescue endothelial cell function whereas the authors did not state the exact time of blood analysis, i.e., before or after revascularization (2).

In another analysis, an altered expression of miRNAs in patients with chronic total coronary artery occlusion and insufficient collateral arteries was recently demonstrated,

with a significantly elevated level of miR-126, miR-423-5p, miR-30d and miR-10b. Even though probably suitable as biomarkers, it remains to be elucidated, however, whether these miRNAs are upregulated due to direct effects on collateral artery development or whether they are part of other pathways that affect collateral vessel growth (13). Schober *et al.* demonstrated that miR-126-5p promotes endothelial proliferation and limits atherosclerosis by suppressing Dlk1 (12).

The recent *Genet Mol Res* article by Li *et al.* adds to the growing body of scientific findings the role of miR-126 in myocardial ischemia reperfusion injury. Revascularization is the gold standard for the treatment of myocardial infarction. Reviewing the current literature, Li *et al.* explain that while the exact underlying pathophysiology still needs to be elucidated injury resulting from reperfusion is a significant cause of cardiac cell death after ischemia and accounts for a relevant portion of the necrosis. Thus not only ischemia itself but also sudden blood flow restoration has a harmful effect on cell survival and apoptosis (14,15). In order to assess the role of miR-126 in reperfusion injury *in vitro*, miR-126 inhibitor and mimic were transfected into rat myocardial H9c2 cells and the cells were subjected to simulated ischemia-reperfusion injury afterwards. miR-126 as assessed by real-time PCR was significantly downregulated after ischemia-reperfusion. The miR-126 inhibitor reduced injury-induced myocardial cell apoptosis and caspase 3 protein expression. For the mimic contrary effects were observed. For *in vivo* analysis, female Wistar rats underwent lentivirus miR-126 mimic or inhibitor injection and 7 days later 30 min of myocardial ischemia induced by ligation of the left anterior descending coronary artery followed by 2 h of reperfusion. Infarction area size was significantly smaller in miR-126 inhibitor-injected animals and larger in mimic-injected animals in comparison to the control group, respectively (14). Obviously, miR-126 affects regulation of myocardial cell apoptosis in ischemia-reperfusion injury.

Contrastingly, in other investigations hypoxia itself resulted in a profound up-regulation of endothelial miR-126 (8). Furthermore, miR-126 upregulation was shown to have the capability of re-inducing angiogenesis, promoting re-endothelialization and of re-activating endothelial (progenitor) cells whilst inhibiting apoptosis and thus to contribute to healing mechanisms and to exhibit vasculoprotective effects (8). Thus, there obviously are substantial differences in the regulatory effects during hypoxia and reperfusion as well as varying effects on the



vascular system and cardiomyocytes including the involved downstream targets and mediating components. A major contributing factor in this setting might be actual blood flow and thus shear stress within the vessel. Additionally, a key role of heat shock protein 70 in influencing myocardial cell apoptosis has been suggested (14,16).

As explained earlier, miRNAs have been considered as sensitive diagnostic and prognostic biomarkers. Considering its pathophysiological implications, miR-126 may have a potential value in diagnosing not only myocardial ischemia, acute myocardial infarction, but also in stroke and other acute or chronic ischemic diseases (2,8). Additionally, the abovementioned findings underline the powerful therapeutic implications of targeting miRNAs in cardiovascular diseases. Further elucidating both conditions, hypoxia and reperfusion, could thus substantially contribute to the development of novel therapeutic agents.

### Acknowledgements

SP Guenther was supported by a grant from the German Heart Foundation. S Schrepfer was funded by the German Research Foundation [Deutsche Forschungsgemeinschaft (DFG), SCHR992/3-1 and SCHR992/4-1].

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Rottiers V, Näär AM. MicroRNAs in metabolism and metabolic disorders. *Nat Rev Mol Cell Biol* 2012;13:239-250.
2. Hsu A, Chen SJ, Chang YS, et al. Systemic approach to identify serum microRNAs as potential biomarkers for acute myocardial infarction. *Biomed Res Int* 2014;2014:418628.
3. Li Z, Li N, Wu M, et al. Expression of miR-126 suppresses migration and invasion of colon cancer cells by targeting CXCR4. *Mol Cell Biochem* 2013;381:233-242.
4. Banerjee N, Kim H, Talcott S, et al. Pomegranate polyphenolics suppressed azoxymethane-induced colorectal aberrant crypt foci and inflammation: possible role of miR-126/VCAM-1 and miR-126/PI3K/AKT/mTOR. *Carcinogenesis* 2013;34:2814-2822.
5. Crawford M, Brawner E, Batte K, et al. MicroRNA-126 inhibits invasion in non-small cell lung carcinoma cell lines. *Biochem Biophys Res Commun* 2008;373:607-612.
6. van Solingen C, Seghers L, Bijkerk R, et al. Antagomir-mediated silencing of endothelial cell specific microRNA-126 impairs ischemia-induced angiogenesis. *J Cell Mol Med* 2009;13:1577-1585.
7. Welten SM, Goossens EA, Quax PH, et al. The multifactorial nature of microRNAs in vascular remodelling. *Cardiovasc Res* 2016;110:6-22.
8. Chistiakov DA, Orekhov AN, Bobryshev YV. The role of miR-126 in embryonic angiogenesis, adult vascular homeostasis, and vascular repair and its alterations in atherosclerotic disease. *J Mol Cell Cardiol* 2016;97:47-55.
9. van Solingen C, Bijkerk R, de Boer HC, et al. The Role of microRNA-126 in vascular homeostasis. *Curr Vasc Pharmacol* 2015;13:341-351.
10. Fish JE, Santoro MM, Morton SU, et al. miR-126 regulates angiogenic signaling and vascular integrity. *Dev Cell* 2008;15:272-284.
11. Renna NF, de Las Heras N, Miatello RM. Pathophysiology of vascular remodeling in hypertension. *Int J Hypertens* 2013;2013:808353.
12. Schober A, Nazari-Jahantigh M, Wei Y, et al. MicroRNA-126-5p promotes endothelial proliferation and limits atherosclerosis by suppressing Dlk1. *Nat Med* 2014;20:368-376.
13. Hakimzadeh N, Nossent AY, van der Laan AM, et al. Circulating microRNAs characterizing patients with insufficient coronary collateral artery function. *PLoS One* 2015;10:e0137035.
14. Li B, Tao Y, Huang Q. Effect and mechanism of miR-126 in myocardial ischemia reperfusion. *Genet Mol Res* 2015;14:18990-18998.
15. Qiao X, Xu J, Yang QJ, et al. Transient acidosis during early reperfusion attenuates myocardium ischemia reperfusion injury via PI3k-Akt-eNOS signaling pathway. *Oxid Med Cell Longev* 2013;2013:126083.
16. Jiang C, Ji N, Luo G, et al. The effects and mechanism of miR-92a and miR-126 on myocardial apoptosis in mouse ischemia-reperfusion model. *Cell Biochem Biophys* 2014;70:1901-1906.

**Cite this article as:** Guenther SP, Schrepfer S. miR-126: a potential new key player in hypoxia and reperfusion? *Ann Transl Med* 2016;4(19):377. doi: 10.21037/atm.2016.08.22

## miR-21 alters circulating Treg function in vascular disease—hope for restoring immunoregulatory responses in atherosclerosis?

Emer E. Hackett, Frederick J. Sheedy

School of Biochemistry & Immunology, Trinity Biomedical Sciences Institute, Trinity College, Dublin, Ireland

*Correspondence to:* Frederick J. Sheedy, School of Biochemistry & Immunology, Trinity Biomedical Sciences Institute, Trinity College, Dublin, Ireland. Email: fsheedy@tcd.ie.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Li S, Fan Q, He S, *et al.* MicroRNA-21 negatively regulates Treg cells through a TGF- $\beta$ 1/Smad-independent pathway in patients with coronary heart disease. *Cell Physiol Biochem* 2015;37:866-78.

Submitted Nov 21, 2016. Accepted for publication Nov 25, 2016.

doi: 10.21037/atm.2016.12.72

View this article at: <http://dx.doi.org/10.21037/atm.2016.12.72>

The small molecule regulators of gene expression, microRNAs, have emerged as important mediators in a variety of cellular processes linked to disease. Much emphasis has been placed on measuring the expression of these molecules as novel biomarkers of disease. In particular, miRNA profiles of cancer have revealed both tumor specific signatures as well as highlighting common miRNAs with central roles in malignancy. Chief amongst these is microRNA-21, miR-21, the expression of which is enhanced in multiple solid tumors and lymphomas (1) where it regulates transformation by limiting the expression of various tumor suppressor genes (2). Outside of tumors, miR-21 is also highly expressed in cells of the immune system and its expression can be modulated by inflammatory stimuli during immune cell activation (3), which traffic through the circulation. miRNAs can also be found in circulation in secreted vesicles including exosomes released from various cell types associated with disease (4). This has led to an explosion in interest in measuring serum miRNA profiles during disease pathogenesis. A recently published article in *Cellular Physiology and Biochemistry* by Li and colleagues (5) measured miR-21 in serum from patients with vascular disease has revealed novel ways miR-21 can control circulating immune cell diversity and function, which impacts upon disease.

Although other groups measured circulating miR-21 in patients with atherosclerosis previously and found an increased level of miR-21 associated with increased risk of stroke (6), the study by Li and colleagues examines miR-21 in

serum in various cohorts of patients with increasing severity. In particular, miR-21 expression is increased over 5-fold in peripheral blood mononuclear cells (PBMCs—a fraction of white blood cells including B, T-cells, monocytes, dendritic cells and neutrophils), in patients with severe vascular disease and a history of myocardial infarction relative to control patients with chest pains yet no vascular disease. Interestingly, in other intermediate patients groups with progressing from stable to unstable angina and increasing incidence of vascular disease there is also an increase in PBMC miR-21 expression implicating the induction of miR-21 in progression of atherosclerosis.

While atherosclerosis is associated with the recruitment of immune cells to areas of lipid deposition along blood vessels, particularly pro-inflammatory monocytes which foster and promote plaque inflammation (7), it is emerging that regulatory immune cells including FoxP3<sup>+</sup> T-cells (Treg) infiltrate atherosclerotic plaques (8). However, the number and function of these cells decreases both in plaques and in circulation as disease progresses, consistent with a breakdown in tolerance and the appearance of pro-inflammatory T cells (9-12). The Li study confirmed this decline in circulating Treg numbers alongside the expression levels of TGF- $\beta$  and FoxP3 mRNA in PBMC from atherosclerotic patients, reflected in the serum by decreased TGF- $\beta$  protein levels when measured by ELISA. These decreases, like miR-21, are associated with severity of vascular disease and are more pronounced in cohorts with history of unstable angina and myocardial infarction. These

intriguing findings suggest that an increase in PBMC miR-21 as disease progresses alters T-cell function to promote an immune-regulatory environment, specifically by reducing Treg numbers and activity.

Previous work in the area of T-cell biology has implicated miR-21 in multiple levels of control of T-cell fate, function and diversity and this new data showing modulation of miR-21 negatively regulating T-reg function in atherosclerosis, while increasing our knowledge also increases the complexity whereby miR-21 can control immune function. miR-21 is known to be specifically expressed in the Treg subset (13) and promotes Treg differentiation by positively regulating expression of FoxP3 itself, the Treg-specific transcription factor. miR-21 also has the potential to positively regulate Treg activity by directly targeting a proposed negative regulator of TGF- $\beta$  signaling, SMAD7 (14,15). In contrast, other studies have suggested that miR-21 in fact restrains Treg activity through intrinsic T-reg-specific mechanisms and limiting FOXP3 activity (16) or indirectly by promoting the activity of Th17 cells, whose pro-inflammatory nature counters Treg function (14). With this in mind, Li and colleagues found that the decrease in TGF- $\beta$  and FoxP3 expression in PBMC from diseased patients was consistent with a decrease in mRNA expression of the miR-21 target *SMAD7*. While this suggests the increase in PBMC miR-21 has functional consequences by reducing the expression of a known miR-21 target, it also suggests the net anti-inflammatory effects of this on T-reg numbers and activity may proceed through an alternative mechanism.

Although it is likely that miR-21 controls Treg activity through repression of alternative target mRNAs, it is also possible that during atherosclerosis progression, the increase in PBMC miR-21 which alters Treg activity, may occur in other cell types which drive suppression of Tregs. In particular, the role of inflammatory monocytes, known to increase during hyperlipidemia (17), or Th17 cells, known to negatively regulate Treg activity through miR-21 in other contexts (14), could mediate the effects here. Indeed, the cells expressing induced-miR-21 in disease remain unidentified and although Treg numbers decrease with vascular disease, the possibility exists that the remaining cells enhance miR-21 to mediate suppressive effects. Although the current study did not examine miR-21 in Treg in atherosclerotic plaques, miR-21 has been reported to be up-regulated in human plaques (18) and differences in local and peripheral regulatory T-cell function may exist. The authors highlight a previous study which demonstrated

that Tregs derived from umbilical cord blood (13), which represented a heterogenous population distinct from those of circulating PBMC, also express miR-21 to promote Treg activity through FoxP3.

Examining other autoimmune diseases reveals interesting differences relating to circulating miR-21 levels and immune cell function. For example, rheumatoid arthritis patients, characterized by chronic inflammation in synovial joints, display decreased serum miR-21 levels (15). This is associated with decreased Treg function and an increase in the number and activity of the Th17 subset, promoting chronic inflammation. In this particular situation, loss of miR-21 allowed expression of another predicted target, *STAT3*, which acts as a Th17-specific transcription factor. Again, the cells in which miR-21 expression is lost in during disease are not clear, although here it is likely that Th17 polarization is promoted by an increase in *STAT3* expression in T-cell precursors. This has the net effect of limiting Treg activity. In another model of autoimmune disease, the EAE model of multiple sclerosis, targeting miR-21 using antisense was shown to block Th17 mediated inflammation leading to reduced disease burden (14). In mouse models of SLE, both targeting of miR-21 by antisense and miR-21 deletion decreased Th17 cells with a consequent increase in Tregs, conferring protection against disease (19,20). These results confirm that the miR-21/Treg axis is active in disease and confirmation of the results in the Li *et al.* study using similar tools for vascular disease are forthcoming. In particular, identifying the mRNA targets specific to each disease through which miR-21 has its effects on immune cell function will add to the complexity behind control of T-cell fate by this important miRNA and will allow more specific targeting for improved therapies.

In summary, the study by Li *et al.* provides primary patient data highlighting an important role for miR-21 in the pathogenesis of vascular disease. Notably, as disease progresses there is a loss of an important immunoregulatory T-cell subset in circulation which is driven by miR-21 and identifying the cellular and molecular mechanisms controlling this will illuminate our knowledge of T-cell function, tolerance and inflammation in atherosclerosis. In particular, identifying the signals, such as lipids or lipid-induced inflammatory mediators, which drive miR-21 expression during disease progression as well as identifying the overexpressing subsets in PBMCs and correlating these observed differences with local immune cells in the plaque, will improve our understanding of miR-21 function in immunity.

## Acknowledgements

None.

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

- Volinia S, Calin GA, Liu CG, et al. A microRNA expression signature of human solid tumors defines cancer gene targets. *Proc Natl Acad Sci U S A* 2006;103:2257-2261.
- Krichevsky AM, Gabriely G. miR-21: a small multi-faceted RNA. *J Cell Mol Med* 2009;13:39-53.
- Sheedy FJ. Turning 21: Induction of miR-21 as a Key Switch in the Inflammatory Response. *Front Immunol* 2015;6:19.
- D'Souza-Schorey C, Clancy JW. Tumor-derived microvesicles: shedding light on novel microenvironment modulators and prospective cancer biomarkers. *Genes Dev* 2012;26:1287-1299.
- Li S, Fan Q, He S, et al. MicroRNA-21 negatively regulates Treg cells through a TGF- $\beta$ 1/Smad-independent pathway in patients with coronary heart disease. *Cell Physiol Biochem* 2015;37:866-878.
- Tsai PC, Liao YC, Wang YS, et al. Serum microRNA-21 and microRNA-221 as potential biomarkers for cerebrovascular disease. *J Vasc Res* 2013;50:346-354.
- Moore KJ, Sheedy FJ, Fisher EA. Macrophages in atherosclerosis: a dynamic balance. *Nat Rev Immunol* 2013;13:709-721.
- de Boer OJ, van der Meer JJ, Teeling P, et al. Low numbers of FOXP3 positive regulatory T cells are present in all developmental stages of human atherosclerotic lesions. *PLoS One* 2007;2:e779.
- Hansson GK, Hermansson A. The immune system in atherosclerosis. *Nat Immunol* 2011;12:204-212.
- Mor A, Planer D, Luboshits G, et al. Role of naturally occurring CD4<sup>+</sup> CD25<sup>+</sup> regulatory T cells in experimental atherosclerosis. *Arterioscler Thromb Vasc Biol* 2007;27:893-900.
- Rohm I, Atiskova Y, Drobnik S, et al. Decreased regulatory T cells in vulnerable atherosclerotic lesions: imbalance between pro- and anti-inflammatory cells in atherosclerosis. *Mediators Inflamm* 2015;2015:364710.
- Liu ZD, Wang L, Lu FH, et al. Increased Th17 cell frequency concomitant with decreased Foxp3<sup>+</sup> Treg cell frequency in the peripheral circulation of patients with carotid artery plaques. *Inflamm Res* 2012;61:1155-1165.
- Rouas R, Fayyad-Kazan H, El Zein N, et al. Human natural Treg microRNA signature: role of microRNA-31 and microRNA-21 in FOXP3 expression. *Eur J Immunol* 2009;39:1608-1618.
- Murugaiyan G, da Cunha AP, Ajay AK, et al. MicroRNA-21 promotes Th17 differentiation and mediates experimental autoimmune encephalomyelitis. *J Clin Invest* 2015;125:1069-1080.
- Dong L, Wang X, Tan J, et al. Decreased expression of microRNA-21 correlates with the imbalance of Th17 and Treg cells in patients with rheumatoid arthritis. *J Cell Mol Med* 2014;18:2213-2224.
- Bhairavabhotla R, Kim YC, Glass DD, et al. Transcriptome profiling of human FoxP3<sup>+</sup> regulatory T cells. *Hum Immunol* 2016;77:201-213.
- Swirski FK, Libby P, Aikawa E, et al. Ly-6Chi monocytes dominate hypercholesterolemia-associated monocytosis and give rise to macrophages in atheromata. *J Clin Invest* 2007;117:195-205.
- Raitoharju E, Lyytikäinen LP, Levula M, et al. miR-21, miR-210, miR-34a, and miR-146a/b are up-regulated in human atherosclerotic plaques in the Tampere Vascular Study. *Atherosclerosis* 2011;219:211-217.
- Garchow BG, Bartulos Encinas O, Leung YT, et al. Silencing of microRNA-21 in vivo ameliorates autoimmune splenomegaly in lupus mice. *EMBO Mol Med* 2011;3:605-615.
- Garchow B, Kiriakidou M. MicroRNA-21 deficiency protects from lupus-like autoimmunity in the chronic graft-versus-host disease model of systemic lupus erythematosus. *Clin Immunol* 2016;162:100-106.

**Cite this article as:** Hackett EE, Sheedy FJ. miR-21 alters circulating Treg function in vascular disease—hope for restoring immunoregulatory responses in atherosclerosis? *Ann Transl Med* 2017;5(1):21. doi: 10.21037/atm.2016.12.72

# The role of circulating microRNAs in acute coronary syndromes: ready for prime time?

Gert Klug, Bernhard Metzler

Cardiology and Angiology, University Clinic of Internal Medicine III, Medical University of Innsbruck, Anichstraße 35, A-6020 Innsbruck, Austria

*Correspondence to:* Gert Klug. Cardiology and Angiology, University Clinic of Internal Medicine III, Medical University of Innsbruck, Anichstrasse 35, A-6020 Innsbruck, Austria. Email: Gert.Klug@tirol-kliniken.at.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Navickas R, Gal D, Laucevičius A, *et al.* Identifying circulating microRNAs as biomarkers of cardiovascular disease: a systematic review. *Cardiovasc Res* 2016;111:322-37.

Submitted Oct 18, 2016. Accepted for publication Oct 24, 2016.

doi: 10.21037/atm.2016.11.64

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.11.64>

Recently Navickas *et al.* published a review on the role of microRNAs (miRs) as biomarkers of cardiovascular disease in *Cardiovascular Research* (1). Based on a systematic literature research their aim was to determine the diagnostic and prognostic value of miRs in healthy subjects, subjects with stable coronary artery disease and patients with different forms of unstable coronary artery disease (unstable angina, non-STEMI and STEMI). They identified n=487 papers and extracted n=19 studies, reporting on 52 different miRs, after a rigorous quality check. The largest amount of evidence, through all stages of cardiovascular disease, was found for miR-133a/b (5 studies), miR-208a/b (6 studies) and miR-499 (7 studies). Furthermore the promising role of miR-1 (3 studies) in the diagnosis of acute coronary syndromes and the regulation of miR-145 in STEMI patients is highlighted. A meta-analysis, however, is not presented because of heterogeneous study designs and analytical reasons (1).

Especially in patients with acute coronary syndromes the role of novel biomarkers is rapidly evolving (2). Cardiac troponin measured with standard (3) and high-sensitive (4) assays has improved our abilities to define patients with acute coronary syndromes (5), estimate the amount of myocardial necrosis (3), predict functional impairment (6,7) and prognosis (8). Another well-established biomarker in acute coronary syndromes is NT-pro-BNP (9). Today these two biomarkers impact clinical decisions of cardiologist every day. However their diagnostic performance is

hampered by diagnostic windows, their relatively low specificity (10) and their correlation with renal function (11). Therefore there is indeed a need to identify novel biomarkers in acute coronary syndromes.

miR-1 is very specific for cardiac skeletal muscle and plays an important role during cardiogenesis and proliferation of cardiomyocytes (12). Three different studies, with a total of n=583 patients with acute coronary syndromes and n=259 controls (13-15), investigated the role of miR-1 in the initial diagnosis of patients with chest pain and suspected acute coronary syndrome. Wang *et al.* observed that miR-1 levels in patients with acute myocardial infarction are elevated compared to controls, but the diagnostic performance of miR-1 was inferior to cardiac troponin I (AUC: 0.85 *vs.* 0.99) (13). Oerlemans *et al.* described an increase in miR-1 even in patients with initially negative troponin levels or in patients presenting within 3 hours after symptom onset (14). Furthermore, Widera *et al.* showed that miR-1 levels are significantly higher in patients with NSTEMI or STEMI than in patients with unstable angina, although they did not predict mortality at 6 months (15). Anyhow, these results identify miR-1 as one of the most promising miRs for the early diagnosis of acute coronary syndromes, especially in the combination with other biomarkers.

The largest study included in this review was performed by Devaux *et al.* (16). It prospectively investigated the use of six different miRs in n=1,155 patients with acute chest pain and suspected acute myocardial infarction. Finally,



n=179 patients were diagnosed as NSTEMI and n=45 patients as suffering from a STEMI. miR-133a, miR-208b and miR-499 were identified as univariate predictors of myocardial infarction. As noted by Navickas *et al.* all these three miRs control cardiomyocyte identity (17). However, their predictive value did not remain significant after correction for troponin levels. Furthermore, the area under the receiver operating curves were low (AUC: 0.53–0.76) compared to high sensitive troponin (AUC: 0.94). The miR-133a, miR-208b and miR-499 levels were significantly higher in STEMI patients than in NSTEMI patients (16). These findings are in line with the findings of Widera *et al.* on miR-133a and miR-208a (15), although both studies failed to demonstrate an independent prognostic value of all miRs studied.

The potential unique role of miR-133a in STEMI patients is further highlighted by a study by Eitel *et al.* (18). In this study miR-133a levels were associated with decreased myocardial salvage, larger infarct size and microvascular obstruction (19) as determined by cardiac magnetic resonance in a clearly defined study population of n=216 consecutive STEMI patients. Although miR-133a was a univariate predictor of mortality and MACE (HR: 1.28) the use of cardiac magnetic resonance for infarct characterisation (20) allowed the authors to demonstrate, that this association is not independent of infarct characteristics (18).

Another study focusing exclusively on patients with STEMI was performed by Dong *et al.* (21) who investigated the prognostic value of miR-145, which regulates vascular smooth muscle cell and cardiomyocyte differentiation and has been shown to correlate with infarct size (22). In n=245 with STEMI they demonstrated that miR-145 levels above the median predicted 12-months MACE independent of NT-pro-BNP, creatine kinase or troponin levels (HR: 5.6) (21). Interestingly miR-145 levels have been observed to be generally lower in patients with severe coronary artery disease or acute coronary syndromes which might indicate altered expression of miR-145 in these patients (23). As these findings seem controversial further research should clarify the role of miR-145 in cardiovascular disease.

Navickas *et al.* have done a valuable work in identifying five, out of more than 2,000 described in humans, miRs which have great potential to improve our daily clinical work in the future. Their review is based on the data of 19 studies with more than 6,000 participants (1). These miRs are ready for prime time in cardiovascular research

but further studies are warranted to provide reliable and standardised quantification with faster PCR and microarray technologies. Then it should be possible to include these promising biomarkers in controlled, large-scale, well-powered trials and, perhaps someday, into clinical practice.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Navickas R, Gal D, Laucevičius A, et al. Identifying circulating microRNAs as biomarkers of cardiovascular disease: a systematic review. *Cardiovasc Res* 2016;111:322-337.
2. Reinstadler SJ, Feistritzer HJ, Reindl M, et al. Combined biomarker testing for the prediction of left ventricular remodelling in ST-elevation myocardial infarction. *Open Heart* 2016;3:e000485.
3. Klug G, Mayr A, Mair J, et al. Role of biomarkers in assessment of early infarct size after successful p-PCI for STEMI. *Clin Res Cardiol* 2011;100:501-510.
4. Reinstadler SJ, Feistritzer HJ, Klug G, et al. High-sensitivity troponin T for prediction of left ventricular function and infarct size one year following ST-elevation myocardial infarction. *Int J Cardiol* 2016;202:188-193.
5. Thygesen K, Alpert JS, Jaffe AS, et al. Third universal definition of myocardial infarction. *Glob Heart* 2012;7:275-295.
6. Mayr A, Mair J, Klug G, et al. Cardiac troponin T and creatine kinase predict mid-term infarct size and left ventricular function after acute myocardial infarction: a cardiac MR study. *J Magn Reson Imaging* 2011;33:847-854.
7. Feistritzer HJ, Klug G, Reinstadler SJ, et al. Novel biomarkers predicting cardiac function after acute myocardial infarction. *Br Med Bull* 2016;119:63-74.
8. Sherwood MW, Morrow DA, Scirica BM, et al. Early dynamic risk stratification with baseline troponin levels and 90-minute ST-segment resolution to predict 30-day cardiovascular mortality in ST-segment elevation myocardial infarction: analysis from CLopidogrel as Adjunctive Reperfusion Therapy (CLARITY)-

- Thrombolysis in Myocardial Infarction (TIMI) 28. *Am Heart J* 2010;159:964-971.e1.
9. Mayr A, Mair J, Schocke M, et al. Predictive value of NT-pro BNP after acute myocardial infarction: relation with acute and chronic infarct size and myocardial function. *Int J Cardiol* 2011;147:118-123.
  10. Haaf P, Drexler B, Reichlin T, et al. High-sensitivity cardiac troponin in the distinction of acute myocardial infarction from acute cardiac noncoronary artery disease. *Circulation* 2012;126:31-40.
  11. Reinstadler SJ, Klug G, Feistritz HJ, et al. Copeptin testing in acute myocardial infarction: ready for routine use? *Dis Markers* 2015;2015:614145.
  12. Zhao Y, Samal E, Srivastava D. Serum response factor regulates a muscle-specific microRNA that targets Hand2 during cardiogenesis. *Nature* 2005;436:214-220.
  13. Wang GK, Zhu JQ, Zhang JT, et al. Circulating microRNA: a novel potential biomarker for early diagnosis of acute myocardial infarction in humans. *Eur Heart J* 2010;31:659-666.
  14. Oerlemans MI, Mosterd A, Dekker MS, et al. Early assessment of acute coronary syndromes in the emergency department: the potential diagnostic value of circulating microRNAs. *EMBO Mol Med* 2012;4:1176-1185.
  15. Widera C, Gupta SK, Lorenzen JM, et al. Diagnostic and prognostic impact of six circulating microRNAs in acute coronary syndrome. *J Mol Cell Cardiol* 2011;51:872-875.
  16. Devaux Y, Mueller M, Haaf P, et al. Diagnostic and prognostic value of circulating microRNAs in patients with acute chest pain. *J Intern Med* 2015;277:260-271.
  17. Xin M, Olson EN, Bassel-Duby R. Mending broken hearts: cardiac development as a basis for adult heart regeneration and repair. *Nat Rev Mol Cell Biol* 2013;14:529-541.
  18. Eitel I, Adams V, Dieterich P, et al. Relation of circulating MicroRNA-133a concentrations with myocardial damage and clinical prognosis in ST-elevation myocardial infarction. *Am Heart J* 2012;164:706-714.
  19. Klug G, Mayr A, Schenk S, et al. Prognostic value at 5 years of microvascular obstruction after acute myocardial infarction assessed by cardiovascular magnetic resonance. *J Cardiovasc Magn Reson* 2012;14:46.
  20. Klug G, Metzler B. Assessing myocardial recovery following ST-segment elevation myocardial infarction: short- and long-term perspectives using cardiovascular magnetic resonance. *Expert Rev Cardiovasc Ther* 2013;11:203-219.
  21. Dong YM, Liu XX, Wei GQ, et al. Prediction of long-term outcome after acute myocardial infarction using circulating miR-145. *Scand J Clin Lab Invest* 2015;75:85-91.
  22. Meder B, Keller A, Vogel B, et al. MicroRNA signatures in total peripheral blood as novel biomarkers for acute myocardial infarction. *Basic Res Cardiol* 2011;106:13-23.
  23. Gao H, Guddeti RR, Matsuzawa Y, et al. Plasma Levels of microRNA-145 Are Associated with Severity of Coronary Artery Disease. *PLoS One* 2015;10:e0123477.

**Cite this article as:** Klug G, Metzler B. The role of circulating microRNAs in acute coronary syndromes: ready for prime time? *Ann Transl Med* 2016;4(24):537. doi: 10.21037/atm.2016.11.64

## Should we expect novel biomarkers of myocardial infarction?

Marek Kiliszek<sup>1</sup>, Agata Maciejak<sup>2</sup>

<sup>1</sup>Department of Cardiology and Internal Diseases, Military Institute of Medicine, Warsaw, Poland; <sup>2</sup>Institute of Biochemistry and Biophysics, Polish Academy of Sciences, Warsaw, Poland

*Correspondence to:* Marek Kiliszek. Department of Cardiology and Internal Diseases, Military Institute of Medicine, 128 Szaserów Str., 04-141 Warsaw, Poland. Email: kiliszek@mp.pl.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Yao XL, Lu XL, Yan CY, *et al.* Circulating miR-122-5p as a potential novel biomarker for diagnosis of acute myocardial infarction. *Int J Clin Exp Pathol* 2015;8:16014-9.

Submitted May 09, 2016. Accepted for publication May 12, 2016.

doi: 10.21037/atm.2016.05.33

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.05.33>

Recent studies have revealed important roles for microRNAs (miRNAs) in cardiovascular disease, including acute myocardial infarction (AMI). miRNAs are small, 20–25 nucleotides long, non-coding RNA molecules, which inhibit gene expression by promoting mRNA degradation or preventing translation (1,2). Although the biological functions of miRNAs are not fully understood, numerous studies have shown that some miRNAs have unique expression profiles in certain tissues or cell types (3). Recent discoveries have revealed the existence of freely circulating, stably expressed miRNAs in human blood cells or plasma/serum (4,5). The circulating miRNAs have been shown to be sensitive and informative biomarkers in the diagnosis of cardiovascular diseases. The article by Yao and colleagues (6) describes circulating miRNA-122-5p as a potential novel biomarker for diagnosis of acute myocardial infarction. In that study, the authors investigated the level of miRNA-122-5p by quantitative real-time PCR (RT-qPCR) and found that its expression was up-regulated at 4, 8, 12, and 24 h in AMI patients compared to non-AMI controls, and displayed similar trends to the cTnI concentrations. A high correlation was observed between the circulating miR-122-5p and cTnI concentrations. The receiver operating characteristic (ROC) curve analysis showed that miRNA-122-5p in plasma had considerable diagnostic accuracy for AMI with an area under curve (AUC) of 0.855. The results suggest that miRNA-122-5p could leak from cardiac myocytes into the circulation during the early stages of AMI.

This circulating miR-122-5p could be a useful biomarker

for the diagnosis of AMI. Furthermore, miRNA expression analysis, particularly when combined with clinical parameters, such as cTnI concentrations, provides better understanding of the changes that occur in the myocardium and determine the potential role of extracellular miRNA-122-5p as a paracrine signaling molecule. The authors took into account that the small sample size was a major limitation of the study.

The authors do not state how they found out that this miR-122-5p might be elevated in AMI—until now it was mainly linked with liver injury (7). However, some papers showing upregulation of miR-122 (miR-122 and miR-122-5p are originating from the same hairpin, see e.g., <https://www.exiqon.com/mirsearch>) in AMI have also been published (8,9). Of note, the study by Li *et al.* (9) was performed using miRNA microarrays representing 1,205 human miRNAs and the results were validated on a large group of patients (two independent cohorts of 111 and 428 patients). Therefore, we have another replication showing increase in miR-122-5p expression in patients with AMI.

Interestingly, numerous studies have indicated that heart-specific miRNAs could be released into the circulation during AMI, making them potentially useful in aiding diagnosis or guiding therapy in acute coronary syndrome (10). Such miRNAs can easily be detected in the circulation and serve as potential biomarkers for cardiovascular diseases. For instance, Gidlöf *et al.* (11) found that the plasma level of some cardiac-associated miRNAs, such as miRNA-1, miRNA-133a, miRNA-208b, and miRNA-499-5p significantly increased in STEMI patients. D'Alessandra *et al.* (12)



have reported that in an acute hind-limb ischemia, unlike in AMI, plasma levels of miRNA-1, miRNA-133a, miRNA-133b, and miRNA-499-5p did not increase, indicating that they are ideal biomarkers for AMI. Ai *et al.* (13) showed that circulating miRNA-1 level was significantly higher in AMI patients compared with non-AMI group and the level returned to normal on discharge following medication. Corsten *et al.* (14) evaluated plasma levels of heart-associated miRNAs (miR-1, miR-133a, miR-208b, and miR-499), fibrosis-associated miRNAs (miRNA-21 and miRNA-29b), and leukocyte-associated miRNAs (miRNA-146, miRNA-155, and miRNA-223) in patients with various cardiac damage including AMI, viral myocarditis, diastolic dysfunction, and acute HF. miR-208b and miRNA-499 were found to be significantly elevated in AMI patients compared to healthy controls. The ROC curve analysis revealed a good diagnostic value of miRNA-208b and miRNA-499 as biomarkers for AMI.

Circulating miRNA-122-5p could broaden our understanding of the role of miRNAs in the pathogenesis of acute coronary syndromes and, if confirmed, it could become a novel biomarker for AMI. We need publications on large cohorts of patients with clinical follow-up, to see if expression of miR-122-5p gives us additional predictive value to clinical data and classical biomarkers. The real revolution is just behind the doorstep—first attempts are currently being made to use miR antagonists for the treatment of patients: now those with hepatitis C, but a novel field in cardiology is opening up (15).

## Acknowledgements

*Funding:* This work was supported by the National Science Centre, Poland (2014/13/N/NZ5/01403).

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

- Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281-297.
- Inui M, Martello G, Piccolo S. MicroRNA control of signal transduction. *Nat Rev Mol Cell Biol* 2010;11:252-263.
- Sood P, Krek A, Zavolan M, et al. Cell-type-specific signatures of microRNAs on target mRNA expression. *Proc Natl Acad Sci U S A* 2006;103:2746-2751.
- Gilad S, Meiri E, Yegorov Y, et al. Serum microRNAs are promising novel biomarkers. *PLoS One* 2008;3:e3148.
- Chen X, Ba Y, Ma L, et al. Characterization of microRNAs in serum: a novel class of biomarkers for diagnosis of cancer and other diseases. *Cell Res* 2008;18:997-1006.
- Yao XL, Lu XL, Yan CY, et al. Circulating miR-122-5p as a potential novel biomarker for diagnosis of acute myocardial infarction. *Int J Clin Exp Pathol* 2015;8:16014-16019.
- Tan Y, Ge G, Pan T, et al. Serum MiRNA panel as potential biomarkers for chronic hepatitis B with persistently normal alanine aminotransferase. *Clin Chim Acta* 2015;451:232-239.
- D'Alessandra Y, Carena MC, Spazzafumo L, et al. Diagnostic potential of plasmatic MicroRNA signatures in stable and unstable angina. *PLoS One* 2013;8:e80345.
- Li X, Yang Y, Wang L, et al. Plasma miR-122 and miR-3149 Potentially Novel Biomarkers for Acute Coronary Syndrome. *PLoS One* 2015;10:e0125430.
- Ahlin F, Arfvidsson J, Vargas KG, et al. MicroRNAs as circulating biomarkers in acute coronary syndromes: A review. *Vascul Pharmacol* 2016;81:15-21.
- Gidlöf O, Andersson P, van der Pals J, et al. Cardiospecific microRNA plasma levels correlate with troponin and cardiac function in patients with ST elevation myocardial infarction, are selectively dependent on renal elimination, and can be detected in urine samples. *Cardiology* 2011;118:217-226.
- D'Alessandra Y, Devanna P, Limana F, et al. Circulating microRNAs are new and sensitive biomarkers of myocardial infarction. *Eur Heart J* 2010;31:2765-2773.
- Ai J, Zhang R, Li Y, et al. Circulating microRNA-1 as a potential novel biomarker for acute myocardial infarction. *Biochem Biophys Res Commun* 2010;391:73-77.
- Corsten MF, Dennert R, Jochems S, et al. Circulating MicroRNA-208b and MicroRNA-499 reflect myocardial damage in cardiovascular disease. *Circ Cardiovasc Genet* 2010;3:499-506.
- Miravirsin Study in Null Responder to Pegylated InterCirculating MicroRNA-208b and MicroRNA-499 reflect myocardial damage in cardiovascular disease. *feron Alpha Plus Ribavirin Subjects With Chronic Hepatitis C*. Accessed 29 April, 2016. Available online: <https://clinicaltrials.gov/show/NCT01727934>

**Cite this article as:** Kiliszek M, Maciejak A. Should we expect novel biomarkers of myocardial infarction? *Ann Transl Med* 2016;4(11):227. doi: 10.21037/atm.2016.05.33

# The hunt for fatal myocardial infarction biomarkers: predictive circulating microRNAs

Francesco Russo<sup>1</sup>, Milena Rizzo<sup>2,3</sup>, Kirstine Belling<sup>1</sup>, Søren Brunak<sup>1</sup>, Lasse Folkersen<sup>4</sup>

<sup>1</sup>Novo Nordisk Foundation Center for Protein Research, Faculty of Health and Medical Sciences, University of Copenhagen, Copenhagen, Denmark; <sup>2</sup>Institute of Clinical Physiology (IFC), National Research Council (CNR), Pisa, Italy; <sup>3</sup>Tuscan Tumor Institute, Florence, Italy; <sup>4</sup>Center for Biological Sequence analysis, Technical University of Denmark, Lyngby, Denmark

*Correspondence to:* Francesco Russo. Novo Nordisk Foundation Center for Protein Research, Faculty of Health and Medical Sciences, University of Copenhagen, 2200 Copenhagen, Denmark. Email: francesco.russo@cpr.ku.dk

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Bye A, Røsjø H, Nauman J, *et al.* Circulating microRNAs predict future fatal myocardial infarction in healthy individuals - The HUNT study. *J Mol Cell Cardiol* 2016;97:162-168.

Submitted Jul 14, 2016. Accepted for publication Jul 18, 2016.

doi: 10.21037/atm.2016.08.21

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.08.21>

## Introduction

The concept of a hype cycle is a well-established business concept, in which novel ideas are said to have an initial wave of hype followed by disillusionment. Only after that, the novel concept takes off and become truly useful entering a so-called plateau of productivity. In biomedical science, the field of microRNAs (miRNAs) certainly had a peak of interest in the end of the last decade. This led by high impact publications (1) and characterization of both novel miRNA-entities as well as their associations to a broad range of diseases. Nonetheless, no clear pharmaceutical successes emerged: miRNA targets are being pursued as therapeutic targets, but none have as of yet successfully made it through clinical trials (2). Likewise the use of miRNA-based treatment strategies targeting regular mRNA is an area of interest (3). In this editorial we focus on a third aspect of miRNAs: the use of miRNAs as prognostic biomarkers in disease, asking the question if miRNAs are now entering this plateau of productivity in which actual benefit will be seen.

We focus on the recent paper by Bye *et al.*: “Circulating microRNAs predict future fatal myocardial infarction in healthy individuals - The HUNT study” (4). This paper is of particular interest because it presents strong evidence for prognostic benefit of miRNAs. The study was based on the

HUNT cohort, a Norwegian biobank-initiative in which an impressive 88% of the adult population of the Nord-Trøndelag County participated, giving blood samples and questionnaire information in 1984, 1995, and 2006. With the unique advantage of having both frozen serum and decades of follow-up information, the study was designed as a prospective nested case-control design with fatal acute myocardial infarct (AMI) as endpoint and controls matched on risk factors such as body mass index (BMI), total cholesterol and high-density lipoprotein cholesterol (HDL-C) (4). The main discovery phase results yielded 12 miRNAs that were associated with future AMI. This editorial will discuss the perspectives of these findings as well as considerations for similar future miRNA studies.

## The methodology of miRNA normalization

A key consideration in biomarker studies is the existence of similar studies. For miRNA biomarkers predicting AMI, several studies already exist that address AMI risk. Typically the main focus is the discovery of biomarkers for immediate use, such as distinguishing patients with ST-elevated AMI from patients with stable ischemic heart disease (5), or between ongoing AMI and healthy controls (6). The most comparable study to the interest of this editorial is the one Zampetaki *et al.* In this study,

the main focus was the prediction of future AMI and the authors did find association of miR-126, miR-223 and miR-197 to the disease (7). It is noteworthy that none of these miRNAs were identified by Bye *et al.* (4). Hence, for the overall purpose of using miRNAs as predictive biomarkers it prompts an important discussion on methodological choice.

Bye *et al.* suggested that one main discrepancy reason could be the choice of data normalization method and the platform for miRNA analysis. In fact, while Bye *et al.* used a panel of seven reference genes for normalization and quality control by means of the RNA Spike-in kit including cel-miR-39-3p, UniSp2, UniSp4, UniSp5 and UniSp6, Zampetaki *et al.* (7) solely used U6, which is not a suitable endogenous control for the quantification of circulating miRNAs based on previous works (8,9). It has been shown that the Spike-in system improves the quality of the normalization step (10). The normalization method for circulating miRNA quantification is one of the critical aspects in this field and from this point of view the normalization procedure used in the work of Bye *et al.* is the most robust to date.

### Sample collection and processing in miRNA analysis

Another crucial aspect for the analysis of circulating miRNAs is the collection and processing of blood samples. In fact, it has been suggested that blood must be processed within a few hours after collection in order to prevent cell-derived miRNA contamination from red blood cells or platelets (11-13). Unfortunately, many studies do not follow this suggestion or do not report this important information. Moreover, it has been shown that the difference between serum and plasma strongly affects the spectrum of circulating miRNA in blood (11) demonstrating higher miRNA concentrations in serum samples compared to the corresponding plasma samples. Considering that Bye *et al.* and Zampetaki *et al.* extracted miRNAs from serum and plasma, respectively, and that they used two different RNA isolation kits (miRCURY RNA isolation and miRNeasy kit, respectively), it is plausible that the two studies did not find the same miRNAs. All these considerations point the attention on the fact that, given the numerous factors that generate variability in circulating miRNA studies, it is now mandatory to develop standard protocols for blood specimen collection and processing to allow the comparison across studies.

### Using small RNA-seq to improve the quality of the results

Circulating miRNAs are considered novel non-invasive biomarkers. Yet, the mechanism of action at the molecular level both in healthy and disease is still largely unknown. Since there is a great opportunity to establish a new paradigm of intercellular communication, the National Institutes of Health (NIH) funded a novel Common Fund's Extracellular RNA Communication (ERC) program "(I) to discover fundamental biological principles about the mechanisms of extracellular RNA (exRNA) generation, secretion, and transport; (II) to identify and develop a catalogue of exRNA in normal human body fluids; (III) and to investigate the potential for using exRNAs as therapeutic molecules or biomarkers of disease". In order to disseminate the knowledge derived from this program, the results are shared through the exRNA research portal, a community-wide resource for exRNA standards, protocols and data. These efforts have already generated new small RNA-seq data for several conditions (including cardiovascular diseases and cancer), biofluids (e.g., plasma and serum) and RNA sources (e.g., exosomes and other extracellular vesicles). Since the quality, the amount and the specific body fluid are important factors (as discussed above), RNA-seq is likely to be the future standard technique in this field. Still, small RNA-seq is not the common method used as shown in the miRandola database, the circulating RNAs database (14). In the work of Bye *et al.* and in many other published studies, qRT-PCR has been used as golden standard for miRNA quantification. Since in this context the normalization step is crucial and there is no clear agreement in the scientific society, using small RNA-seq could solve this crucial problem, increasing the quality of the results. Overall, rigorous standardized methods and analyses are needed in this field. It has been reported that many confounding factors exist in the phases of processing, extraction and quantification of exRNAs.

### Statistical considerations in search for predictive biomarkers

In understanding biomarker discovery studies it is important to be very aware of the statistical pitfalls associated with them (15). The archetypical discovery study pitfall is that of the winner's curse; that testing hundreds of metrics will inevitably yield significant findings by chance and that their effect estimates will be inflated (16). In the Bye *et al.* study (4)

the study design was built around a discovery phase as well as a validation phase. In the discovery phase, 76 miRNAs were tested, of which 12 were selected at an uncorrected  $P < 0.05$  significance threshold. This alone is of course a clear example of test metric inflation and winner's curse, and it follows that the  $\Delta\Delta Cq$  values should decrease in the validation phase, which in fact they do. However, the study also included a validation phase within which it was shown that ten of the 12 miRNAs were significantly associated with future AMI at  $P < 0.05$ . No metric was provided reporting with a formal multiple testing correction of 12 miRNAs, but it is reported that four of the 12 miRNAs also were significant at  $P < 0.01$ . Further, Bye *et al.* presents a combination signature of five other miRNAs. These resulted from the testing of 4,095 different combinations with no independent replication. It is from this signature that the 0.91 AUC is concluded.

The replication and validation setup is a good strategy to amend winners curse problems. Ultimately, however, the proper question to debate here is of course whether these statistics will hold up in a general case. The combination signature score of 0.91 AUC finding was based on a large un-replicated multiple burden and so is highly likely to decrease on independent validation. However, the individual miRNA scores were independently replicated suggesting that they are true in a general case. And that is a novelty in a field burdened by underpowered discovery studies without genuine follow up, so we believe that at least the four strongly replicating miRNAs, let-7g-5p, miR-26a-5p, miR-106a-5p, and miR-151a-5p, could play important future roles in the field of AMI prognostics.

## Conclusions

Current prediction algorithms in clinical use include the European Society of Cardiology's Systematic Coronary Risk Evaluation (SCORE) and the Framingham risk score (17,18). These algorithms have some impact in a clinical setting; by accurately assigning patients to risk-groups they can prompt important discussions of smoking patterns, LDL-cholesterol levels and overall healthy lifestyle. However, the scores are still too inaccurate to clearly pinpoint the individuals who will in fact become future patients. This is the reason why predictive biomarkers are of such interest. Having the ability to identify individuals who has a high risk of adverse events with only low chance of false positives; that is a hallmark of precision medicine, and one that is not possible only with the current life-style

based clinical scores.

While the work with miRNA biomarkers for AMI is still in its infancy, studies like Bye *et al.* pave a way for a future in which life-style scores could be supplemented with simple and cheap blood-test-based biomarker predictions, and resultant in early and accurate intervention. More accurate intervention, importantly, also means less wasteful and non-required treatment of individuals who are in fact not at risk. And this optimization of the health care system really is the grand perspective to have in mind when considering precision medicine in general, and studies like Bye *et al.* in particular.

## Acknowledgements

*Funding:* This work was supported by the Novo Nordisk Foundation (grant agreement NNF14CC0001) and a grant from the Danish Innovation Fund (145-2014-5).

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

1. Sonkoly E, Wei T, Janson PC, et al. MicroRNAs: novel regulators involved in the pathogenesis of psoriasis? *PLoS One* 2007;2:e610.
2. Li Z, Rana TM. Therapeutic targeting of microRNAs: current status and future challenges. *Nat Rev Drug Discov* 2014;13:622-638.
3. van Rooij E, Purcell AL, Levin AA. Developing microRNA therapeutics. *Circ Res* 2012;110:496-507.
4. Bye A, Røsjø H, Nauman J, et al. Circulating microRNAs predict future fatal myocardial infarction in healthy individuals - The HUNT study. *J Mol Cell Cardiol* 2016;97:162-168.
5. Wei T, Folkersen L, Ehrenborg E, et al. MicroRNA 486-3P as a stability marker in acute coronary syndrome. *Biosci Rep* 2016.36.
6. Zhang L, Chen X, Su T, et al. Circulating miR-499 are novel and sensitive biomarker of acute myocardial infarction. *J Thorac Dis* 2015;7:303-308.
7. Zampetaki A, Willeit P, Tilling L, et al. Prospective study on circulating MicroRNAs and risk of myocardial infarction. *J Am Coll Cardiol* 2012;60:290-299.
8. Xiang M, Zeng Y, Yang R, et al. U6 is not a suitable

- endogenous control for the quantification of circulating microRNAs. *Biochem Biophys Res Commun* 2014;454:210-214.
9. Benz F, Roderburg C, Vargas Cardenas D, et al. U6 is unsuitable for normalization of serum miRNA levels in patients with sepsis or liver fibrosis. *Exp Mol Med* 2013;45:e42.
  10. Marabita F, de Candia P, Torri A, et al. Normalization of circulating microRNA expression data obtained by quantitative real-time RT-PCR. *Brief Bioinform* 2016;17:204-212.
  11. Wang K, Yuan Y, Cho JH, et al. Comparing the MicroRNA spectrum between serum and plasma. *PLoS One* 2012;7:e41561.
  12. Cheng HH, Yi HS, Kim Y, et al. Plasma processing conditions substantially influence circulating microRNA biomarker levels. *PLoS One* 2013;8:e64795.
  13. El-Khoury V, Pierson S, Kaoma T, et al. Assessing cellular and circulating miRNA recovery: the impact of the RNA isolation method and the quantity of input material. *Sci Rep* 2016;6:19529.
  14. Russo F, Di Bella S, Nigita G, et al. miRandola: extracellular circulating microRNAs database. *PLoS One* 2012;7:e47786.
  15. Ioannidis JP, Greenland S, Hlatky MA, et al. Increasing value and reducing waste in research design, conduct, and analysis. *Lancet* 2014;383:166-175.
  16. Young NS, Ioannidis JP, Al-Ubaydli O. Why current publication practices may distort science. *PLoS Med* 2008;5:e201.
  17. Lloyd-Jones DM, Wilson PW, Larson MG, et al. Framingham risk score and prediction of lifetime risk for coronary heart disease. *Am J Cardiol* 2004;94:20-24.
  18. Conroy RM, Pyörälä K, Fitzgerald AP, et al. Estimation of ten-year risk of fatal cardiovascular disease in Europe: the SCORE project. *Eur Heart J* 2003;24:987-1003.

**Cite this article as:** Russo F, Rizzo M, Belling K, Brunak S, Folkersen L. The hunt for fatal myocardial infarction biomarkers: predictive circulating microRNAs. *Ann Transl Med* 2016;4(Suppl 1):S1. doi: 10.21037/atm.2016.08.21



# Correlations between microRNAs and their target genes in skeletal myoblasts cell therapy for myocardial infarction

Andrea Rognoni<sup>1</sup>, Chiara Cavallino<sup>1,2</sup>, Francesco Rametta<sup>2</sup>, Angelo Sante Bongo<sup>1</sup>

<sup>1</sup>Coronary Care Unit and Catheterization Laboratory, “Maggiore della Carità Hospital”, Novara, Italy; <sup>2</sup>Division of Cardiology, Sant’Andrew Hospital, Vercelli, Italy

*Correspondence to:* Andrea Rognoni, MD, FSCAI. Coronary Care Unit and Catheterization Laboratory, “Maggiore della Carità” Hospital, Corso Mazzini 18, 28100 Novara, Italy. Email: arognoni@hotmail.com.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Liu Q, Du GQ, Zhu ZT, *et al.* Identification of apoptosis-related microRNAs and their target genes in myocardial infarction post-transplantation with skeletal myoblasts. *J Transl Med* 2015;13:270.

Submitted May 30, 2016. Accepted for publication May 31, 2016.

doi: 10.21037/atm.2016.05.65

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.05.65>

In the setting of ischemic heart disease in which revascularization therapies (both percutaneous and surgery) and pharmaceutical therapy are able to contribute to ventricular remodeling, nowadays we also have some indications about promising options to prevent or reverse the ventricular remodelling process and consequent heart failure. Stem cells provide an alternative curative intervention for the infarcted heart by compensating for the cardiomyocyte loss subsequent to myocardial injury. The presence of resident stem and progenitor cell populations in the heart, and nuclear reprogramming of somatic cells with genetic induction of pluripotency markers are the emerging new developments in stem cell-based regenerative medicine. However, until safety and feasibility of these cells are established by extensive experimentation in *in vitro* and *in vivo* experimental models, skeletal muscle-derived myoblasts, and bone marrow cells remain the most well-studied donor cell types for myocardial regeneration and repair (1,2). Furthermore in the literature we can find a lot of indications about microRNAs as novel and alternative cardiac biomarkers. These are 22-nucleotide-long non-coding RNAs that regulate gene expression at post-transcriptional level. Several recent studies have shown that miRNAs play a physiological role in cardiovascular homeostasis and in the pathogenesis of cardiovascular disease. Expression-pattern studies of myocardial tissue reveal that several miRNAs are up- or down-regulated during myocardial infarction (3,4).

During nearly two decades of cell therapy research for treatment of ischemic heart disease, stem cell transplantation, either alone or in combination with the other therapeutic interventions, has demonstrated promise as a novel curative strategy (5). The prime advantage of the heart cell therapy using stem cells is its capability to replace the loss-of-functioning cardiomyocytes to preserve the deterioration of left ventricular function (6,7). Of all stem cells used, skeletal muscle-derived myoblasts have been investigated in experimental and clinical studies; in fact skeletal myoblasts and bone marrow derived stem cells remain the most well-characterized studied therapy for myocardial reparability in the patients with ischemic heart disease (8,9). Skeletal myoblasts constitute the renewable source of progenitor cells in skeletal muscle that participate in the repair process in the event of injury. The most important characteristics of skeletal myoblasts that make them suitable for use are their autologous availability, potential to expand *in vitro*, resistance to ischemia, low risk of tumorigenesis and myogenic differentiation potential (10,11). In the paper of Liu *et al.*, published on *Journal of Transplantation Medicine* in 2015, the authors analyzed the correlations between microRNAs and their target genes in skeletal myoblasts cell therapy for myocardial infarction. Their final data showed that the down regulation of apoptosis-regulatory microRNAs and in turn up regulation of target genes may partially account for rescue effect of skeletal myoblasts

therapy for myocardial infarction (12). Since the early 2000s we can find in the literature a lot of *in vitro* researches that have assessed the performance of skeletal myoblasts for the treatment of ischemic and non ischemic cardiomyopathies in animal models (13-16). All these studies showed that skeletal myoblasts prevented left ventricular remodeling, preserved ejection fraction, left ventricular pressure wall thickness and left ventricular pressure. Skeletal myoblasts were, also, used in post myocardial patients in some clinical settings (17,18). Even in these cases, results from heart function were promising increasing in left ventricular ejection fraction and in segmental contractility on echocardiography. The interesting feature of the paper of Liu *et al.* (12) was to compare the expression of microRNAs in post myocardial infarction rats with or without skeletal myoblasts cell transplantation. The authors focused their research on new apoptosis-associated microRNAs and their target genes; in particular they showed that four microRNAs were down regulated in the skeletal myoblasts treated group compared with the untreated group. Some studies in the recent past showed the role of microRNAs; some of this have distinct roles in modulating skeletal and cardiac muscle proliferation and differentiation (19-21); other studies have, also, demonstrated that microRNA-206 and -1 directly down regulated gap junction coupling after the initiation of myoblast fusion *in vitro* and *in vivo* and inhibit Cx 43 expression during myoblast differentiation without altering Cx43 microRNA levels (22). The paper of Liu *et al.*, (12) is the first comparative *in vitro* study of microRNA and microRNA expression in myocardial infarction heart treated with skeletal myoblasts transplantation using surgical route. We hope as interesting for the future skeletal myoblasts transplantation combined with transplantation of other cell types or growth factor; a combined therapy based on simultaneous delivery of skeletal myoblasts and bone marrow stem cells may be more effective with either cell type alone as partially demonstrated in the recent past (23). In-depth mechanistic studies, genetic reprogramming or pharmacological manipulation, and combinatorial approach involving skeletal myoblast transplantation with other relevant interventions, such as growth factor administration, can help safety and therapeutic efficacy; in fact simultaneous insertion of skeletal myoblasts with multiple growth factors promoted their ability to integrate with host myocytes. Skeletal myoblasts remain the most well-studied donor cell type.

## Acknowledgements

None.

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

1. Cleland JG, McGowan J. Heart failure due to ischaemic heart disease: epidemiology, pathophysiology and progression. *J Cardiovasc Pharmacol* 1999;33:S17-S29.
2. Frangogiannis NG. The immune system and cardiac repair. *Pharmacol Res* 2008;58:88-111.
3. Rognoni A, Cavallino C, Lupi A, et al. Novel biomarkers in the diagnosis of acute coronary syndromes: the role of circulating miRNAs. *Expert Rev Cardiovasc Ther* 2014;12:1119-1124.
4. Baskerville S, Bartel DP. Microarray profiling of microRNAs reveals frequent coexpression with neighboring miRNAs and host genes. *RNA* 2005;11:241-247.
5. Leri A, Hosoda T, Rota M, et al. Myocardial Regeneration by Exogenous and Endogenous Progenitor Cells. *Drug Discov Today Dis Mech* 2007;4:197-203.
6. Kajstura J, Rota M, Whang B, et al. Bone marrow cells differentiate in cardiac cell lineages after infarction independently of cell fusion. *Circ Res* 2005;96:127-137.
7. Haider HKh, Ye L, Jiang S, et al. Angiomyogenesis for cardiac repair using human myoblasts as carriers of human vascular endothelial growth factor. *J Mol Med (Berl)* 2004;82:539-549.
8. Mias C, Lairez O, Trouche E, et al. Mesenchymal stem cells promote matrix metalloproteinase secretion by cardiac fibroblasts and reduce cardiac ventricular fibrosis after myocardial infarction. *Stem Cells* 2009;27:2734-2743.
9. Ince H, Petzsch M, Rehders TC, et al. Transcatheter transplantation of autologous skeletal myoblasts in postinfarction patients with severe left ventricular dysfunction. *J Endovasc Ther* 2004;11:695-704.
10. Menasché P. Skeletal myoblasts as a therapeutic agent. *Prog Cardiovasc Dis* 2007;50:7-17.
11. Al Attar N, Carrion C, Ghostine S, et al. Long-term (1 year) functional and histological results of autologous skeletal muscle cells transplantation in rat. *Cardiovasc Res* 2003;58:142-148.

12. Liu Q, Du GQ, Zhu ZT, et al. Identification of apoptosis-related microRNAs and their target genes in myocardial infarction post-transplantation with skeletal myoblasts. *J Transl Med* 2015;13:270.
13. Al Attar N, Carrion C, Ghostine S, et al. Long-term (1 year) functional and histological results of autologous skeletal muscle cells transplantation in rat. *Cardiovasc Res* 2003;58:142-148.
14. Taylor DA, Atkins BZ, Hungspreugs P, et al. Regenerating functional myocardium: improved performance after skeletal myoblast transplantation. *Nat Med* 1998;4:929-933.
15. Hata H, Matsumiya G, Miyagawa S, et al. Grafted skeletal myoblast sheets attenuate myocardial remodeling in pacing-induced canine heart failure model. *J Thorac Cardiovasc Surg* 2006;132:918-924.
16. He KL, Yi GH, Sherman W, et al. Autologous skeletal myoblast transplantation improved hemodynamics and left ventricular function in chronic heart failure dogs. *J Heart Lung Transplant* 2005;24:1940-1949.
17. Menasché P, Haggè AA, Scorsin M, et al. Myoblast transplantation for heart failure. *Lancet* 2001;357:279-280.
18. Haggè AA, Carrion C, Menasché P, et al. Viability and differentiation of autologous skeletal myoblast grafts in ischaemic cardiomyopathy. *Lancet* 2003;361:491-492.
19. Conboy IM, Conboy MJ, Smythe GM, et al. Notch-mediated restoration of regenerative potential to aged muscle. *Science* 2003;302:1575-1577.
20. Chen JF, Mandel EM, Thomson JM, et al. The role of microRNA-1 and microRNA-133 in skeletal muscle proliferation and differentiation. *Nat Genet* 2006;38:228-233.
21. Kim HK, Lee YS, Sivaprasad U, et al. Muscle-specific microRNA miR-206 promotes muscle differentiation. *J Cell Biol* 2006;174:677-687.
22. Anderson C, Catoe H, Werner R., et al. MIR-206 regulates connexin43 expression during skeletal muscle development. *Nucleic Acids Res* 2006;34:5863-5871.
23. Ye L, Haider HKh, Sim EK, et al. Adult stem cells for cardiac repair: a choice between skeletal myoblasts and bone marrow stem cells. *Exp Biol Med (Maywood)* 2006;231:8-19.

**Cite this article as:** Rognoni A, Cavallino C, Rametta F, Bongo AS. Correlations between microRNAs and their target genes in skeletal myoblasts cell therapy for myocardial infarction. *Ann Transl Med* 2016;4(15):292. doi: 10.21037/atm.2016.05.65



# Is the regulation of SIRT1 by miRNA-34a the key to mesenchymal stem cell survival?

Michael A. Bellio<sup>1</sup>, Wayne Balkan<sup>1</sup>, Joshua M. Hare<sup>1</sup>, Ivonne Hernandez Schulman<sup>1,2</sup>

<sup>1</sup>Interdisciplinary Stem Cell Institute, <sup>2</sup>Division of Nephrology and Hypertension, University of Miami Miller School of Medicine, Miami, FL, USA  
 Correspondence to: Ivonne Hernandez Schulman, MD. Associate Professor of Clinical Medicine, Division of Nephrology and Hypertension, Program Director, Interdisciplinary Stem Cell Institute, University of Miami Miller School of Medicine, Biomedical Research Building/Room 810, 1501 N.W. 10th Avenue, Miami, FL 33136, USA. Email: ischulman@med.miami.edu.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Brambilla E, Le Teuff G, Marguet S, *et al.* Prognostic effect of tumor lymphocytic infiltration in resectable non-small-cell lung cancer. *J Clin Oncol* 2016;34:1223-30.

Submitted May 12, 2016. Accepted for publication May 17, 2016.

doi: 10.21037/atm.2016.05.45

View this article at: <http://dx.doi.org/10.21037/atm.2016.05.45>

Mesenchymal stem cells (MSCs) are currently being used clinically to treat cardiovascular diseases, including ischemic heart disease, heart failure, and peripheral vascular disease (1,2). To date, these trials have proven MSCs to be a safe and effective option for the improvement of vascular function, reduction of scar size, and reversal of remodeling in heart failure (3,4). Despite the positive data being collected, there are still a number of challenges that reduce the effectiveness of MSC cell therapy. Cell survival and engraftment in the hostile microenvironment of the diseased myocardium severely limits MSC regenerative potential. Research focusing on MSC biology, including the identification of genes and molecules that regulate aspects of regeneration, is critical towards the establishment of effective cell production and delivery strategies (5,6).

Gene therapy and the manipulation of protein expression via genetic editing and targeted microRNA (miRNA) technologies has become a promising avenue for the enhancement of cellular regenerative potential (7). The delivery of genetically modified MSCs to ischemic hearts in pre-clinical studies has advanced our understanding of which genes could potentially improve the efficacy of MSC therapy. Overexpression of genes such as extracellular superoxide dismutase (ecSOD) and vascular endothelial growth factor (VEGF) in MSCs has improved survival and tissue-repair when cells are injected into sites of ischemic

injury (8,9). Similarly, modification of miRNAs via lentiviral induced expression or silencing has made it possible to target entire pathways rather than individual genes. Delivery of MSCs transduced to overexpress miRNA-126 enhanced ischemic angiogenesis through the activation of AKT and ERK regulated signaling pathways (10). Together, these studies have provoked further investigation to establish the optimal gene targeting systems that will optimize MSC therapeutic applications.

In an intriguing study published in *Stem Cell Research and Therapy*, Zhang *et al.* demonstrated a novel role for miRNA-34a in the regulation of apoptosis and senescence by MSCs via the silent information regulator 1 (SIRT1)-mediated pathways (11). As a well-studied miRNA, the down-regulation of miRNA-34a promotes proliferation, increases stress resistance, and promotes cell migration in a variety of cell types. These outcomes are particularly appealing to a cell-therapy application where cell survival and tissue integration is at risk. The therapeutic advantages of modifying miRNA-34a expression are further showcased through the feasibility of specific alteration using oligonucleotide mimetic or inhibition. Particularly in cancer therapy, induced expression of miRNA-34a safely reduced tumor growth and survival (12). In this context, the authors describe that MSC apoptosis and decline of cell proliferation correlates with miRNA-34a upregulation. This response serves as a promising therapeutic target in MSCs

because recent studies have demonstrated a consistent role of miRNA-34a in cell cycle progression, particularly due to its regulation of genes such as p53, c-kit, SIRT1, and Notch (13). Based on these findings, the authors formulated and tested the hypothesis that overexpression of miRNA-34a exacerbates hypoxia and serum starvation-induced apoptosis and senescence in MSCs.

The combination of hypoxia and serum starvation is a common inducer of MSC apoptosis, and is a reliable model to test molecular mechanisms that may be altered upon cell injection into hypoxic tissue (14). Zhang *et al.* used this model to test the potency of miRNA-34a overexpression and silencing on cell apoptosis via the detection of annexin-V positive cells. As expected, those MSCs engineered to overexpress miRNA-34a exhibited a greater amount of apoptosis upon hypoxic serum starvation conditions, while silencing miRNA-34a significantly reduced this effect. These findings were then expanded upon in order to identify the key molecular players involved in the miRNA-34a response.

SIRT1 is an established and key target of miRNA-34a due to its endogenous regulation of anti-apoptotic and pro-proliferative pathways. Acting as a NAD-dependent protein deacetylase, SIRT1 has been described as a longevity factor, targeting transcription factors such as forkhead box proteins (FOX) and p53, while tightly regulating cellular resistance to oxidative damage (15). However, it had not been previously shown whether miRNA-34a manipulation and subsequent regulation of SIRT1 affects MSC biology. The authors used SIRT1 gene silencing paired with miRNA-34a inhibition to test the involvement of SIRT1 in the miRNA-34a-mediated apoptotic response. The authors demonstrated that miRNA-34a regulates SIRT1 and the downstream pro-apoptotic factor FOXO3a. Moreover, silencing SIRT1 expression abolished the anti-apoptotic effects of inhibiting miRNA-34a. This result was confirmed by the alteration of apoptotic markers caspase-3, poly ADP ribose polymerase (PARP), and mitochondrial cytochrome C. Lastly, markers of DNA damage and senescence decreased with miRNA-34a inhibition, providing further evidence for the multifaceted effects of miRNA-34a in MSCs.

Although these findings serve as a promising first step toward establishing a genetic strategy designed to effectively preserve the viability of MSCs delivered into the ischemic myocardium, many questions are still left to be resolved. The broad range of RNA targets of miRNA-34a may be beneficial or problematic, and the long-term effect

of miRNA-34a inhibition on MSCs has not been explored. The goal of MSC therapy is for transplanted MSCs to provide lasting reparative effects on myocardial structure and function by engrafting into the myocardium. miRNA-34a overexpression vectors are currently being investigated in translational models of cancer to halt unregulated cell proliferation and cancer metastasis (16). Whether the permanent inhibition of miRNA-34a in MSCs is a safe option that does not pose tumorigenic risk needs to be determined. Additionally, genes that influence MSC differentiation and angiogenesis, two important factors for the regenerative response, are known targets of miRNA-34a. Expression of platelet-derived growth factor receptor (PDGFR), which is down regulated by miRNA-34a, plays a significant role in MSC mediated vasculogenesis (17,18). It would be important to investigate how the regulation of multiple pathways by miRNA-34a would impact the *in vivo* differentiation and tissue integration of MSCs. Lastly, the experiments performed by Zhang *et al.* were all done in 5% oxygen (11), a level that is hypoxic compared to standard laboratory culture conditions (21% oxygen) but not compared to the physiological oxygen levels in the mammalian heart. It has been reported that ischemia in the heart could decrease oxygen levels from a physiologic 5% to a low of 1–3% oxygen (19). To confirm the effectiveness of miRNA-34a silencing in ischemia, similar experiments would need to be conducted at lower oxygen tensions.

In summary, Zhang *et al.* have presented a novel and promising genetic targeting strategy that could enhance the effectiveness of MSC therapy in ischemic heart disease. Their findings demonstrate that miRNA-34a inhibition results in greater viability of MSCs in an *in vitro* model of hypoxia and serum starvation, warranting further investigation in an *in vivo* translational model.

### Acknowledgements

*Funding:* This work is supported by NIH grants, UM1 HL113460, R01 HL084275, and R01 HL110737, the Starr Foundation and the Soffer Family Foundation.

### Footnote

*Conflicts of Interest:* Dr. Hare has a patent for cardiac cell-based therapy; he holds equity in Vestion Inc., maintains a professional relationship with Vestion as a consultant and member of the Board of Directors and Scientific Advisory Board, and is a shareholder in Longeveron LLC. The other

authors have no conflicts of interest to declare.

## References

- Sanina C, Hare JM. Mesenchymal stem cells as a biological drug for heart disease: where are we with cardiac cell-based therapy? *Circ Res* 2015;117:229-233.
- Karantalis V, Hare JM. Use of mesenchymal stem cells for therapy of cardiac disease. *Circ Res* 2015;116:1413-1430.
- Hare JM, Fishman JE, Gerstenblith G, et al. Comparison of allogeneic vs autologous bone marrow-derived mesenchymal stem cells delivered by transcatheter injection in patients with ischemic cardiomyopathy: the POSEIDON randomized trial. *JAMA* 2012;308:2369-2379.
- Premier C, Blum A, Bellio MA, et al. Allogeneic mesenchymal stem cells restore endothelial function in heart failure by stimulating endothelial progenitor cells. *EBioMedicine* 2015;2:467-475.
- Kanashiro-Takeuchi RM, Schulman IH, Hare JM. Pharmacologic and genetic strategies to enhance cell therapy for cardiac regeneration. *J Mol Cell Cardiol* 2011;51:619-625.
- Golpanian S, Schulman IH, Ebert RF, et al. Concise review: review and perspective of cell dosage and routes of administration from preclinical and clinical studies of stem cell therapy for heart disease. *Stem Cells Transl Med* 2016;5:186-191.
- Nowakowski A, Walczak P, Janowski M, et al. Genetic engineering of mesenchymal stem cells for regenerative medicine. *Stem Cells Dev* 2015;24:2219-2242.
- Pan Q, Qin X, Ma S, et al. Myocardial protective effect of extracellular superoxide dismutase gene modified bone marrow mesenchymal stromal cells on infarcted mice hearts. *Theranostics* 2014;4:475-486.
- Lai T, Li M, Zheng L, et al. Over-expression of VEGF in marrow stromal cells promotes angiogenesis in rats with cerebral infarction via the synergistic effects of VEGF and Ang-2. *J Huazhong Univ Sci Technolog Med Sci* 2012;32:724-731.
- Chen JJ, Zhou SH. Mesenchymal stem cells overexpressing MiR-126 enhance ischemic angiogenesis via the AKT/ERK-related pathway. *Cardiol J* 2011;18:675-681.
- Zhang F, Cui J, Liu X, et al. Roles of microRNA-34a targeting SIRT1 in mesenchymal stem cells. *Stem Cell Res Ther* 2015;6:195.
- Li L, Xie X, Luo J, et al. Targeted expression of miR-34a using the T-VISA system suppresses breast cancer cell growth and invasion. *Mol Ther* 2012;20:2326-2334.
- Ye Z, Fang J, Dai S, et al. MicroRNA-34a induces a senescence-like change via the down-regulation of SIRT1 and up-regulation of p53 protein in human esophageal squamous cancer cells with a wild-type p53 gene background. *Cancer Lett* 2016;370:216-221.
- Zhu W, Chen J, Cong X, et al. Hypoxia and serum deprivation-induced apoptosis in mesenchymal stem cells. *Stem Cells* 2006;24:416-425.
- Bordone L, Guarente L. Calorie restriction, SIRT1 and metabolism: understanding longevity. *Nat Rev Mol Cell Biol* 2005;6:298-305.
- Misso G, Di Martino MT, De Rosa G, et al. Mir-34: a new weapon against cancer? *Mol Ther Nucleic Acids* 2014;3:e194.
- Peng Y, Guo JJ, Liu YM, et al. MicroRNA-34A inhibits the growth, invasion and metastasis of gastric cancer by targeting PDGFR and MET expression. *Biosci Rep* 2014.34.
- Gomes SA, Rangel EB, Premier C, et al. S-nitrosoglutathione reductase (GSNOR) enhances vasculogenesis by mesenchymal stem cells. *Proc Natl Acad Sci U S A* 2013;110:2834-2839.
- Roy S, Khanna S, Wallace WA, et al. Characterization of perceived hyperoxia in isolated primary cardiac fibroblasts and in the reoxygenated heart. *J Biol Chem* 2003;278:47129-47135.

**Cite this article as:** Bellio MA, Balkan W, Hare JM, Schulman IH. Is the regulation of SIRT1 by miRNA-34a the key to mesenchymal stem cell survival? *Ann Transl Med* 2016;4(12):243. doi: 10.21037/atm.2016.05.45

# My heart will go on—beneficial effects of anti-MiR-30 after myocardial infarction

Yuhuang Li, Lars Maegdefessel

Department of Medicine, Karolinska Institutet, Solna, Stockholm 17176, Sweden

*Correspondence to:* Lars Maegdefessel, MD, PhD. Center for Molecular Medicine L8:03, Department of Medicine, Karolinska Institutet and University Hospital, Stockholm 17176, Sweden. Email: lars.maegdefessel@ki.se.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

**Abstract:** MicroRNAs play key roles in the regulation of diverse cellular processes and during cardiovascular disease development and progression, such as myocardial infarction (MI) and heart failure (HF). A recent manuscript by Shen and colleagues provided evidence that the miR-30-CSE-H<sub>2</sub>S axis contributes to the protection against cardiomyocyte ischemic injury by regulating hydrogen sulfide (H<sub>2</sub>S) production. Inhibition of the miR-30 family after MI injury offers potential therapeutic value to ‘keep our heart going on’. As this study highlights miRNAs as promising future therapeutic targets, their translational applicability to utilization in humans needs to be viewed with caution.

**Keywords:** microRNAs; myocardial Infarction (MI); therapeutics; cardiomyocytes

Submitted Feb 22, 2016. Accepted for publication Feb 25, 2016.

doi: 10.21037/atm.2016.03.12

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.03.12>

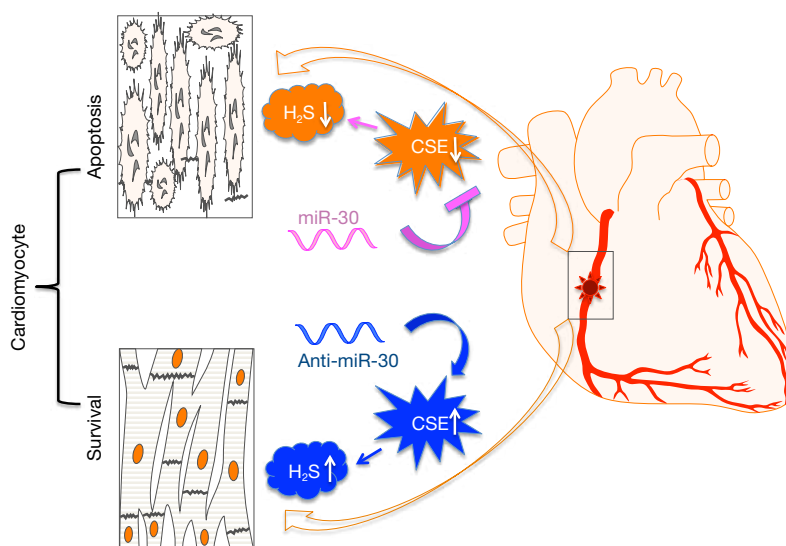
MicroRNAs (miRNAs) are small, non-coding RNA molecules, approximately 22 nucleotides in length, which act as master regulators of gene expression (1). Mature miRNAs are single stranded RNAs that block translation—or induce degradation of mRNA by base pairing to partially or perfectly complementary sites on their target mRNA, usually in the 3'-UTR (2). Accumulated data have indicated that miRNAs play key roles in the regulation of diverse cellular processes and during cardiovascular disease development and progression, such as atherosclerosis, hypertension, arrhythmias, left ventricular hypertrophy, myocardial infarction (MI), as well as heart failure (HF) (3,4).

Despite all available therapeutic approaches, MI still ranks as the main cause of death worldwide (5). Pathophysiological landmarks of MI include acute myocardial damage, post-ischemic neovascularization, and cardiac remodeling (6). Acute myocardial damage is attributed by ischemic cellular hypoxia, which results in an increase of reactive oxygen species during early reperfusion, endothelial cell (EC) activation, cytokine production in the damaged area, priming and recruiting of

neutrophils bordering the infarcted region, and ultimately cardiomyocyte death, endothelial capillary impairment and post-ischemic neovascularization. The remodeling process is initially an adaptive mechanism to maintain adequate cardiac function, but eventually leads to fibrosis, left ventricular dilatation, and HF (7).

Evidentially, all these processes are tightly regulated by miRNAs (6). Primarily, miRNAs participate in the regulation of cardiomyocyte fate, such as the pro-apoptotic miRs-15, -34, -320, and -140; the anti-apoptotic miRs-24 and -214; the pro-proliferative miR-17-92 cluster, miRs-199a and -590; and anti-proliferative miRs-15 and -133. Secondly, several miRNAs have been described to induce (miR-210) or repress (miR-15, -24, -26, and -17-92 cluster) angiogenesis. Furthermore, miRNAs modulate cardiac remodeling via inhibiting (miR-15 and -34) or enhancing (miR-1, -21, -24, -126, -155, -221, and -499) the function of stem and/or progenitor cells.

A recent manuscript by Shen and colleagues identifies a novel and crucial role for the miR-30 family in acute MI injury and cardiac function (8). In this study, the authors



**Figure 1** Therapeutic effect of anti-miR-30 on cardiomyocyte survival in myocardial ischemia.

emphasized on the cardio-protective role of hydrogen sulfide ( $H_2S$ ), predominantly deriving from L-cysteine and being catalyzed by cystathionine-c-lyase (CSE). Their experiments provide additional evidence that the miR-30-CSE- $H_2S$  axis contributes to the protection against cardiomyocyte ischemic injury, both *in vitro* and *in vivo*, by regulating  $H_2S$  production.

The authors were able to discover significant deduction of CSE and  $H_2S$  in MI, which negatively correlates with expression levels of the miR-30 family. Of importance, they could confirm that CSE is a direct target of the miR-30 family. The miR-30 family regulates CSE mRNA and protein levels, thus affects  $H_2S$  production and hypoxia-induced cardiomyocyte injury. In addition, silencing the miR-30 family not only protects against cardiomyocyte apoptosis during hypoxia, but also increases cardiac function evaluated by echocardiography after MI induction in mice, which turns out to be a CSE-dependent mechanism, as these effects were absent in genetically mutated CSE knockout mice. In contrast, delivery of miR-30b in mice greatly aggravated MI injury. Taken together, the study is able to demonstrate for the very first time that the miR-30 family regulates  $H_2S$  production by directly targeting CSE. Inhibition of the miR-30 family after MI injury offers clear therapeutic value to ‘keep our heart going on’ (Figure 1).

One limitation to the translational feasibility of the study is that the authors did not evaluate potential off-target effects in other organ systems than the targeted heart (e.g.,

liver, kidney) in which systemically administered miRNA modulators assimilate to a much higher extent. Further translationally focused pre-clinical studies should take these considerations into account. One promising solution may be local delivery of miRNAs highlighted in the past by Hinkel *et al.* (9). In their study, locked nucleic acid-modified antisense miR-92a (LNA-92a) was applied either regionally (antegrade or retrograde) with a catheter or systemically (intravenously). They confirmed that LNA-92a reduced miR-92a expression in the infarct zone regardless of the application venue. However, catheter-based delivery, but not intravenous infusion, reduced the infarct size compared with control LNA-treated pigs, which correlated with an improved ejection fraction and left ventricular end-diastolic pressure, while not accumulating in other organs (such as kidney and liver). In addition, as reported by previous studies (10,11), both down-regulated miR-30a and up-regulated miR-30c/d could aggravate myocardial hypertrophy, indicating that distinctive modulation of a miR-30 family subtype would need to be taken into account when developing future therapeutic cardiomyocyte rescue strategies after acute MI in humans. Last but not the least, endogenous  $H_2S$  level subsequent to miR-30 modulation should be tightly controlled due to its latent toxicity.

The therapeutic value of miRNA in cardiovascular as well as other diseases is obvious. By having unique expression profiles and higher stability in biological samples, miRNAs quickly emerged as novel biomarkers.

The ability to regulate multiple genes in various disease-contributing signaling pathways makes them promising future therapeutic targets. Noteworthy, translational applicability to utilization in humans needs to be viewed with caution, especially issues relating to potential immune stimulatory effects, mode of delivery, and off-target effects. These obstacles need to be overcome to bring miRNA therapeutics into mainstream clinical practice.

### Acknowledgements

*Funding:* Research in the Maegdefessel laboratory on non-coding RNAs in cardiovascular diseases is supported by the Swedish Heart-Lung-Foundation (20120615, 20130664, and 20140186), the Ragnar Söderberg Foundation (M55/14), the Swedish Research Council (2015-03140), and the European Research Council (ERC-StG NORVAS).

### Footnote

*Conflicts of Interest:* Yuhuang Li is a CERIC (Center of Excellence for Research in Inflammatory and Cardiovascular Diseases) scholar at the Karolinska Institute, Stockholm, Sweden. The other author has no conflicts of interest to declare.

### References

1. Yates LA, Norbury CJ, Gilbert RJ. The long and short of

- microRNA. *Cell* 2013;153:516-519.
2. Thum T, Condorelli G. Long noncoding RNAs and microRNAs in cardiovascular pathophysiology. *Circ Res* 2015;116:751-762.
  3. Romaine SP, Tomaszewski M, Condorelli G, et al. MicroRNAs in cardiovascular disease: an introduction for clinicians. *Heart* 2015;101:921-928.
  4. Small EM, Olson EN. Pervasive roles of microRNAs in cardiovascular biology. *Nature* 2011;469:336-342.
  5. Hausenloy DJ, Yellon DM. Ischaemic conditioning and reperfusion injury. *Nat Rev Cardiol* 2016. [Epub ahead of print].
  6. Boon RA, Dimmeler S. MicroRNAs in myocardial infarction. *Nat Rev Cardiol* 2015;12:135-142.
  7. Fiedler J, Thum T. MicroRNAs in myocardial infarction. *Arterioscler Thromb Vasc Biol* 2013;33:201-205.
  8. Shen Y, Shen Z, Miao L, et al. miRNA-30 family inhibition protects against cardiac ischemic injury by regulating cystathionine- $\gamma$ -lyase expression. *Antioxid Redox Signal* 2015;22:224-240.
  9. Hinkel R, Penzkofer D, Zühlke S, et al. Inhibition of microRNA-92a protects against ischemia/reperfusion injury in a large-animal model. *Circulation* 2013;128:1066-1075.
  10. Pan W, Zhong Y, Cheng C, et al. MiR-30-regulated autophagy mediates angiotensin II-induced myocardial hypertrophy. *PLoS One* 2013;8:e53950.
  11. Jentzsch C, Leierseder S, Loyer X, et al. A phenotypic screen to identify hypertrophy-modulating microRNAs in primary cardiomyocytes. *J Mol Cell Cardiol* 2012;52:13-20.

**Cite this article as:** Li Y, Maegdefessel L. My heart will go on—beneficial effects of anti-MiR-30 after myocardial infarction. *Ann Transl Med* 2016;4(7):144. doi: 10.21037/atm.2016.03.12



# MicroRNA-499-5p: a therapeutic target in the context of cardiovascular disease

**Menno Hoekstra**

Division of Biopharmaceutics, Leiden Academic Centre for Drug Research, Gorlaeus Laboratories, Einsteinweg 55, 2333CC Leiden, The Netherlands

*Correspondence to:* Menno Hoekstra, PhD. Division of Biopharmaceutics, Leiden Academic Centre for Drug Research, Gorlaeus Laboratories, Einsteinweg 55, 2333CC Leiden, The Netherlands. Email: hoekstra@lacdr.leidenuniv.nl.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* O Sullivan JF, Neylon A, McGorrian C, *et al.* miRNA-93-5p and other miRNAs as predictors of coronary artery disease and STEMI. *Int J Cardiol* 2016;224:310-6.

Submitted Oct 19, 2016. Accepted for publication Oct 24, 2016.

doi: 10.21037/atm.2016.11.61

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.11.61>

Cardiovascular disease remains a major cause of death worldwide. Atherosclerosis, narrowing of the arteries due to the accumulation of cholesterol in macrophages within the sub-endothelial space of the vessel wall, is the primary underlying cause of cardiovascular disease. Hypercholesterolemia is therefore an established risk factor for the development of atherosclerotic lesions and cardiovascular disease (1,2). Other major cardiovascular risk factors include smoking, high blood pressure, and the presence of type II diabetes (1,2). Unfortunately, it is still difficult to reliably predict the risk for the development of cardiovascular disease and the associated mortality in individual subjects. A clear need therefore exists for the discovery of sensitive cardiovascular disease biomarkers.

MicroRNAs constitute a large family of highly conserved non-coding RNAs that inhibit the protein expression of their target genes through modulation of the mRNA translation rate and/or stability (3). Initial findings from the oncology research field have indicated that these ~22 nucleotide long RNA species may be useful as biomarkers since several tumorous tissues display a specific change in their microRNA expression profile as compared to their related non-diseased tissues (4-7). In follow-up studies we have been able to show that the presence of cardiovascular disease in humans subjects, e.g. in unstable and stable angina pectoris patients, coincides with a change in the microRNA profile of peripheral blood mononuclear cells (PBMCs) (8).

More specifically, we observed that the relative expression levels of miR-135a and miR-147 were respectively 5-fold higher and 4-fold lower in PBMC fractions from coronary artery disease patients as compared to those of unaffected controls. Our studies also revealed that the PMBC fractions of stable and unstable angina pectoris patients could be distinguished based upon the expression level of three specific microRNAs. Relative expression levels of miR-134, miR-370, and miR-198 were higher in unstable subjects that had experienced ischemic chest pain at rest within the preceding 48 h versus those that had stable effort angina of >6 months duration. It thus appears that microRNAs may be able to also serve as biomarkers of (unstable) coronary artery disease.

In the context of our previous observations, in this commentary, I would like to highlight the paper by O Sullivan *et al.* that has recently been published in the *International Journal of Cardiology* (9) as it provides strong novel support for the predictive power of microRNA signatures in the cardiovascular disease setting. Similar as in our study, O Sullivan *et al.* aimed to show the differential expression of microRNAs in patients with stable coronary artery disease, unstable coronary artery disease and control subjects with normal coronary angiograms. Their unstable group of patients consisted of subjects who presented with ST-Elevation Myocardial Infarction (STEMI) as diagnosed per 2013 ACCF/AHA Guidelines for the Management of



ST-Elevation Myocardial Infarction. All STEMI group inclusions therefore had presented within 12 h of chest pain onset, undergone primary percutaneous intervention (PCI), and had coronary artery occlusion confirmed by angiography. A clear improvement as compared to our studies was that O Sullivan *et al.* included such a high number of patients that they could also correct for possible interactions with known cardiovascular risk factors. Levels of miR-93-5p (increased), miR-146a-5p (decreased), miR-16-5p (increased), and miR-15a-5p (increased) were all significantly changed in plasma specimens of stable coronary artery disease patients as compared to those obtained from controls (9). However, the most striking finding of the study by O Sullivan *et al.* was that inclusion of plasma miR-499a-5p levels (increased in STEMI patients versus controls) in the prediction model significantly enhanced the sensitivity as compared to traditional risk factors alone to identify subjects suffering from myocardial infarction (9). As such, the data from O Sullivan *et al.* suggest that high plasma miR-499a-5p levels may serve as a complementary biomarker for the presence of acute coronary syndromes.

Expression profiling in pigs has suggested that miR-499-5p is highly conserved and preferentially expressed in the myocardium (10). MiR-499-5p is therefore generally regarded to be a cardio-specific microRNA in humans. Importantly, the observation by O Sullivan *et al.* that the presence of the cardiac microRNA miR-499a-5p in plasma contains high predictive power in the context of acute coronary syndromes does not stand by itself. Olivieri *et al.* showed that median circulating levels of miR-499-5p were significantly higher at admission in acute myocardial infarction patients that died within the following year as compared to those that survived the cardiovascular event (11). Furthermore, a step-wise increase in plasma miR-499-5p levels was observed as compared to healthy controls in subjects suffering from acute heart failure without evidence of acute myocardial infarction and those that did display acute non-ST elevation myocardial infarction (12,13). Moreover, studies by Gidlöf *et al.* (13) and D'Allesandra *et al.* (14) have indicated that miR-499-5p levels are transiently elevated in plasma of human subjects in response to the development of myocardial infarction. In further support of the notion that miR-499-5p levels can be used as a highly sensitive biomarker of acute cardiovascular events, Gidlöf *et al.* also observed that circulating miR-499-5p levels can reliably predict the presence of STEMI (13,14).

In agreement with the aforementioned human findings,

induction of myocardial infarction induced a rapid, but transient, increase in plasma levels of miR-499-5p that peaked at 24 hours after the coronary artery occlusion in both mice and pigs (13,14). The miR-499-5p plasma profile in the murine myocardial infarction model was not mimicked by that of other microRNAs supposedly expressed specifically in cardiac muscle (14). The increase in plasma miR-499-5p levels is thus probably not due to a non-specific secretion of the microRNA from cardiac tissue, e.g. in response to myocardial infarction-associated necrosis of cardiomyocytes. Interestingly, a rapid decrease in miR-499-5p levels has been detected in response to hypoxia in cultured rat cardiomyocytes *in vitro* (15). Based upon these combined findings, one can assume that a decrease in cardiomyocyte miR-499-5p levels and concomitant rise in plasma miR-499-5p levels may therefore be a general biomarker of cardiac distress. A higher plasma level of miR-499-5p thus would associate with a higher degree of cardiac dysfunction. In accordance, athletes that have immensely challenged their heart through running a marathon also display a transient rise in circulating miR-499-5p levels (16).

The question remains as to whether the increase in plasma miR-499-5p levels is only a biomarker of cardiac dysfunction/hypoxia or if this microRNA actually plays a role in the pathogenesis of acute cardiovascular events. In their elegant study, Li *et al.* have recently addressed this issue. Overexpression of miR-499-5p in cultured cardiomyocytes lowers programmed cell death protein 4 (neoplastic transformation inhibitor; PDCD4) mRNA expression which translates into a decreased apoptosis rate, while miR-499-5p inhibition increases PDCD4 transcript and protein levels and induces cardiomyocyte apoptosis (15). In line with an inverse relation between miR-499-5p levels and cardiomyocyte death, a lower miR-499-5p expression can be found in infarcted (dying) versus non-infarcted (healthy) cardiac tissue (15). Importantly, overexpression of miR-499-5p in cardiomyocytes was able to protect the heart against myocardial infarction-associated tissue damage *in vivo*. A remarkable ~50% decrease in infarct size was noted in miR-499-5p agomir-treated mice as compared to controls (15). It has been suggested that microRNAs circulating in the plasma compartment, i.e., in membrane vesicles, can be transferred to recipient cells to facilitate cell-to-cell communication (17). When taking the *in vitro* and *in vivo* findings from Li *et al.* into account, it can be hypothesized that, under myocardial infarction conditions, hypoxic cardiomyocytes release miR-499-5p for subsequent transfer to and incorporation by unaffected cells to confer

protection against myocardial infarction-associated cellular apoptosis and tissue death.

In conclusion, the study by O Sullivan *et al.* has (1) provided substantial new support for the relevance of miR-499-5p as non-invasive biomarker of acute coronary events and (2) highlighted the general potential of circulating microRNAs as predictors of disease. The recent discovery that miR-499-5p may play a protective role in cardiomyocytes has opened up new possibilities to treat subjects at risk of developing acute cardiovascular syndromes. Thus far, phase I and II clinical trials involving microRNA-based therapies, i.e. treatment of cancer patients with a liposome-formulated mimic of the tumor suppressor miR-34 (MRX34), have not yet yielded valuable drugs (18). However, it is conceivable that therapeutic approaches aimed at increasing plasma miR-499-5p levels will be developed in the future that can be of benefit for high risk cardiovascular disease patients.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The author has no conflicts of interest to declare.

### References

- Gordon T, Kannel WB. Multiple risk functions for predicting coronary heart disease: the concept, accuracy, and application. *Am Heart J* 1982;103:1031-1039.
- Kannel WB. Lipids, diabetes, and coronary heart disease: insights from the Framingham Study. *Am Heart J* 1985;110:1100-1107.
- Nelson P, Kiriakidou M, Sharma A, et al. The microRNA world: small is mighty. *Trends Biochem Sci* 2003;28:534-540.
- Coulouarn C, Factor VM, Andersen JB, et al. Loss of miR-122 expression in liver cancer correlates with suppression of the hepatic phenotype and gain of metastatic properties. *Oncogene* 2009;28:3526-3536.
- Busacca S, Germano S, De Cecco L, et al. MicroRNA signature of malignant mesothelioma with potential diagnostic and prognostic implications. *Am J Respir Cell Mol Biol* 2010;42:312-319.
- Raponi M, Dossey L, Jatko T, et al. MicroRNA classifiers for predicting prognosis of squamous cell lung cancer. *Catheter Cardiovasc Interv* 2013;81:E1-E8.
- Viswanathan SR, Powers JT, Einhorn W, et al. Lin28 promotes transformation and is associated with advanced human malignancies. *Nat Genet* 2009;41:843-848.
- Hoekstra M, van der Lans CA, Halvorsen B, et al. The peripheral blood mononuclear cell microRNA signature of coronary artery disease. *Biochem Biophys Res Commun* 2010;394:792-797.
- O Sullivan JF, Neylon A, McGorrian C, et al. miRNA-93-5p and other miRNAs as predictors of coronary artery disease and STEMI. *Int J Cardiol* 2016;224:310-316.
- Reddy AM, Zheng Y, Jagadeeswaran G, et al. MicroRNA classifiers for predicting prognosis of squamous cell lung cancer. *Cancer Res* 2009;69:5776-5783.
- Olivieri F, Antonicelli R, Spazzafumo L, et al. Admission levels of circulating miR-499-5p and risk of death in elderly patients after acute non-ST elevation myocardial infarction. *Int J Cardiol* 2014;172:e276-e278.
- Olivieri F, Antonicelli R, Lorenzi M, et al. Diagnostic potential of circulating miR-499-5p in elderly patients with acute non ST-elevation myocardial infarction. *Int J Cardiol* 2013;167:531-536.
- Gidlöf O, Andersson P, van der Pals J, et al. Cardiospecific microRNA plasma levels correlate with troponin and cardiac function in patients with ST elevation myocardial infarction, are selectively dependent on renal elimination, and can be detected in urine samples. *Cardiology* 2011;118:217-226.
- D'Alessandra Y, Devanna P, Limana F, et al. Circulating microRNAs are new and sensitive biomarkers of myocardial infarction. *Eur Heart J* 2010;31:2765-2773.
- Li Y, Lu J, Bao X, et al. MiR-499-5p protects cardiomyocytes against ischaemic injury via anti-apoptosis by targeting PDCD4. *Oncotarget* 2016;7:35607-35617.
- Baggish AL, Park J, Min PK, et al. Rapid upregulation and clearance of distinct circulating microRNAs after prolonged aerobic exercise. *J Appl Physiol* 2014;116:522-531.
- Turchinovich A, Tonevitsky AG, Burwinkel B. Extracellular miRNA: A Collision of Two Paradigms. *Trends Biochem Sci* 2016;41:883-892.
- Shah MY, Ferrajoli A, Sood AK, et al. microRNA Therapeutics in Cancer — An Emerging Concept. *EBioMedicine* 2016;12:34-42.

**Cite this article as:** Hoekstra M. MicroRNA-499-5p: a therapeutic target in the context of cardiovascular disease. *Ann Transl Med* 2016;4(24):539. doi: 10.21037/atm.2016.11.61

# Early detection of myocardial infarction – microRNAs right at the time?

Nicolle Kränkel<sup>1,2</sup>, Stefan Blankenberg<sup>3,4</sup>, Tanja Zeller<sup>3,4</sup>

<sup>1</sup>Department of Cardiology, Charité Universitätsmedizin, Campus Benjamin Franklin, Berlin, Germany; <sup>2</sup>German Center for Cardiovascular Research, partner site Berlin, Berlin, Germany; <sup>3</sup>Univeristy Heart Center Hamburg, Clinic for General and Interventional Cardiology, Hamburg, Germany; <sup>4</sup>German Center for Cardiovascular Research, partner site Hamburg/Lübeck/Kiel, Hamburg, Germany

*Correspondence to:* Tanja Zeller. Univeristy Heart Center Hamburg, Clinic for General and Interventional Cardiology (Genomics and Systems Biology), Hamburg, Germany. Email: t.zeller@uke.de.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Wang KJ, Zhao X, Liu YZ, *et al.* Circulating MiR-19b-3p, MiR-134-5p and MiR-186-5p are Promising Novel Biomarkers for Early Diagnosis of Acute Myocardial Infarction. *Cell Physiol Biochem* 2016;38:1015-29.

Submitted Oct 19, 2016. Accepted for publication Oct 23, 2016.

doi: 10.21037/atm.2016.12.12

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.12.12>

Acute myocardial infarction (AMI) is the major cause of death and disability worldwide and the risk of death is highest within the first hours from chest pain onset (1). Hence, an early and accurate diagnosis of AMI in patients presenting with chest pain is of paramount importance for effective treatment and improvement of outcome prognosis (2). In addition to the clinical symptomatic, electrocardiographic and imaging findings, the rise or fall of cardiac troponins, released from the damaged cardiomyocytes, are important biomarkers for the diagnosis of AMI (2). However, troponins can also be released in other cardiac pathologies, such as myocarditis, and chest pain is often misunderstood or difficult to allocate by the patient. Therefore, novel biomarkers are needed to fine-tune AMI diagnosis and potentially enable personalized treatment in the future.

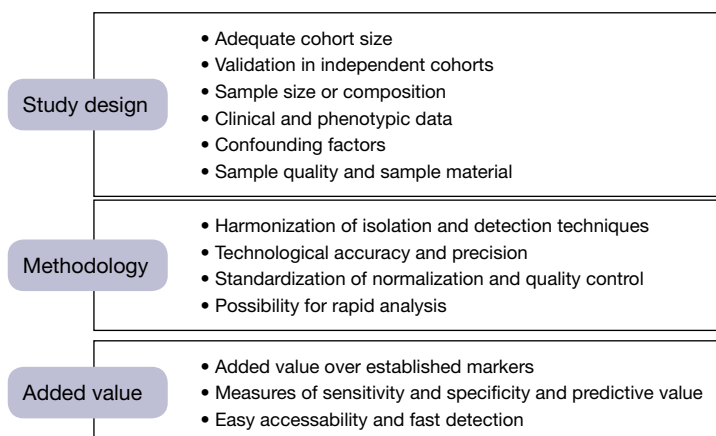
Implementation of novel biomarkers into clinical practice is an important area of biomedical research. A biomarker is a characteristic that is objectively measured and evaluated as an indicator of normal biological processes, pathogenic processes, or pharmacological responses to a therapeutic intervention, and the overall expectation of a biomarker is to enhance the ability of the clinician to optimally manage the patient (3). To evaluate a biomarker, important measures are its sensitivity (the ability to detect the disease, i.e., identification of true-positives) and specificity (the ability to detect absence of disease, i.e., identification of true-negatives), as well as its

positive and negative predictive values (3).

Recently, new technologies have been applied for screening potential novel biomarkers, taking into account not only proteins but also additional promising molecules such as nucleic acids (4). From these analyses, non-coding RNA species have been characterized including microRNAs (miRNAs).

miRNAs are small sequences of endogenous RNA molecules regulating gene expression at a post-transcriptional level (5). miRNAs are involved in a broad range of biological processes and their dysregulation impacts disease development (5). Because of their remarkable stability in biological fluids such as serum/plasma and urine (5) and their tissue-specificity, there is a great interest in miRNAs as novel circulating biomarkers.

Several lines of evidence support this role of miRNAs: studies—using human blood samples as well as animal models, and focusing on the diagnostic potential, compared miRNAs in AMI patients to controls as well as to established biomarkers e.g. troponin (6-9). The most-reported miRNAs were the cardiac-specific miRNAs miR-1, miR-133a, miR-208b and miR-499 (10) that showed high sensitivity and specificity as biomarkers for AMI (10). Similar, miRNAs were identified that facilitate differentiation of unstable angina pectoris (UAP) from stable AP (SAP) or non-coronary chest pain (NCCP). These include the non-cardiac specific miRs 134, 198, 370 (UAP/SAP) and miRs



**Figure 1** Main challenges to overcome in miRNA analyses for diagnostic purposes. miRNA, microRNA.

132, 150, 186 (UAP/NCCP) (11), indicating that also non-cardiac specific miRNAs—in combination - can be of diagnostic value.

In this issue of Cellular Physiology and Biochemistry Wang *et al.* (12) investigated a signature consisting of three (non-cardiac) circulating miRNAs, namely miR-19b-3p, miR-134-5p and miR-186-5p, for its potential use as a biomarker for (fine tuning) AMI diagnosis. The authors present results of a longitudinal assessment of these three miRNAs over different time points after chest pain onset and explored the diagnostic potential of the single miRNAs and in combination. All three miRNAs were upregulated after the onset of chest pain within a range of four to 72 hours, and showed a positive correlation with cardiac troponin I (cTnI). Not surprisingly, levels of cTnI were also increased already at the time of admission. Calculation of the area under the ROC curve (AUC)—as an indicator for the discriminatory performance of the miRNAs, showed good AUC values indicating a discriminatory ability of the three miRNAs—singly and as a combined miRNA score. Interestingly, Wang *et al.* also investigated the influence of heparin and medication on the circulating levels of the three miRNAs and concluded that neither administration of heparin nor the administration of ACE inhibitors, beta-blockers, nitrates, statins, aspirin and clopidogrel) had an influence on the miRNA levels.

Although the concept to explore miRNAs for diagnosis of AMI is very attractive, there are still various challenges to face due to preanalytical and analytical factors influencing data quality. These factors include the study design, the choice of material, isolation, detection and processing techniques as well as normalization strategies, and the

influence of drugs and other, non-cardiac disease and phenotypes (*Figure 1*).

An appropriate study design is a foremost requirement for reliable biomarker identification, ensuring adequate sample size for analysis and accounting for possible confounders (5). Wang *et al.* (12) used a comparatively small group of AMI patients and controls. Aside from the small study cohort, other studies of that purpose usually match one or even two control subjects to one “case” according to a panel of pre-defined factors, rather than merely excluding differences of the overall groups or validate their initial findings in much larger independent study cohorts. The three miRNAs investigated in the study of Wang *et al.* (12) have all been described already to be involved in development as well as pathologies of the heart (13), such as heart failure, but also in other context, such as gastric cancer (14). Therefore, great attention needs to be paid to the selection of case as well as control groups and potential confounding factors influencing miRNA levels. Furthermore, only a small number of non-cardiac miRNAs have been selected by the authors, apparently based on the current literature. Thus, other miRNA with potentially better performance might have been missed.

The detection of miRNAs currently relies on real-time PCR techniques, and thus depends on the limitations raised by the current lack of standardized isolation, detection and normalization strategies. To date, several approaches are used including manual or commercial isolation techniques, different reverse transcription and PCR approaches as well as different normalization methods such as the use of synthetic spike-in material (as in the current study) or the use of an average of the Ct values of all tested miRs (4,5).

To eliminate this technical and analytical variability and thereby avoiding artifactual data generation, consensus on standard methods for all steps is imperative.

Of great interest is that Wang *et al.* (12) directly investigated the effect of heparin and medication on the miRNA levels of interest. However—surprisingly—no significant influence on miRNA levels were found, although at least for miR-19b-3p and miR-186-5p a trend was observed which might reach statistical significance with a higher number of patients tested. Indeed, several other studies have demonstrated effects of statins and anti-platelet drugs on the levels of various circulating miRNAs (15). And heparin is known to interfere with a number of diagnostic markers and had previously been described to alter circulating miRNA levels (16,17).

For the translation of a potential biomarker candidate into clinical application it is of importance to investigate whether the novel marker shows an added value to established markers. For this, potential novel biomarkers for the diagnosis of AMI, i.e., circulating miRNAs, need to be compared directly to cardiac troponins—the gold-standard biomarker currently used for diagnosis of AMI. Since the introduction of highly sensitive troponin assays into clinical practice already substantially improved the early diagnosis of AMI, a direct comparison of the circulating miRNAs to high sensitivity assayed troponins is mandatory. Also, to accurately judge the diagnostic performance of circulating miRNAs, measures such as sensitivity, specificity as well as negative and positive predictive values are needed.

Lastly, independent of the purely methodological aspects, research is also required to address other relevant questions concerning the translation of miRNAs into clinical practice, such as a feasible time frame for qPCR analysis before patient treatment, ethical issues as well as optimal cost-benefit analysis.

## Acknowledgements

None.

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

1. Twerenbold R, Reichlin T, Reiter M, et al. High-sensitive cardiac troponin: friend or foe? *Swiss Med Wkly* 2011;141:w13202.
2. Keller T, Zeller T, Peetz D, et al. Sensitive troponin I assay in early diagnosis of acute myocardial infarction. *N Engl J Med* 2009;361:868-877.
3. Zeller T, Blankenberg S. Blood-based gene expression tests: promises and limitations. *Circ Cardiovasc Genet* 2013;6:139-140.
4. Santovito D, Weber C. Zooming in on microRNAs for refining cardiovascular risk prediction in secondary prevention. *Eur Heart J* 2016. [Epub ahead of print].
5. Siemelink MA, Zeller T. Biomarkers of coronary artery disease: the promise of the transcriptome. *Curr Cardiol Rep* 2014;16:513.
6. Wang GK, Zhu JQ, Zhang JT, et al. Circulating microRNA: a novel potential biomarker for early diagnosis of acute myocardial infarction in humans. *Eur Heart J* 2010;31:659-666.
7. D'Alessandra Y, Devanna P, Limana F, et al. Circulating microRNAs are new and sensitive biomarkers of myocardial infarction. *Eur Heart J* 2010;31:2765-2773.
8. Olivieri F, Antonicelli R, Lorenzi M, et al. Diagnostic potential of circulating miR-499-5p in elderly patients with acute non ST-elevation myocardial infarction. *Int J Cardiol* 2013;167:531-536.
9. Devaux Y, Mueller M, Haaf P, et al. Diagnostic and prognostic value of circulating microRNAs in patients with acute chest pain. *J Intern Med* 2015;277:260-271.
10. Schulte C, Zeller T. microRNA-based diagnostics and therapy in cardiovascular disease—Summing up the facts. *Cardiovasc Diagn Ther* 2015;5:17-36.
11. Zeller T, Keller T, Ojeda F, et al. Assessment of microRNAs in patients with unstable angina pectoris. *Eur Heart J* 2014;35:2106-2114.
12. Wang KJ, Zhao X, Liu YZ, et al. Circulating MiR-19b-3p, MiR-134-5p and MiR-186-5p are Promising Novel Biomarkers for Early Diagnosis of Acute Myocardial Infarction. *Cell Physiol Biochem* 2016;38:1015-1029.
13. Zou M, Wang F, Gao R, et al. Autophagy inhibition of hsa-miR-19a-3p/19b-3p by targeting TGF- R II during TGF- 1-induced fibrogenesis in human cardiac fibroblasts. *Sci Rep* 2016;6:24747.
14. Zhang J, Song Y, Zhang C, et al. Circulating MiR-16-5p and MiR-19b-3p as Two Novel Potential Biomarkers to Indicate Progression of Gastric Cancer. *Theranostics* 2015;5:733-745.
15. Willeit P, Zampetaki A, Dudek K, et al. Circulating microRNAs as novel biomarkers for platelet activation.

- Circ Res 2013;112:595-600.
16. Kaudewitz D, Zampetaki A, Mayr M. MicroRNA Biomarkers for Coronary Artery Disease? *Curr Atheroscler Rep* 2015;17:70.
  17. Boeckel JN, Thomé CE, Leistner D, et al. Heparin selectively affects the quantification of microRNAs in human blood samples. *Clin Chem* 2013;59:1125-1127.

**Cite this article as:** Kränkel N, Blankenberg S, Zeller T. Early detection of myocardial infarction—microRNAs right at the time? *Ann Transl Med* 2016;4(24):502. doi: 10.21037/atm.2016.12.12



# Circulating micro ribonucleic acids in cardiovascular disease: a look beyond myocardial injury

Johannes Mair

Department of Internal Medicine III – Cardiology and Angiology, Innsbruck Medical University, A-6020 Innsbruck, Austria

*Correspondence to:* Johannes Mair, MD. Department of Internal Medicine III – Cardiology and Angiology, Innsbruck Medical University, Anichstrasse 35, A-6020 Innsbruck, Austria. Email: Johannes.Mair@i-med.ac.at.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, Jiangsu, China).

*Comment on:* Navickas R, Gal D, Laucevičius A, *et al.* Identifying circulating microRNAs as biomarkers of cardiovascular disease: a systematic review. *Cardiovasc Res* 2016;111:322-37.

Submitted Sep 30, 2016. Accepted for publication Oct 05, 2016.

doi: 10.21037/atm.2016.10.67

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.10.67>

Laboratory markers are of significant clinical importance in the evaluation of patients with suspected cardiac diseases. They have evolved as essential tools in cardiology over the last 50 years, i.e., lipid testing for primary and secondary prevention, creatine kinase isoenzyme MB and subsequently the more sensitive and specific cardiac troponin (cTn) testing for the diagnosis and management of acute myocardial infarction (AMI), and more recently natriuretic peptide (NP) testing for the diagnosis (in particular exclusion), risk stratification, and monitoring of heart failure (HF) (1-3). We are beginning an era when it may be possible for biomarkers to direct treatment to optimize patient management. This is already the case with cTn (1,4) but should be the final goal with all cardiac biomarkers. However, there are still some open major clinical issues, e.g., the diagnosis of myocardial ischemia. Despite huge research efforts in recent years, which were triggered by the great clinical significance and economic impact of cardiac diseases, biomarkers for the prediction of coronary artery disease (CAD) and for risk stratification in stable CAD or the general population have not yet fulfilled their manifest promise so far (5). The most established marker in this respect is high-sensitivity C-reactive protein (hs-CRP) which still remains controversial (3,6).

The 1990s were the golden era of cardiac biomarkers with the implementation of cTn and NP routine testing. Numerous additional biomarkers were discovered and immunoassays were developed which were also suitable

for routine measurement. The main focus was on markers of coronary plaque formation, plaque destabilization (e.g., myeloperoxidase), intracoronary thrombus formation (coagulation and platelet activation, reduced endogenous fibrinolytic activity), and markers of myocardial ischemia (e.g., ischemia modified albumin). However, the vast majority of these markers did not make the way from research to routine application due to either preanalytical, analytical issues, or because the clinical impact for risk stratification was limited as these markers did not add much to traditional risk factors and even in multimarker approach improved risk stratification and patient reclassification only very modestly above established routine biomarkers (5). Importantly, they did not lead to direct information about how to improve patient management. More recently, copeptin, a very unspecific marker of endogenous stress, was suggested for rapidly ruling out AMI in the emergency department. However, no significant benefit compared to high-sensitivity cTn (hs-cTn) testing could be convincingly demonstrated (7). During this period also genomic biomarkers entered the field and have been particularly popular in the last two decades. Almost all of the candidate-gene era genetic biomarkers of cardiovascular disease failed to be validated after an initial period of enthusiasm (8). Rare variants may be potent but because they are rare, they do not identify large numbers of additional patients at risk. Common variants such as single genetic variants confer extremely small risks such that the usual way of calculating



risk by assessment of traditional cardiovascular risk factors is better than analyses for these commonly occurring variations in deoxy ribonucleic acid (DNA) sequences. Consequently, the current consensus is not to test for commonly occurring genetic variants with weak effects (9).

Another currently very popular research topic is circulating plasma micro ribonucleic acid (miRNA) testing (10). miRNAs are small (typically less than 25 nucleotides), single-stranded, endogenous, non-coding RNAs that post-transcriptionally regulate gene expression by destabilizing messenger RNA (mRNA) or translation repression and thereby preventing proteins synthesis (11,12). Interestingly, each miRNA can target several mRNA while each mRNA can be targeted by multiple miRNAs (12). Eventually miRNAs are secreted from cells into blood being packaged in microparticles, but they are also found bound with proteins or high-density lipoproteins. The biological function of circulating miRNAs remains to be established. It is unclear whether circulating miRNAs are messengers in the cell-to-cell communication with active secretion or merely degradation products without any biological function with passive release as necrosis associated biomarkers.

More than 1,000 miRNAs have been identified in the human genome, but based on their tissue distribution and physiological function in the regulation of angiogenesis, apoptosis, and cell differentiation and proliferation miRNA-1, -133, -145, -208, and -499 appear to be most promising candidate markers for testing their diagnostic and prognostic potential in cardiovascular diseases (10). Regarding cardiac-specificity miRNAs-208 and -499 are promising, and in fact, particularly miRNA-499 and miRNA-208b were evaluated in patients with suspected acute coronary syndromes (ACS) with a rapid increase early after AMI with a high sensitivity within 3 hours from symptom onset (13). The hope still is to identify a miRNA profile (e.g., miRNA-1, -499, and -21) specific for myocardial ischemia (14), which would be of particular clinical interest. First studies, however, could not demonstrate an additive value of miRNA to hs-cTn testing for AMI diagnosis (13). In patients with CAD miRNA-132, miRNA-150, and miRNA-186 appear to be associated with ACS (15), and miRNAs (e.g., miRNA-145) appear to be associated with presence of CAD as well (16). However, the published data on the value of miRNAs for diagnosis and in particular for risk stratification in various cardiac diseases is still contradictory and inconclusive (10), and large clinical studies with appropriate pre-analytics and analytics remain to be done to demonstrate the additive value of miRNA

measurement to conventional cardiac biomarker testing convincingly.

Currently miRNA testing is also time consuming with demanding pre-analytics and analytics (10), which precludes widespread routine use. It is very important to prepare cell free plasma to avoid in-vitro contamination from blood cells, but the methods of plasma preparation are frequently not sufficiently given in publications. Hemolysis must be avoided during blood collection and should be ruled out by oxyhemoglobin testing before miRNA testing in plasma samples. Whole blood must be processed immediately for plasma preparation as well. *In vitro* miRNA contamination from blood cells may be a particular problem if miRNAs are tested in stored frozen plasma samples which were not collected and prepared with the aim of testing miRNAs, and consequently this may lead to erroneous results and publications. Heparin plasma is not suitable for miRNA testing because heparin may inhibit complementary DNA (cDNA) synthesis and polymerase chain reaction (PCR), and quantitative reverse transcription PCR (qRT-PCR) is the most widely used method for circulating miRNA determination. Thus, it is also important to know whether patients were treated with heparin before blood collection. Another unresolved issue is the lack of harmonization of methods and of test result normalization (e.g., synthetic spike-in control miRNAs *vs.* expression or mean expression value of one or better a panel of commonly expressed miRNAs in a sample that are not associated with diseases), which makes it very difficult to compare published study results. Synthetic spike-in RNAs have the additional advantage that this can be also used to monitor the efficiency of RNA isolation, cDNA synthesis, and PCR amplification as well as to reveal potential presence of nucleases in the sample.

In conclusion, the role of biomarkers in cardiovascular diseases, such as AMI and HF, is very well established with cTn and NP testing as essential parts of patient evaluation with suspected AMI or HF (1,2,17,18). Given this powerful role of established cardiac biomarkers it is very difficult to demonstrate a significant benefit of add-on testing of new biomarkers compared with established markers (i.e., hs-cTn, NP, and hs-CRP) in cardiac diseases. Therefore, as with other heavily investigated novel markers, the coming years will show whether miRNA testing will make the way from research to routine use after an initial hype at the beginning of research, particularly as hs-cTn assays already entered routine use (17) and as even more sensitive research cTn assays (“ultra-sensitive”) have been developed with

significant clinical potential (17,19). In contrast to standard cardiac biomarker testing including ultra-sensitive cTn a lot of pre-analytical and analytical issues of miRNA testing also have to be solved before routine testing is feasible.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The author has no conflicts of interest to declare.

### References

1. Thygesen K, Alpert JS, Jaffe AS, et al. Third universal definition of myocardial infarction. *Eur Heart J* 2012;33:2551-2567.
2. Ponikowski P, Voors AA, Anker SD, et al. 2016 ESC Guidelines for the diagnosis and treatment of acute and chronic heart failure: The Task Force for the diagnosis and treatment of acute and chronic heart failure of the European Society of Cardiology (ESC) Developed with the special contribution of the Heart Failure Association (HFA) of the ESC. *Eur Heart J* 2016;37:2129-2200.
3. Catapano AL, Graham I, De Backer G, et al. ESC/EAS Guidelines for the Management of Dyslipidaemias: The Task Force for the Management of Dyslipidaemias of the European Society of Cardiology (ESC) and European Atherosclerosis Society (EAS) Developed with the special contribution of the European Association for Cardiovascular Prevention & Rehabilitation (EACPR). *Eur Heart J* 2016.2016. [Epub ahead of print].
4. Roffi M, Patrono C, Collet JP, et al. 2015 ESC Guidelines for the management of acute coronary syndromes in patients presenting without persistent ST-segment elevation: Task Force for the Management of Acute Coronary Syndromes in Patients Presenting without Persistent ST-Segment Elevation of the European Society of Cardiology (ESC). *Eur Heart J* 2016;37:267-315.
5. Melander O, Newton-Cheh C, Almgren P, et al. Novel and conventional biomarkers for prediction of incident cardiovascular events in the community. *JAMA* 2009;302:49-57.
6. Biasucci LM, Koenig W, Mair J, et al. How to use C-reactive protein in acute coronary care. *Eur Heart J* 2013;34:3687-3690.
7. Reinstadler SJ, Klug G, Feistritz HJ, et al. Copeptin testing in acute myocardial infarction: ready for routine use? *Dis Markers* 2015;2015:614145.
8. Ntzani EE, Rizos EC, Ioannidis JP. Genetic effects versus bias for candidate polymorphisms in myocardial infarction: case study and overview of large-scale evidence. *Am J Epidemiol* 2007;165:973-984.
9. Faergeman O. Genes and cardiovascular risk. *Eur Heart J* 2013;34:949-950.
10. Navickas R, Gal D, Laucevičius A, et al. Identifying circulating microRNAs as biomarkers of cardiovascular disease: a systematic review. *Cardiovasc Res* 2016;111:322-337.
11. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281-297.
12. Bauersachs J, Thum T. Biogenesis and regulation of cardiovascular microRNAs. *Circ Res* 2011;109:334-347.
13. Devaux Y, Vausort M, Goretti E, et al. Use of circulating microRNAs to diagnose acute myocardial infarction. *Clin Chem* 2012;58:559-567.
14. Oerlemans MI, Mosterd A, Dekker MS, et al. Early assessment of acute coronary syndromes in the emergency department: the potential diagnostic value of circulating microRNAs. *EMBO Mol Med* 2012;4:1176-1185.
15. Zeller T, Keller T, Ojeda F, et al. Assessment of microRNAs in patients with unstable angina pectoris. *Eur Heart J* 2014;35:2106-2114.
16. Gao H, Guddeti RR, Matsuzawa Y, et al. Plasma Levels of microRNA-145 Are Associated with Severity of Coronary Artery Disease. *PLoS One* 2015;10:e0123477.
17. Thygesen K, Mair J, Giannitsis E, et al. How to use high-sensitivity cardiac troponins in acute cardiac care. *Eur Heart J* 2012;33:2252-2257.
18. Thygesen K, Mair J, Mueller C, et al. Recommendations for the use of natriuretic peptides in acute cardiac care: a position statement from the Study Group on Biomarkers in Cardiology of the ESC Working Group on Acute Cardiac Care. *Eur Heart J* 2012;33:2001-2006.
19. Bonaca MP, O'Malley RG, Murphy SA, et al. Prognostic performance of a high-sensitivity assay for cardiac troponin I after non-ST elevation acute coronary syndrome: Analysis from MERLIN-TIMI 36. *Eur Heart J Acute Cardiovasc Care* 2015;4:431-440.

**Cite this article as:** Mair J. Circulating micro ribonucleic acids in cardiovascular disease: a look beyond myocardial injury. *Ann Transl Med* 2016;4(Suppl 1):S30. doi: 10.21037/atm.2016.10.67

# Clinical utility of novel biomarkers in acute myocardial infarction

Thomas Stiermaier<sup>1,2</sup>, Holger Thiele<sup>1,2</sup>, Ingo Eitel<sup>1,2</sup>

<sup>1</sup>Department of Cardiology, Angiology, Intensive Care Medicine, Medical Clinic II, University Heart Center of Lübeck, Lübeck, Germany; <sup>2</sup>German Center for Cardiovascular Research (DZHK), partner site Hamburg/Kiel/Lübeck, Lübeck, Germany

*Correspondence to:* Ingo Eitel, MD. Medical Clinic II, University Heart Center Lübeck, University Hospital Schleswig-Holstein, Ratzeburger Allee 160, 23538 Lübeck, Germany. Email: ingo.eitel@uksh.de.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Feistritzer HJ, Klug G, Reinstadler SJ, *et al.* Novel biomarkers predicting cardiac function after acute myocardial infarction. *Br Med Bull* 2016;119:63-74.

Submitted Oct 07, 2016. Accepted for publication Oct 12, 2016.

doi: 10.21037/atm.2016.12.06

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.12.06>

Over the past decades, biomarkers of myocardial injury, particularly cardiac troponin (cTn), creatine kinase and its isoenzyme creatine kinase myocardial band, have been extremely valuable for the diagnosis and clinical decision making in patients with suspected acute coronary syndrome (ACS). These established markers of necrosis were recently complemented by numerous novel biomarkers reflecting causes and consequences of myocardial infarction (e.g., inflammation, endothelial dysfunction, or hemodynamic stress). Therefore, research efforts have been directed to determine the additional diagnostic and prognostic value of these novel biomarkers.

While the diagnosis and management of ST-elevation myocardial infarction is solely based on clinical and electrocardiographic findings, the identification of patients with non-ST-elevation myocardial infarction (NSTEMI) requires a more sophisticated approach including the measurement of cTn and other biomarkers of myocardial necrosis (1,2). In patients with NSTEMI, levels of cTn usually rise rapidly within 1 hour after symptom onset and remain elevated for a variable period of time (1). Advances in technology have led to an improvement in cTn assays and have refined the ability to detect and quantify myocardial injury. By using conventional, non-high-sensitive cTn (hs-cTn) assays elevated troponin concentrations in the peripheral blood may be detected delayed which necessitates serial testing to ascertain the diagnosis in most cases. Novel biomarkers were thought to bridge this “troponin-blind” gap and enable immediate decision making in patients

presenting with acute chest pain.

Copeptin has emerged as one of these promising biomarkers that may overcome this lack of sensitivity within the first hours after symptom onset although non-specific to myocardial injury. It is released in response to endogenous and/or hemodynamic stress and a dual-marker strategy with conventional cTn and copeptin showed a high negative predictive value for the early rule-out of NSTEMI including a reduction of the average time-to-diagnosis, therefore adding incremental value to conventional cTn assays (3,4). However, the introduction of hs-cTn assays in clinical routine significantly increased the diagnostic performance of cTn and facilitates to rule out myocardial infarction within 1 hour (1). An additional diagnostic value of copeptin to hs-cTn has not been convincingly established (5,6). Consequently, current guidelines recommend the routine use of copeptin for the early rule-out of NSTEMI only if hs-cTn assays are not available (1).

In view of the limited diagnostic gain in addition to hs-cTn, the focus shifted to the prognostic value of novel cardiac biomarkers. In the *British Medical Bulletin*, Feistritzer and colleagues provide a systematic review of selected biomarkers reflecting myocardial injury, inflammation/fibrosis, and hemodynamics (7). The authors illustrate the pathophysiological background and the prognostic utility of hs-cTn, natriuretic peptides, copeptin, galectin-3, corin, fetuin-A, adiponectin and micro-RNAs. Scientific advances created numerous other markers that may reflect further aspects of coronary artery disease (e.g., plaque

destabilization/rupture or platelet activation) but would exceed the scope of this review (8). In the context of prognostic utility, growth differentiation factor 15 (GDF-15) showed promising results as an independent predictor of mortality and a potential tool to guide therapy and should therefore be mentioned additionally (8,9). However, although most of these novel biomarkers can improve risk stratification in patients with acute myocardial infarction to some extent, the question regarding their actual role in clinical routine arises (8). As nicely illustrated by Feistritzer and colleagues, the obligatory measurement of hs-cTn already provides substantial prognostic information and is pivotal for patient management (7). In addition, only natriuretic peptides were extensively validated and have proven prognostic utility on top of cTn although without definitive treatment implications (10). Other novel biomarkers have not yet shown useful regarding therapeutic decision making and their incremental prognostic value over and above established biomarkers and risk scores is only marginal. Similarly, multi-marker approaches involving several novel biomarkers failed to clearly outperform the prognostic usefulness of cTn (11). Therefore, guidelines for the management of patients with acute myocardial infarction do not recommend their routine assessment (1,2).

Several aspects might explain why novel biomarkers have not yet found their way into clinical routine. Established biomarkers, basically hs-cTn and natriuretic peptides, as well as clinical scores such as the Global Registry of Acute Coronary Events (GRACE) or the Thrombolysis In Myocardial Infarction (TIMI) Risk Score enable excellent risk stratification, are widely available and cost effective (1). Moreover, imaging modalities provide further prognostic insights. Echocardiography to evaluate left ventricular function and contraction abnormalities is recommended in all patients with acute myocardial infarction (1,2). Cardiovascular magnetic resonance (CMR) imaging allows for a more detailed tissue characterization and is increasingly available in clinical routine. Parameters derived from these imaging modalities, particularly CMR, have demonstrated incremental prognostic information regarding hard clinical endpoints whereas many biomarkers have only been linked to impaired left ventricular function or adverse remodeling (12). Therefore, the combination of established biomarkers, clinical risk scores and imaging parameters facilitates a sufficient risk stratification from a clinician's point of view. Novel biomarkers have to outperform these markers regarding sensitivity, specificity and/or a quicker release kinetic while being similarly cost effective and easily

available to play a role in clinical practice. Since this is very difficult to achieve a potential alternative application of novel biomarkers is the guidance of treatment decisions to enable a more tailored patient management. However, a benefit of biomarker based approaches remains to be proven.

In summary, several novel biomarkers reflecting different pathophysiological aspects of acute myocardial infarction emerged during the last decade. However, few of them have proven valuable in clinical routine given the excellent diagnostic and prognostic performance of established evaluation strategies including cTn, clinical risk scores and imaging markers of myocardial damage. While cTn is highly specific for myocardial necrosis, it provides no information on the etiology of myocyte death. Thus, more research is necessary to determine new thresholds or additional helpful biomarkers to distinguish between etiologies in the heterogeneous field of ACS (e.g., in patients with atrial fibrillation, hypertension, Takotsubo syndrome, and/or kidney disease). Finally, an individualized biomarker-guided management according to cTn and other promising biomarkers like copeptin and/or GDF-15 (e.g., immediate coronary intervention in NSTEMI patients with high-risk biomarker features) should be addressed in upcoming ACS trials to evaluate if there is room for other novel biomarkers for clinical decision making next to cTn.

## Acknowledgements

None.

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

1. Roffi M, Patrono C, Collet JP, et al. 2015 ESC Guidelines for the management of acute coronary syndromes in patients presenting without persistent ST-segment elevation: Task Force for the Management of Acute Coronary Syndromes in Patients Presenting without Persistent ST-Segment Elevation of the European Society of Cardiology (ESC). *Eur Heart J* 2016;37:267-315.
2. Task Force on the management of ST-segment elevation acute myocardial infarction of the European Society of Cardiology (ESC), Steg PG, James SK, et al. ESC

- Guidelines for the management of acute myocardial infarction in patients presenting with ST-segment elevation. *Eur Heart J* 2012;33:2569-619.
3. Maisel A, Mueller C, Neath SX, et al. Copeptin helps in the early detection of patients with acute myocardial infarction: primary results of the CHOPIN trial (Copeptin Helps in the early detection Of Patients with acute myocardial INfarction). *J Am Coll Cardiol* 2013;62:150-160.
  4. Möckel M, Searle J, Hamm C, et al. Early discharge using single cardiac troponin and copeptin testing in patients with suspected acute coronary syndrome (ACS): a randomized, controlled clinical process study. *Eur Heart J* 2015;36:369-376.
  5. Giannitsis E, Kehayova T, Vafaie M, et al. Combined testing of high-sensitivity troponin T and copeptin on presentation at prespecified cutoffs improves rapid rule-out of non-ST-segment elevation myocardial infarction. *Clin Chem* 2011;57:1452-1455.
  6. Karakas M, Januzzi JL Jr, Meyer J, et al. Copeptin does not add diagnostic information to high-sensitivity troponin T in low- to intermediate-risk patients with acute chest pain: results from the rule out myocardial infarction by computed tomography (ROMICAT) study. *Clin Chem* 2011;57:1137-1145.
  7. Feistritz HJ, Klug G, Reinstadler SJ, et al. Novel biomarkers predicting cardiac function after acute myocardial infarction. *Br Med Bull* 2016;119:63-74.
  8. Hochholzer W, Morrow DA, Giugliano RP. Novel biomarkers in cardiovascular disease: update 2010. *Am Heart J* 2010;160:583-594.
  9. Eitel I, Blase P, Adams V, et al. Growth-differentiation factor 15 as predictor of mortality in acute reperfused ST-elevation myocardial infarction: insights from cardiovascular magnetic resonance. *Heart* 2011;97:632-640.
  10. Thygesen K, Mair J, Mueller C, et al. Recommendations for the use of natriuretic peptides in acute cardiac care: a position statement from the Study Group on Biomarkers in Cardiology of the ESC Working Group on Acute Cardiac Care. *Eur Heart J* 2012;33:2001-2006.
  11. O'Malley RG, Bonaca MP, Scirica BM, et al. Prognostic performance of multiple biomarkers in patients with non-ST-segment elevation acute coronary syndrome: analysis from the MERLIN-TIMI 36 trial (Metabolic Efficiency With Ranolazine for Less Ischemia in Non-ST-Elevation Acute Coronary Syndromes-Thrombolysis In Myocardial Infarction 36). *J Am Coll Cardiol* 2014;63:1644-1653.
  12. Eitel I, de Waha S, Wöhrle J, et al. Comprehensive prognosis assessment by CMR imaging after ST-segment elevation myocardial infarction. *J Am Coll Cardiol* 2014;64:1217-1226.

**Cite this article as:** Stiermaier T, Thiele H, Eitel I. Clinical utility of novel biomarkers in acute myocardial infarction. *Ann Transl Med* 2016;4(24):491. doi: 10.21037/atm.2016.12.06

# Circulating microRNA biomarkers for cardiovascular risk prediction: are we approaching clinical application?

Maurice W. J. de Ronde<sup>1,2</sup>, Yigal M. Pinto<sup>3</sup>, Sara-Joan Pinto-Sietsma<sup>1,2</sup>

<sup>1</sup>Department of Vascular Medicine, <sup>2</sup>Department of Clinical Epidemiology, Biostatistics and Bioinformatics, <sup>3</sup>Department of Experimental Cardiology, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands

*Correspondence to:* Sara-Joan Pinto-Sietsma. Academic Medical Center, Meibergdreef 9, 1105 AZ, Room J1B-214, Amsterdam, The Netherlands. Email: pintosj@gmail.com.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Karakas M, Schulte C, Appelbaum S, *et al.* Circulating microRNAs strongly predict cardiovascular death in patients with coronary artery disease—results from the large AtheroGene study. *Eur Heart J* 2016. [Epub ahead of print].

Submitted Oct 07, 2016. Accepted for publication Oct 12, 2016.

doi: 10.21037/atm.2016.12.05

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.12.05>

Patients at high risk of cardiovascular morbidity and mortality are still difficult to identify. Unfortunately, the most widely used standard risk prediction models predict cardiovascular disease (CVD) rather poorly, since these risk calculators are mostly driven by age. Therefore, most young subjects are regarded as low risk merely based on their age, despite sometimes quite obvious adverse risk factors. Circulating biomarkers that can help to improve the identification of individuals at risk are therefore highly needed.

MicroRNAs are short noncoding RNAs that post-transcriptionally regulate gene expression (1). When microRNAs are shed into the circulation, they remain there in a stable state and reflect ongoing processes at a cellular level in the tissues (2,3). Because of these properties, circulating microRNAs have become increasingly popular over the years.

After many small, exploratory studies with divergent results, presently, also larger studies on circulating microRNAs are being published. In a recent study, Karakas *et al.* (4), assessed the prognostic value of 8 circulating microRNAs that had previously been identified to facilitate the diagnosis of unstable angina pectoris (UAP) (5) in a cohort of 430 acute coronary syndrome (ACS) patients and 682 stable angina pectoris (SAP) patients. The authors concluded that 7 of these 8 measured microRNAs are strongly predictive of cardiovascular mortality with areas under the receiver-operating curves (AUCs) up to 0.76;

this represents an important step forward in the circulating microRNA field. However, these studies still struggle with many common pitfalls in microRNA measurement and interpretation of the results remains challenging.

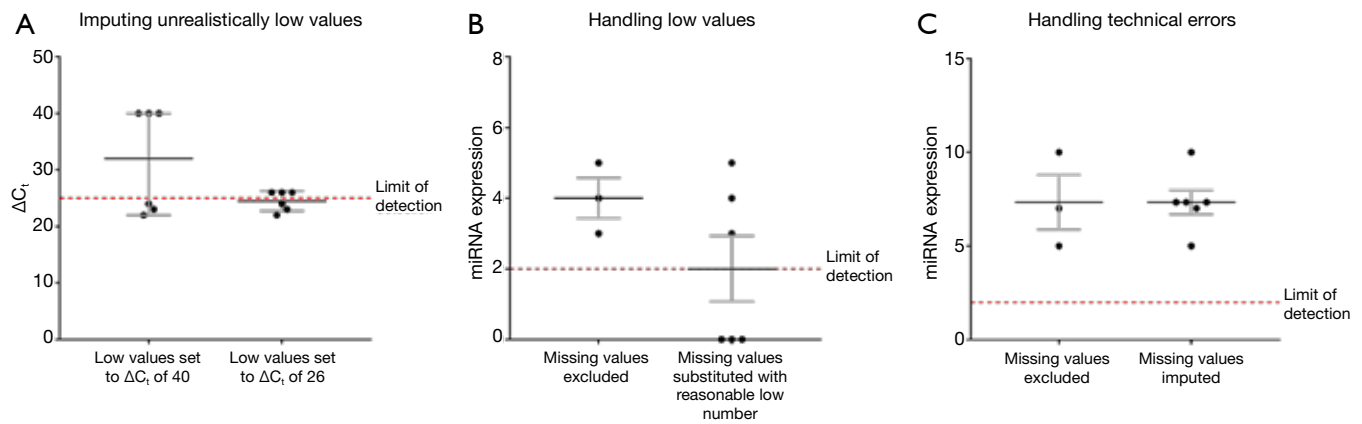
## Pitfalls in the measurement of circulating microRNAs and poor reporting on reverse transcription quantitative polymerase chain reaction (RT-qPCR) methods

Reverse transcription quantitative polymerase chain reaction (RT-qPCR) is the most common way to reliably assess microRNA levels. However, the way in which RT-qPCR measurements are performed and analyzed can have a large influence on the results. It is therefore of great importance to have standardized and validated methods for RT-qPCR microRNA measurement. The authors of the study of Karakas *et al.* (4) thoroughly report on statistical methods, but do not report on the details of the RT-qPCR measurement nor the handling of missing data. It is important to describe the RT-qPCR methods following the MIQE guidelines (6) and to describe how missing data was handled. Differences herein can lead to serious flaws and biases in the results, as explained more in detail below.

## Handling missing data and low values

MicroRNAs often circulate in a very low concentration.





**Figure 1** Pitfalls of microRNA data handling. The following pitfalls can be expected from wrong data handling. (A) shows the effect of using an unrealistically low value to substitute  $C_t$  values  $\geq 40$ . Due to a large gap between the lowest measurable  $\Delta C_t$  of 24.99 and the substituted  $\Delta C_t$  of 40, there is more variation in the data, creating bias in the analysis. Dots are fictional data with mean (black line) and standard deviation (grey whiskers). This is solved by using a more appropriate  $\Delta C_t$  to substitute these low values such as a  $\Delta C_t$  of 26; (B) shows the effect of excluding missing values due to low microRNA expression. When these values are excluded, this falsely leads to increased expression levels. Therefore, missing values due to low microRNA expression must be substituted with a low value (e.g., 0); (C) shows the effect of excluding missing values that occurred due to technical errors. Excluding instead of imputing these missing values from the analysis will lead to an increased standard error of the mean and may therefore mask significance. In (B) and (C), dots are fictional data with mean (black line) and standard error of the mean (grey whiskers).

Even highly sensitive methods like RT-qPCR, need to be stretched to their limit of detection and sometimes certain microRNAs are even totally absent from the circulation. This most often results in missing values that complicate the measurement of microRNAs, since microRNAs can also appear to be missing due to a technical error.

It is most important to distinguish missing values due to a low concentration or complete absence of the microRNA from the circulation, from values that are missing due to a technical error. Missing data due to a low concentration is not missing at random and therefore should be substituted with a low value, whereas missing data due to technical errors occur at random and must be imputed.

The authors of Karakas *et al.* (4) acknowledge the issues related to a low microRNA concentration, since they substitute  $C_t$  values  $\geq 40$  with a  $\Delta C_t$  of 40. Unfortunately, by doing so, they substitute the low microRNA concentrations with an unrealistically low value. We will explain this in detail. In the study, the formula  $C_t(\text{microRNA}) - C_t(\text{cel-miR-39})$  was used to calculate the  $\Delta C_t$ . To produce a reliable qPCR curve based on enough data points to calculate a  $C_t$  from, a minimum  $C_t$  value of 15 is needed. Therefore, the maximum  $\Delta C_t$  that can be calculated is  $[(\text{maximum } C_t \text{ of the microRNA} = 39.99) - (\text{minimum } C_t \text{ cel-}$

$\text{miR-39} = 15)] = 24.99$ . However, in case of a  $C_t$  value of  $\geq 40$ , the authors set the  $\Delta C_t$  to 40, creating a large gap between the lowest value measurable ( $\Delta C_t$  of 24.99) and the substituted low value. This value will contribute tremendously to the mean expression level compared to more realistic substitution values such as a  $\Delta C_t$  of 26 (Figure 1A). If not accounted for, these unrealistic values can influence the results of cox or logistic regression analyses.

Concerning missing values, the authors do not report whether they encountered this and how this was handled. A missing value can occur because the  $C_t$  is 0, meaning that the microRNA was under the detection limit, which also represents a low concentration. Therefore,  $C_t$  values of 0 should also be imputed with a reasonably low number, instead of simply excluding them, which could lead to false higher expression levels (Figure 1B).

A missing value can also occur due to a technical error. By excluding these data, the study will lose power and meaningful differences can be lost as illustrated in Figure 1C. The best way to handle this type of missing data is by a multiple imputation method, in which a weighted average is imputed, taking into account the characteristics of the study population. Failure to handle this issue, might lead to either under- or overestimation of the effect.



### Single versus duplicate or triplicate RT-qPCR measurements

Another major issue concerning the reliability of RT-qPCR measurements of microRNA data is that often single measurements per sample are performed, where duplicate or even triplicate measurements would be more appropriate. Using measurements in duplicate or triplicate highly increases the precision of the microRNA measurements. Besides, replicate measurements are essential to distinguish missing data due to the technical errors from missing data because of low expression, as explained above.

If a single microRNA measurement shows a  $C_t$  value of 0, one would falsely decide that the microRNA is not detectable within the sample, whereas if the measurement would have been done in triplicates, the other two  $C_t$  values might have had values of for instance 32. In this particular example one would conclude, that the first measurement has a technical error and that the true value is the average of the other two measurements. On the other hand, when all three triplicates have a  $C_t$  of 0, it is most likely that this microRNA cannot be measured due to low expression. Unfortunately, the authors do not report whether their RT-qPCR measurements were single or multiple measurements.

### Normalization methods

As the authors state in the discussion, another potential source of bias is the normalization with cel-miR-39. Since cel-miR-39 is not incorporated in microvesicles or protein or lipid complexes, variations in extraction of microRNAs from these vesicles and complexes are not accounted for (7). Therefore, we recently proposed a normalization methods using a panel of endogenous microRNAs, best representing stability of the data and taking in to account technical failures during the RT-qPCR measurement (8). We therefore suggest using these specific panels for specific samples.

### Interpretation of the study results

The authors of the paper by Karakas *et al.* (4) nicely show that circulating microRNAs predict cardiovascular death in coronary artery disease. This is one of the first papers addressing such an important issue in such a large population. Besides, the observed association of 7 of the 8 microRNAs related to cardiovascular mortality, is fairly

strong. On the other hand, because the authors do not report on the number of death and the data handling as discussed above, the observed results might be slightly overestimated. Remarkably, this important observation is mainly present in the ACS group, since when analyzing the overall group, the observed association is slightly attenuated.

The authors speculate on the pathophysiological process behind these microRNAs and state that they could be hypoxia markers. On the other hand, it appears from Table 2 in the paper, that the expression level of all the eight microRNAs in ACS are actually lower than in SAP (higher  $\Delta C_t$  = lower expression level). Therefore, it seems that this intriguing suggestion cannot directly be support by these data. The authors show another intriguing observation, namely, that the association is influenced by vessel disease and left ventricular ejection fraction (LVEF). When the authors correct their data for the number of diseased vessels and LVEF, the association becomes stronger, meaning that this association is mainly present in individuals with a lower atherosclerotic burden and/or a preserved ejection fraction. This suggests that these microRNAs might reflect another underlying pathophysiological mechanism strongly related to cardiovascular death. Taken together, Karakas *et al.* importantly advance this field by showing predictive power well beyond currently used risk scores, and suggest that the circulating microRNAs measured are related to cardiovascular death. Obviously, more detailed analysis of the technical details of their RT-qPCR measurement may improve signal to noise ratio and further facilitate the use of these biomarkers.

### Relation to other microRNAs studies

In this study, microRNAs that were previously associated with UAP were identified (5). Although only small studies have compared UAP with controls (9-11), none of these markers that were identified in the study of Zeller *et al.* (5), have been found previously. Additionally, since UAP is atherosclerosis without substantial ischemia, some overlap with previously found markers for SAP in earlier studies would have been expected. However, none of the eight microRNAs that were measured in the study of Karakas *et al.* (4) were found in a total of 17 earlier studies on SAP markers (12). Although, the study of Zeller *et al.* (5) is the first to consistently show an association of microRNAs with UAP in three independent cohorts, we maintain to wonder why so many different study of microRNA biomarkers

produce different results. This might be a cause of the many pitfalls of microRNA measurement, the lack of standard methods for measurements and normalization or handling of missing data. On the other hand the study of Karakas *et al.* (4) remains one of the largest and most thoroughly performed studies in the field of circulating microRNAs. Moreover, they found highly interesting markers for coronary artery disease risk prediction, despite the raised issues concerning data handling. However, subsequent external validation with more accurate measurements are needed to assess if these circulating microRNA biomarkers could be of value in clinical practice.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281-297.
2. Gupta SK, Bang C, Thum T. Circulating microRNAs as biomarkers and potential paracrine mediators of cardiovascular disease. *Circ Cardiovasc Genet* 2010;3:484-488.
3. Mitchell PS, Parkin RK, Kroh EM, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 2008;105:10513-10518.
4. Karakas M, Schulte C, Appelbaum S, et al. Circulating microRNAs strongly predict cardiovascular death in patients with coronary artery disease—results from the large AtheroGene study. *Eur Heart J* 2016. [Epub ahead of print].
5. Zeller T, Keller T, Ojeda F, et al. Assessment of microRNAs in patients with unstable angina pectoris. *Eur Heart J* 2014;35:2106-2114.
6. Bustin SA, Benes V, Garson JA, et al. The MIQE guidelines: minimum information for publication of quantitative real-time PCR experiments. *Clin Chem* 2009;55:611-622.
7. Zampetaki A, Mayr M. Analytical challenges and technical limitations in assessing circulating miRNAs. *Thromb Haemost* 2012;108:592-598.
8. Kok MG, Halliani A, Moerland PD, et al. Normalization panels for the reliable quantification of circulating microRNAs by RT-qPCR. *FASEB J* 2015;29:3853-3862.
9. Adachi T, Nakanishi M, Otsuka Y, et al. Plasma microRNA 499 as a biomarker of acute myocardial infarction. *Clin Chem* 2010;56:1183-1185.
10. D'Alessandra Y, Carena MC, Spazzafumo L, et al. Diagnostic potential of plasmatic MicroRNA signatures in stable and unstable angina. *PLoS One* 2013;8:e80345.
11. Guo M, Mao X, Ji Q, et al. miR-146a in PBMCs modulates Th1 function in patients with acute coronary syndrome. *Immunol Cell Biol* 2010;88:555-564.
12. Romaine SP, Tomaszewski M, Condorelli G, et al. MicroRNAs in cardiovascular disease: an introduction for clinicians. *Heart* 2015;101:921-928.

**Cite this article as:** de Ronde MW, Pinto YM, Pinto-Sietsma SJ. Circulating microRNA biomarkers for cardiovascular risk prediction: are we approaching clinical application? *Ann Transl Med* 2016;4(24):490. doi: 10.21037/atm.2016.12.05

# Exosomes: scytales in the damaged heart

Lara Ottaviani<sup>1</sup>, Leon J. De Windt<sup>1</sup>, Paula A. da Costa Martins<sup>1,2</sup>

<sup>1</sup>Department of Cardiology, CARIM School for Cardiovascular Diseases, Faculty of Health, Medicine and Life Sciences, Maastricht University, Maastricht, the Netherlands; <sup>2</sup>Department of Physiology and Cardiothoracic Surgery, Faculty of Medicine, University of Porto, Porto, Portugal

*Correspondence to:* Paula A. da Costa Martins, PhD. Department of Cardiology, CARIM School for Cardiovascular Diseases, Faculty of Health, Medicine and Life Sciences, Maastricht University, Maastricht, the Netherlands. Email: p.dacostamartins@maastrichtuniversity.nl

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Chistiakov DA, Orekhov AN, Bobryshev YV. Cardiac extracellular vesicles in normal and infarcted heart. *Int J Mol Sci* 2016;17:E63.

Submitted Apr 20, 2016. Accepted for publication Apr 29, 2016.

doi: 10.21037/atm.2016.05.17

View this article at: <http://dx.doi.org/10.21037/atm.2016.05.17>

## Exosomes and intracellular communication

As cells in our organism are constantly sending out and receiving signals, cell-cell communication is an essential way to maintain process homeostasis while allowing adaptation to external stimuli. Disturbances in cell-to-cell communication will result in disease. Cells communicate with each other via extracellular molecules such as nucleotides, short peptides, proteins or lipids that are released to the extracellular space and bind receptors on other cells, therefore inducing signaling and modifying the molecular status of the recipient cells. In addition to such molecules, cells also release membrane vesicles, representing a rich source of small molecules such as messenger RNAs, microRNAs (miRNAs) and long noncoding RNAs (lncRNAs), small amounts of DNA and low molecular weight lipids and proteins (including transcription factors and cytokines), all of which can also alter recipient cells that encounter such structures. Although initially thought as cellular debris and a sign of cellular death, in recent years more interest has been dedicated to extracellular vesicles (EVs) as mediators of long-range cellular communication by their presence in most body fluids. According to their size, EVs can be classified in microvesicles (MVs) (0.1 to 1  $\mu$ m), exosomes (20 to 100 nm) and apoptotic bodies (ABs) (0.5 to 2  $\mu$ m). (1) While MVs (and ABs) are assembled by budding from the plasma membranes, exosomes are raised from endosomal vesicles and formed as intraluminal vesicles by inward budding of the limiting membrane of the multivesicular bodies, which allows the internalization of small proteins, mRNAs, miRNAs and DNA. Once the

multivesicular bodies fuse with the cell membrane, exosomes are released (2) and free to interact with target cells in different ways: (I) exosomes can fuse in a non-selective way releasing their content into the target cell; (II) hemifusion followed by a complete fusion between the exosome and the cell membrane; (III) internalization by phagocytosis; and (IV) specific interactions among exosomes and target cells mediated by extracellular matrix components (3).

It is important to underline that exosomes, although generated by different cell types expressing distinct markers, also carry common surface markers such as heat shock protein HSP70 and tetraspanins CD9 and CD63, making their characterization a difficult process (4). For research purposes, it is fundamental to purify EV populations, but despite a large body of literature describing protocols on this matter, a gold standard method for the isolation of exosomes remains to be established. This is a critical point while studying exosome-mediated cell communication since different isolation methods still result in different yield and purity populations (5).

## Exosomes and intracardiac cell communication

Heart failure is one of the leading causes of morbidity and mortality worldwide (6). Cardiac remodeling is often a maladaptive mechanism that, independently of the nature of the initial insult, will ultimately cause ventricular dysfunction (7). Proper cardiac function does not only rely on cardiac muscle cells which account for one third of the total cell number of the myocardium, but rather on a

balance with other cell types including smooth muscle cells, endothelial cells, fibroblasts and immune cells. These very distinct cell types do not function isolated from each other but rather interact physically and/or via different autocrine, paracrine and endocrine factors. Indeed, the myocardium secretes exosomes, which are involved in intracellular communication in the adult heart (8). As extracellular spaces have mixed exosome populations from all kinds of cellular sources it is difficult to distinguish exosomes derived from a specific cell type from the intact organ. For this reason, *in vitro* studies have been very helpful not only providing evidence of exosome secretion but also in identifying the exosomal content and function from specific cell types. Early reports using primary cardiomyocytes and relevant cell lines have provided evidence of exosome secretion and detected nucleic acid-containing microvesicles/exosomes in the cell media, which could reprogram fibroblast gene expression (9,10). These investigations introduced a new concept in cardiac cell-cell communication, proposing that exosomes generated by cardiomyocytes are able to transfer protein or genetic information to neighboring cells of the heart.

Recent studies indicate that exosomal content is highly regulated by stress and disease conditions and despite the fact that research on cardiac exosomes is just emerging, a limited number of publications provide strong evidence that exosomes can exert pathological effects during cardiac response to stress as shown for different myocardial diseases as cardiac hypertrophy and peripartum or diabetic cardiomyopathy. Exosomes secreted from fibroblasts are enriched in miR-21\* which once uptaken by cardiomyocytes will down-regulate Sorbin and SH3 domain 2 (SORBS2) or PDZ and LIM domain 5 (PDLIM5), both regulators of cardiac muscle structure and function, and induce cardiac hypertrophy (11). The fact that fibroblast-derived miR-21\* induces cardiomyocyte hypertrophy also demonstrates that miRNA passenger strands can function as mature miRNAs (11). Another study demonstrates how, in women affected with peripartum cardiomyopathy (PPCM), a 16-kDa prolactin fragment (16K PRL) leads to the release of miR-146-enriched exosomes by endothelial cells (12). These exosomes can be uptaken by cardiomyocytes where miR-146a interferes with the physiological metabolism and contractile capacity of the cell, leading to the development of hypertrophy. Furthermore, levels of exosomal miR-146a were higher in plasma from patients with acute PPCM than healthy postpartum controls and patients with dilated cardiomyopathy. Maybe more important to underline is that

standard heart failure therapy in PPCM patients lowered circulating exosomal miR-146a to control levels, indicating miR-146a as a strong potential biomarker for diagnosis and risk stratification of patients with PPCM (12).

As miRNA composition may reflect the metabolic or differentiation state of the exosome-producing cells, circulating exosomes found in body fluids such as plasma and serum are becoming an attractive tool for analytical studies and subsequent disease diagnosis (13). In agreement, the recent work of Chistiakov *et al.* (14), very elegantly underlines that differential exosomal content can lead to new cardiac-specific diagnostic markers by emphasizing the central role of exosomes in cardiac regeneration of the infarcted heart.

### Exosomes and cardiac repair

The type of stress to which myocardial tissue is exposed seems to determine the content of the secreted EVs. In myocardial infarction (MI) where the heart is subjected to ischemic stress signals such as hypoxia, inflammation and injury, cardiomyocytes increase the secretion of MVs and/or exosomes. In addition, the distinct content between exosomes deriving from the border zone of the MI and the ones originating from the healthy myocardium suggests an adaptive response to injury. Indeed, circulating miRNAs are markedly altered after MI. miR-1 and mir-133 are elevated in the serum of patients with acute coronary syndrome, correlating with the levels of the clinical biomarker troponin T (15). Together with miR-499, these cardiac specific miRNAs are released from the infarcted and peri-infarcted myocardium and regulate the expression of sarcomeric genes and ion channels (15,16). miR-1 is also elevated, together with miR-208, in the urine of acute MI patients (17) indicating that circulating miRNAs released from the injured myocardium can travel to distant organs via exosomes as they are stable and protected from degradation by RNases present in the different body fluids. Many of these miRNAs are released immediately after an insult and could therefore, be used as markers for early detection of acute MI. In fact, circulating miR-126 is an important indicator of damage and repair mechanisms in acute MI patients exemplifying that monitoring of exosomal contents after MI can be a factor of prognosis evaluation and prediction (18,19).

Stem cells have been used in an attempt to regenerate damaged tissue when injected in the injured heart region, by engrafting, proliferating, differentiating and repopulating

the myocardium. The therapeutic effects of stem cells more likely result from the secretion of molecules such as growth factors, antioxidants, cytokines, chemokines and miRNAs with a wide-range of physiological effects (20). The review by Chistiakov *et al.* (14) emphasizes the contribution of cardiac progenitor cells (CPCs) as source of exosomes with regenerative properties. Under hypoxic conditions, CPCs are able to secrete pro-regenerative exosomes capable of inducing tube formation, proliferation and migration of endothelial cells (21,22). Elevated levels of miR-132 and miR-146 have been found in exosomes derived from hypoxic CPCs and infusion of these exosomes in a rat model of ischemia-reperfusion injury reduced fibrosis and enhanced heart function (22). These findings demonstrate how hypoxia is able to re-enforce the regenerative capacity of CPCs via exosomes. Also critical is the type of parent cell secreting exosomes. While delivery of CPCs-secreted EVs to infarcted rat hearts resulted in decreased cardiomyocyte apoptosis, reduced collagen deposition, increased blood vessel density, and ultimately in improved cardiac function, treatment with fibroblast-derived EVs did not have any effect (21). The differences observed may relate to elevated paracrine secretion of miR-210, miR-132 and miR-146a-3p from CPCs and not from fibroblasts (21). Cardiosphere-derived cardiac stem cells were reported to employ a similar mechanism via transfer of miR-146a to mediate cardioprotection by inhibiting apoptosis, promoting cardiomyocyte proliferation and inducing angiogenesis (23). Whether this cardioprotective effect is progenitor/cardiac stem cell preparation-dependent or whether it is mediated by a specific combination of miRNAs and other factors is not clear. Nevertheless, these results suggest progenitor and cardiac stem cells as unlimited sources of cardioprotective exosomes with promising therapeutic potential for post-MI cardiac repair.

### Future perspectives/limitations

Only recently, the potential of using exosomes as therapy for MI and other cardiac pathologies has started to be evaluated in preclinical animal models. Among all types of EVs, exosomes are the richest in (micro)RNAs and could constitute a tool for cell- or tissue-specific delivery of RNA molecules of interest after modulating exosomal membrane receptors. One of the main problems in delivering short interfering RNAs (siRNA) or miRNAs *in vivo* is targeting specific tissues and avoiding non-specific delivery. Exosomes, as natural RNA carriers, would also

bypass the issue of immunogenicity of siRNA or miRNA or their cargo vehicle. In fact, different strategies have been used to manipulate exosomal content (24) for specific delivery in the brain and targeting of neurons, microglia and oligodendrocytes (25). Exosomes are versatile carriers of both protective and pathological molecular signals and while they appear to be safe and efficient much effort should be directed to identifying their bioactive molecular content as well as understanding how such content will affect specific signaling pathways and cellular mechanisms in recipient cells.

Besides their promising clinical relevance in the identification of novel biomarkers of cardiac injury in ischemic or other cardiac diseases, the clinical and scientific benefits of studying exosomes as extracellular communicators in cardiac disease are multifold. Such studies will reveal novel mechanisms of how cells and organs communicate among each other, aid in understanding and developing new cell therapies for ischemia as well as providing insights for the development of novel therapeutics, and finally to reveal mechanisms of cell targeting for the discovery of novel candidates and delivery of therapeutic compounds for cardiac diseases. But again, issues such as biosafety and tolerance of exosome-based therapies, which could render their translational power, deserve special attention in the future.

### Acknowledgements

PA da Costa Martins is supported by a MEERVOUD grant from The Netherlands Organisation for Scientific Research (NWO) and is an Established Investigator of the Dutch Heart Foundation. LJ De Windt acknowledges support from the *Netherlands CardioVascular Research Initiative*: the Dutch Heart Foundation, Dutch Federation of University Medical Centers, ZonMW and the Royal Netherlands Academy of Sciences. LJ de Windt was further supported by grant 311549 from the European Research Council (ERC) and a VICI award 918-156-47 from NWO.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Pfeifer P, Werner N, Jansen F. Role and function of



- microRNAs in extracellular vesicles in cardiovascular biology. *Biomed Res Int* 2015;2015:161393.
2. Katzmann DJ, Babst M, Emr SD. Ubiquitin-dependent sorting into the multivesicular body pathway requires the function of a conserved endosomal protein sorting complex, ESCRT-I. *Cell* 2001;106:145-155.
  3. Corrado C, Raimondo S, Chiesi A, et al. Exosomes as intercellular signaling organelles involved in health and disease: basic science and clinical applications. *Int J Mol Sci* 2013;14:5338-5366.
  4. Théry C, Ostrowski M, Segura E. Membrane vesicles as conveyors of immune responses. *Nat Rev Immunol* 2009;9:581-593.
  5. Xu R, Greening DW, Zhu HJ, et al. Extracellular vesicle isolation and characterization: toward clinical application. *J Clin Invest* 2016;126:1152-1162.
  6. Mozaffarian D, Benjamin EJ, Go AS, et al. Heart disease and stroke statistics-2016 update: a report from the American heart association. *Circulation* 2016;133:e38-360.
  7. Philippen LE, Dirx E, da Costa-Martins PA, et al. Non-coding RNA in control of gene regulatory programs in cardiac development and disease. *J Mol Cell Cardiol* 2015;89:51-58.
  8. Barile L, Gherghiceanu M, Popescu LM, et al. Ultrastructural evidence of exosome secretion by progenitor cells in adult mouse myocardium and adult human cardiospheres. *J Biomed Biotechnol* 2012;2012:354605.
  9. Waldenstrom A, Genneback N, Hellman U, et al. Cardiomyocyte microvesicles contain DNA/RNA and convey biological messages to target cells. *PLoS One* 2012;7:e34653.
  10. Malik ZA, Kott KS, Poe AJ, et al. Cardiac myocyte exosomes: stability, HSP60, and proteomics. *Am J Physiol Heart Circ Physiol* 2013;304:H954-H965.
  11. Bang C, Batkai S, Dangwal S, et al. Cardiac fibroblast-derived microRNA passenger strand-enriched exosomes mediate cardiomyocyte hypertrophy. *J Clin Invest* 2014;124:2136-2146.
  12. Halkein J, Tabruyn SP, Ricke-Hoch M, et al. MicroRNA-146a is a therapeutic target and biomarker for peripartum cardiomyopathy. *J Clin Invest* 2013;123:2143-2154.
  13. Vickers KC, Palmisano BT, Shoucri BM, et al. MicroRNAs are transported in plasma and delivered to recipient cells by high-density lipoproteins. *Nat Cell Biol* 2011;13:423-433.
  14. Chistiakov DA, Orekhov AN, Bobryshev YV. Cardiac extracellular vesicles in normal and infarcted heart. *Int J Mol Sci* 2016;17:E63.
  15. Kuwabara Y, Ono K, Horie T, et al. Increased microRNA-1 and microRNA-133a levels in serum of patients with cardiovascular disease indicate myocardial damage. *Circ Cardiovasc Genet* 2011;4:446-454.
  16. He B, Xiao J, Ren AJ, et al. Role of miR-1 and miR-133a in myocardial ischemic postconditioning. *J Biomed Sci* 2011;18:22.
  17. Cheng Y, Wang X, Yang J, et al. A translational study of urine miRNAs in acute myocardial infarction. *J Mol Cell Cardiol* 2012;53:668-676.
  18. Rautou PE, Vion AC, Amabile N, et al. Microparticles, vascular function, and atherothrombosis. *Circ Res* 2011;109:593-606.
  19. Zampetaki A, Willeit P, Tilling L, et al. Prospective study on circulating MicroRNAs and risk of myocardial infarction. *J Am Coll Cardiol* 2012;60:290-299.
  20. van Berlo JH, Molkenin JD. An emerging consensus on cardiac regeneration. *Nat Med* 2014;20:1386-1393.
  21. Barile L, Lionetti V, Cervio E, et al. Extracellular vesicles from human cardiac progenitor cells inhibit cardiomyocyte apoptosis and improve cardiac function after myocardial infarction. *Cardiovasc Res* 2014;103:530-541.
  22. Gray WD, French KM, Ghosh-Choudhary S, et al. Identification of therapeutic covariant microRNA clusters in hypoxia-treated cardiac progenitor cell exosomes using systems biology. *Circ Res* 2015;116:255-263.
  23. Cheng HS, Sivachandran N, Lau A, et al. MicroRNA-146 represses endothelial activation by inhibiting pro-inflammatory pathways. *EMBO Mol Med* 2013;5:949-966.
  24. El-Andaloussi S, Lee Y, Lakhali-Littleton S, et al. Exosome-mediated delivery of siRNA in vitro and in vivo. *Nat Protoc* 2012;7:2112-2126.
  25. Alvarez-Erviti L, Seow Y, Yin H, et al. Delivery of siRNA to the mouse brain by systemic injection of targeted exosomes. *Nat Biotechnol* 2011;29:341-345.

**Cite this article as:** Ottaviani L, De Windt LJ, da Costa Martins PA. Exosomes: scytales in the damaged heart. *Ann Transl Med* 2016;4(11):222. doi: 10.21037/atm.2016.05.17

# Is there a role for microRNAs as novel predictors of prognosis in myocardial infarction?

Robert Adam, Dominic Kelly

Department of Cardiology, Basingstoke and North Hampshire Hospital, Aldermaston Road, Basingstoke, Hampshire, RG24 9NA, UK

*Correspondence to:* Dr. Robert Adam. Department of Cardiology, Basingstoke and North Hampshire Hospital, Aldermaston Road, Basingstoke, Hampshire, RG24 9NA, UK. Email: Robert.Adam@hhft.nhs.uk.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Coskunpinar E, Cakmak HA, Kalkan AK, *et al.* Circulating miR-221-3p as a novel marker for early prediction of acute myocardial infarction. *Gene* 2016;591:90-6.

Submitted Oct 07, 2016. Accepted for publication Oct 12, 2016.

doi: 10.21037/atm.2016.11.50

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.11.50>

MicroRNAs (miRNAs) are endogenous, non-coding, single stranded RNAs of 19–25 nucleotides in length that regulate gene expression at the post-transcription processing steps (1). Although the exact biological functions of miRNAs are still not fully understood they have been implicated in cellular development, differentiation, metabolism and death (2,3).

The discovery that stable miRNAs could be identified in human plasma/serum (4) enabled researchers to investigate their expression levels in various cardiovascular conditions (5). Circulating levels of specific miRNAs have been found to be significantly increased in AMI patients compared to controls, leading to speculation that they may have a role to play as novel diagnostic biomarkers (6).

In this edition of *Gene*, Coskunpinar *et al.* present data from a small (27 patients), single centre study, demonstrating the use of circulating miRNAs as novel markers for early prediction of acute myocardial infarction and LV dysfunction post AMI (7). Real time PCR was used to determine the expression levels of 1,116 miRNAs in 27 patients post AMI and 16 control subjects. Six miRNAs were identified as being significantly upregulated in the AMI group compared with the control group.

The most promising miRNA, miR-221-3p was shown to correlate modestly with GRACE and SYNTAX scores as well as serum troponin. In practical terms, the paper suggests that this may represent a novel marker for the early prediction of AMI. However, when miR-221-3p was assessed via receiver operator curves for the prediction of

AMI, the AUC was calculated at 0.881, which was inferior to that of the current standard clinical test of troponin (AUC 0.954). Moreover, the addition of miR-221-3p to troponin did not appear to add additional diagnostic capacity.

Ongen *et al.* also observed an inverse relationship with LV ejection fraction. Such relationships would be expected of any biomarker which is collinear with troponin and is therefore not unusual. Moreover, the relationship was demonstrated in uni-variable analysis only and has not been tested in more robust multi-variable analysis.

The observation of relationship between miR-221-3p and LV function is interesting but certainly not a unique finding for miRNAs. Devaux *et al.* (8,9) have previously demonstrated miRNAs to be associated with LV function, remodelling and cardiac contractility post MI in a larger cohort of patients and other studies have shown similar (10).

The utility of miR-221-3p as a prognostic marker has not been tested in this study as no clinical endpoint data were presented. However, one may expect that a biomarker which is associated with Troponin, GRACE and LV function would also be associated with adverse outcomes. Indeed, long term follow up data from the AtheroGene Study identified three miRNAs which precisely predict cardiovascular mortality following AMI establishing their potential as predictors of prognosis (11).

As acknowledged by the authors, the small sample size is the major limitation of this study. Further studies are required both to validate the findings of this small single



centre investigation and also to expand to allow use as a marker of prognosis and adverse LV remodelling. In addition, the authors link the increased expression of miRNAs identified in the study, with the down regulation of four target genes that have previously been found to regulate various cardioprotective molecular pathways. The authors infer that this relationship may hence increase susceptibility to AMI. Further research is required in order to establish any potential causative mechanisms.

In conclusion, this study adds to the expanding list of biomarkers in the prediction of AMI. With over 2,000 miRNAs already catalogued (12) this remains a vast topic with many exciting research avenues to pursue in the future. The ability to identify the genetic functions of specific miRNAs and their influence on particular cellular processes significantly increases our understanding of complex pathophysiological mechanisms involved in the development of conditions such as AMI.

Combining this knowledge with the rapidly evolving science of genetic engineering is already allowing infections such as Hepatitis C to be tackled in revolutionary ways (13) and may radically change the way we manage numerous cardiovascular diseases in the future.

## Acknowledgements

None.

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

1. Bartel DP. MicroRNAs: genomics, biogenesis, mechanism, and function. *Cell* 2004;116:281-297.
2. Ambros V. The functions of animal microRNAs. *Nature* 2004;431:350-355.
3. Plasterk RH. Micro RNAs in animal development. *Cell* 2006;124:877-881.
4. Mitchell PS, Parkin RK, Kroh EM, et al. Circulating microRNAs as stable blood-based markers for cancer detection. *Proc Natl Acad Sci U S A* 2008;105:10513-10518.
5. Small EM, Olson EN. Pervasive roles of microRNAs in cardiovascular biology. *Nature* 2011;469:336-342.
6. Navickas R, Gal D, Laucevičius A, et al. Identifying circulating microRNAs as biomarkers of cardiovascular disease: a systematic review. *Cardiovasc Res* 2016;111:322-337.
7. Coskunpinar E, Cakmak HA, Kalkan AK, et al. Circulating miR-221-3p as a novel marker for early prediction of acute myocardial infarction. *Gene* 2016;591:90-96.
8. Devaux Y, Vausort M, McCann GP, et al. MicroRNA-150: a novel marker of left ventricular remodeling after acute myocardial infarction. *Circ Cardiovasc Genet* 2013;6:290-298.
9. Devaux Y, Vausort M, McCann GP, et al. A panel of 4 microRNAs facilitates the prediction of left ventricular contractility after acute myocardial infarction. *PLoS One* 2013;8:e70644.
10. Gidlöf O, Smith JG, Miyazu K, et al. Circulating cardio-enriched microRNAs are associated with long-term prognosis following myocardial infarction. *BMC Cardiovasc Disord* 2013;13:12.
11. Karakas M, Schulte C, Appelbaum S, et al. Circulating microRNAs strongly predict cardiovascular death in patients with coronary artery disease—results from the large AtheroGene study. *Eur Heart J* 2016. [Epub ahead of print].
12. Romaine SP, Tomaszewski M, Condorelli G, et al. MicroRNAs in cardiovascular disease: an introduction for clinicians. *Heart* 2015;101:921-928.
13. Miravirsin Study in Null Responder to Pegylated Interferon Alpha Plus Ribavirin Subjects With Chronic Hepatitis C. Accessed 29 April, 2016. Available online: <https://clinicaltrials.gov/show/NCT01727934>

**Cite this article as:** Adam R, Kelly D. Is there a role for microRNAs as novel predictors of prognosis in myocardial infarction? *Ann Transl Med* 2016;4(23):473. doi: 10.21037/atm.2016.11.50

# MicroRNAs to take the place of collateral flow index measurements and Rentrop scoring? – Reply to Papageorgiou *et al.*

Nazanin Hakimzadeh<sup>1,2</sup>, Jan J. Piek<sup>2</sup>

<sup>1</sup>Department of Biomedical Engineering & Physics, <sup>2</sup>Department of Cardiology, Academic Medical Center, University of Amsterdam, Amsterdam, The Netherlands

*Correspondence to:* Jan J. Piek, Department of Cardiology, Academic Medical Center, University of Amsterdam, Room B2-250, Meibergdreef 9, 1105 AZ Amsterdam, The Netherlands. Email: j.j.piek@amc.uva.nl.

*Provenance:* This is a Guest Letter to the Editor commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Response to:* Papageorgiou N, Zacharia E, Tousoulis D. Association between microRNAs and coronary collateral circulation: is there a new role for the small non-coding RNAs? *Ann Transl Med* 2016;4:223.

Submitted Jul 02, 2016. Accepted for publication Jul 04, 2016.

doi: 10.21037/atm.2016.07.26

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.07.26>

We thank Papageorgiou *et al.* for their thoughtful comments on our recent publication regarding circulating microRNAs (miRNAs) associated with coronary collateral artery capacity (1). The benefits of a vast collateral network have been widely accepted to prevent mortality in patients with chronic stable coronary artery disease (CAD) (2,3). Identifying patients with a limited collateral network can in turn distinguish patients at risk to substantial debilitation from adverse cardiac events. Nonetheless, current methods to identify patients with insufficient collateralization are limited to invasive intracoronary collateral flow index (CFI) measurements or angiographic grading (4). Clinical parameters associated with collateral development have been identified (5). However, there remained a lack of biomarkers to discriminate between patients with sufficient or insufficient collateralization.

Genetic heterogeneity in CAD patients has been identified at the messenger RNA level of circulating monocytes and macrophage phenotypes (6,7). This led us to hypothesize that differential miRNA expression in patients with insufficient *vs.* sufficient collateralization must also be present. We identified 4 miRNAs that were significantly upregulated in the plasma of patients with low collateral capacity. We further determined that these miRNAs (miR423-5p, miR10b, miR30d, miR126) could serve as circulating biomarkers to significantly distinguish between chronic total occlusion (CTO) patients with high or low collateral capacity. These miRNAs can discriminate

between these patient groups with a positive likelihood ratio between 3.0 and 6.1 depending on the respective miRNA. This revelation can be of immense clinical significance, whereby patients with low collateralization can be potentially identified with a simple blood sample rather than invasive intracoronary catheterization. However, our study warrants further studies with larger patient cohorts, along with examining the utility of these miRNAs as biomarkers for collateralization in CAD patients.

Papageorgiou *et al.* point out that the utility of these miRNAs is also dependent on a generally accepted definition of low collateral capacity, whereby some studies use Rentrop scoring and others use different thresholds of pressure-derived CFI (CFI<sub>p</sub>) to establish the level of collateralization. Rentrop scoring provides only a semi-quantitative measure and is limited to collateral vessels above 100 µm diameter. In relation to CFI<sub>p</sub> measurements, it is important to recognize the difference between CAD and CTO patients, whereby CTO patients provide no variability in coronary lesion severity as compared to CAD patients. As a result, the distribution of CFI<sub>p</sub> measurements in CTO patient populations differs from that in CAD patients. The mean CFI<sub>p</sub> in a cohort of 295 patients was deemed to be 0.39 (5), while in CAD patients the frequency distribution of CFI<sub>p</sub> is dependent on the severity of CAD (2).

As both Papageorgiou *et al.* and we have mentioned, a number of co-existing parameters may affect the diagnostic ability of miRNAs in the general population. Medication

usage, gender, age as well as diabetes mellitus have been shown to affect both collateral vessel development as well as miRNA expression levels (8-11). In our study, gender and age significantly impacted the predictive power of the respective miRNAs as suitable biomarkers. However, miR126 demonstrated significant predictive power to discriminate between patients with high and low collateral capacity even without consideration of age and gender. miRNA126 has been largely linked to angiogenesis, as well as atherosclerosis, whereby the mature miRNA-126-5p plays a role in endothelial turnover (12). Examining larger patient cohorts will likely elucidate the exact threshold of the respective miRNA expression levels that can distinguish between these patient groups, and provide additional insight in the effects of other co-existing parameters on this threshold.

In conclusion, circulating miRNAs offer a new method for patient stratification. Nonetheless, there still remains a large step before miRNA examination can become a part of routine clinical application. As the possibility to distinguish the coronary collateral artery capacity of patients without the need for invasive catheterization is of great clinical significance, our results warrant further investigation in larger CTO patient cohorts, along with examining the predictive power of these miRNAs in CAD patients.

### Acknowledgements

*Funding:* This work was performed within the framework of CTMM, Center for Translational Molecular Medicine ([www.ctmm.nl](http://www.ctmm.nl)), project EMINENCE (grant 01C-204).

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

- Hakimzadeh N, Nossent AY, van der Laan AM, et al. Circulating MicroRNAs Characterizing Patients with Insufficient Coronary Collateral Artery Function. *PLoS One* 2015;10:e0137035.
- Meier P, Gloekler S, Zbinden R, et al. Beneficial effect of recruitable collaterals: a 10-year follow-up study in patients with stable coronary artery disease undergoing quantitative collateral measurements. *Circulation* 2007;116:975-983.
- Meier P, Seiler C. The coronary collateral circulation—clinical relevances and therapeutic options. *Heart* 2013;99:897-898.
- Hakimzadeh N, Verberne HJ, Siebes M, et al. The future of collateral artery research. *Curr Cardiol Rev* 2014;10:73-86.
- van der Hoeven NW, Teunissen PF, Werner GS, et al. Clinical parameters associated with collateral development in patients with chronic total coronary occlusion. *Heart* 2013;99:1100-1105.
- Schirmer SH, Fledderus JO, Bot PT, et al. Interferon-beta signaling is enhanced in patients with insufficient coronary collateral artery development and inhibits arteriogenesis in mice. *Circ Res* 2008;102:1286-1294.
- van der Laan AM, Schirmer SH, de Vries MR, et al. Galectin-2 expression is dependent on the rs7291467 polymorphism and acts as an inhibitor of arteriogenesis. *Eur Heart J* 2012;33:1076-1084.
- Fichtlscherer S, De Rosa S, Fox H, et al. Circulating microRNAs in patients with coronary artery disease. *Circ Res* 2010;107:677-684.
- Tijssen AJ, Creemers EE, Moerland PD, et al. MiR423-5p as a circulating biomarker for heart failure. *Circ Res* 2010;106:1035-1039.
- Dimmeler S, Nicotera P. MicroRNAs in age-related diseases. *EMBO Mol Med* 2013;5:180-190.
- Epstein SE, Lassance-Soares RM, Faber JE, et al. Effects of aging on the collateral circulation, and therapeutic implications. *Circulation* 2012;125:3211-3219.
- Schober A, Nazari-Jahantigh M, Wei Y, et al. MicroRNA-126-5p promotes endothelial proliferation and limits atherosclerosis by suppressing Dlk1. *Nat Med* 2014;20:368-376.

**Cite this article as:** Hakimzadeh N, Piek JJ. MicroRNAs to take the place of collateral flow index measurements and Rentrop scoring?—Reply to Papageorgiou *et al.* *Ann Transl Med* 2016;4(15):297. doi: 10.21037/atm.2016.07.26

# Circulating extracellular vesicles containing miRNAs may have utility as early biomarkers for cardiac injury

Bethany Doran<sup>1</sup>, Deepak Voora<sup>2</sup>

<sup>1</sup>Duke Molecular Physiology Institute, Durham, NC, USA; <sup>2</sup>Duke Center for Applied Genomics & Precision Medicine, Durham, NC, USA

*Correspondence to:* Deepak Voora, MD. 101 Science Dr, 2187 CIEMAS, Campus Box 3382, Durham, NC 27708, USA. Email: Deepak.voora@duke.edu.

*Provenance:* This is a Guest Commentary commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Deddens JC, Vrijssen KR, Colijn JM, *et al.* Circulating Extracellular Vesicles Contain miRNAs and are Released as Early Biomarkers for Cardiac Injury. *J Cardiovasc Transl Res* 2016;9:291-301.

Submitted Sep 28, 2016. Accepted for publication Oct 03, 2016.

doi: 10.21037/atm.2016.10.55

View this article at: <http://dx.doi.org/10.21037/atm.2016.10.55>

Through advancements in prevention, awareness, detection, and treatment modalities, rates of myocardial infarction (MI) have decreased over time. However, MI still remains a significant cause of morbidity and financial burden within the United States, with approximately 3.2 per 1,000 individuals hospitalized per year, corresponding to an estimated 11.3 billion dollars in societal costs (1). The universal definition of MI specifies that patients must have a rise and fall of cardiac biomarkers as well as ischemic symptoms for diagnosis. Currently, circulating biomarkers in plasma including creatinine kinase MB (CK-MB) and cardiac troponin remain the gold standard for detection and validation of type 1 MI. Although assays using high sensitivity troponin are able to detect MI as early as 2–3 hours after cardiac injury with good sensitivity and specificity (2), in certain patients the initial assay may be negative. Because early recognition of MI is associated with improved outcomes, decreased hospital length of stay, and decreased cost (3–6), there is interest into novel diagnostic methods for MI. Beyond diagnosis of MI, improving the detection and etiology of myocardial injury using novel biomarkers may enhance the management of a variety of cardiac conditions.

It is in this context that Deddens and coauthors (7) present a study investigating the possible utility of quantification of microRNAs (miRNAs) and extracellular vesicle (EV) release to assist in early determination of myocardial injury. Using mouse and porcine models, their study demonstrates that circulating EVs as well as miRNAs are significantly increased in animals with induced MI as

compared with sham controls early after ischemia. The authors demonstrate that after ligation of the LAD in mice to induce myocardial ischemia, followed by reperfusion (n=3), the amount of EVs released are significantly greater than the sham arm (n=1) in mice at 150 minutes. Moving to porcine models, the authors serially sample plasma to determine the level of circulating miRNA at different time points. They demonstrate that the circulating miRNAs previously demonstrated to be increased in plasma after MI in humans are also increased in animal models with a significant increase demonstrated in cases as compared to controls 2.5–3.5 hours after ischemia. The authors found that significantly elevated miRNAs include miRNA-1, -208, and -499 but not miRNA-21 or miRNA-146a, and that these levels are higher in EV than in plasma.

Although limited by small study size, the study offers interesting opportunities for translational medicine. Because miRNAs are involved in gene expression at a post translational level, the ability to understand the role of miRNA in pathological processes may also provide possible therapeutic targets (8). Although the purpose of the analysis by Deddens and colleagues was not to determine the exact mechanism and significance of miRNA elevation after MI, identification of key miRNAs after ischemia is an important step in better understanding the physiologic process that occurs when myocardium becomes ischemic.

The importance of miRNAs in the post transcriptional regulation of gene expression is increasingly recognized (9). Prior studies have identified certain miRNAs as markers of cardiac ischemic/reperfusion injury with both regulatory,

protective, and diagnostic utility (10-12). In addition to modifying myocardial gene expression in response to injury, miRNA are secreted in a regulated manner into the circulation by EVs as part of intercellular communication (similar to hormones). The presence of EVs in the circulation provides an important “window” into the injured myocardium that is otherwise inaccessible in the clinical setting. EVs package miRNA in specific proteins (e.g., Ago2 or HDL) which render miRNA highly resistant to degradation. Unlike most extracellular RNA which is rapidly degraded in the absence of RNase inhibitors or strict handling conditions, EV miRNA is robust to degradation under most conditions. This feature coupled with the ability to readily measure RNA in clinical laboratories make EV miRNA an attractive platform for a clinical biomarker.

In addition to detection of myocardial necrosis, miRNA quantification and assessment may be of utility in elucidating the mechanism of myocardial injury, assisting in the prognostic and diagnostic capabilities in acute MI. This highlights possible future avenues of translation into clinical practice, with possible benefit in distinguishing between MI, heart failure, myocarditis, and other processes involving myocardial injury upon patient presentation. Additional potential applications include myocardial monitoring for cardiac allograft rejection, chemotherapy induced cardiomyopathy, asymptomatic severe valvular disease, and risk stratification during exercise stress testing.

Excitingly, miRNAs are also emerging as potential therapeutic targets, thus better understanding of their utility, function, and targets are a priority for investigation (13). EV miRNA are also being developed as therapeutic targets as the extent of their increase has been shown to be associated with worsened myocardial injury (14). Studies have demonstrated improvement in hepatitis C viremia in primate studies (15) and colon cancer (16), however the full potential of miRNA as therapeutic agents is only beginning to be fully realized.

### Acknowledgements

None.

### Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

### References

1. Sugiyama T, Hasegawa K, Kobayashi Y, et al. Differential time trends of outcomes and costs of care for acute myocardial infarction hospitalizations by ST elevation and type of intervention in the United States, 2001-2011. *J Am Heart Assoc* 2015;4:e001445.
2. Keller T, Zeller T, Peetz D, et al. Sensitive troponin I assay in early diagnosis of acute myocardial infarction. *N Engl J Med* 2009;361:868-877.
3. Forberg JL, Henriksen LS, Edenbrandt L, et al. Direct hospital costs of chest pain patients attending the emergency department: a retrospective study. *BMC Emerg Med* 2006;6:6.
4. Task Force for Diagnosis and Treatment of Non-ST-Segment Elevation Acute Coronary Syndromes of European Society of Cardiology. Bassand JP, Hamm CW, Guidelines for the diagnosis and treatment of non-ST-segment elevation acute coronary syndromes. *Eur Heart J* 2007;28:1598-1660.
5. Antman EM, Hand M, Armstrong PW, et al. 2007 Focused Update of the ACC/AHA 2004 Guidelines for the Management of Patients With ST-Elevation Myocardial Infarction: a report of the American College of Cardiology/American Heart Association Task Force on Practice Guidelines: developed in collaboration With the Canadian Cardiovascular Society endorsed by the American Academy of Family Physicians: 2007 Writing Group to Review New Evidence and Update the ACC/AHA 2004 Guidelines for the Management of Patients With ST-Elevation Myocardial Infarction, Writing on Behalf of the 2004 Writing Committee. *Circulation* 2008;117:296-329.
6. Thygesen K, Alpert JS, White HD, et al. Universal definition of myocardial infarction. *Circulation* 2007;116:2634-2653.
7. Deddens JC, Vrijisen KR, Colijn JM, et al. Circulating Extracellular Vesicles Contain miRNAs and are Released as Early Biomarkers for Cardiac Injury. *J Cardiovasc Transl Res* 2016;9:291-301.
8. Li Z, Rana TM. Therapeutic targeting of microRNAs: current status and future challenges. *Nat Rev Drug Discov* 2014;13:622-638.
9. Krol J, Loedige I, Filipowicz W. The widespread regulation of microRNA biogenesis, function and decay. *Nat Rev Genet* 2010;11:597-610.
10. Ren XP, Wu J, Wang X, et al. MicroRNA-320 is involved in the regulation of cardiac ischemia/reperfusion

- injury by targeting heat-shock protein 20. *Circulation* 2009;119:2357-2366.
11. Ji X, Takahashi R, Hiura Y, et al. Plasma miR-208 as a biomarker of myocardial injury. *Clin Chem* 2009;55:1944-1949.
  12. Wang X, Zhang X, Ren XP, et al. MicroRNA-494 targeting both proapoptotic and antiapoptotic proteins protects against ischemia/reperfusion-induced cardiac injury. *Circulation* 2010;122:1308-1318.
  13. van Rooij E, Purcell AL, Levin AA. Developing microRNA therapeutics. *Circ Res* 2012;110:496-507.
  14. Boulanger CM, Scoazec A, Ebrahimian T, et al. Circulating microparticles from patients with myocardial infarction cause endothelial dysfunction. *Circulation* 2001;104:2649-2652.
  15. Lanford RE, Hildebrandt-Eriksen ES, Petri A, et al. Therapeutic silencing of microRNA-122 in primates with chronic hepatitis C virus infection. *Science* 2010;327:198-201.
  16. Huang JL, Urtatiz O, Van Raamsdonk CD, Oncogenic G. Protein GNAQ Induces Uveal Melanoma and Intravasation in Mice. *Cancer Res* 2015;75:3384-3397.

**Cite this article as:** Doran B, Voora D. Circulating extracellular vesicles containing miRNAs may have utility as early biomarkers for cardiac injury. *Ann Transl Med* 2016;4(Suppl 1):S60. doi: 10.21037/atm.2016.10.55



# Long non-coding RNAs in heart failure: a promising future with much to learn

Samir Ounzain, Thierry Pedrazzini

Experimental Cardiology Unit, Department of Medicine, University of Lausanne Medical School, Lausanne, Switzerland

*Correspondence to:* Dr. Samir Ounzain. Experimental Cardiology Unit, Department of Medicine, University of Lausanne Medical School, CH-1011 Lausanne, Switzerland. Email: samir.ounzain@chuv.ch.

*Provenance:* This is a Guest Letter to the Editor commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Response to:* El Azzouzi H, Doevendans PA, Sluijter JP. Long non-coding RNAs in heart failure: an obvious Inc. *Ann Transl Med* 2016;4:182.

Submitted Jul 10, 2016. Accepted for publication Jul 14, 2016.

doi: 10.21037/atm.2016.07.13

View this article at: <http://dx.doi.org/10.21037/atm.2016.07.13>

El Azzouzi and colleagues (1) recently published a perspective describing our work, in which we characterised the murine long noncoding RNA (lncRNA) transcriptome post myocardial infarction (MI) (2). This perspective nicely summarised key aspects of our work. However, we would like to respond with some additional comments to clarify and expand important points resulting from their perspective and our work.

Considering the emerging roles for lncRNAs in dictating the gene regulatory networks underpinning developmental and disease processes (3,4), we set out to characterise this landscape within the infarcted mouse heart (2). We utilised an extremely deep RNA sequencing approach coupled with *de novo* transcript reconstruction to comprehensively profile and discover novel lncRNAs. As discussed, we identified approximately 1,500 previously unannotated lncRNAs, with a significant fraction being differentially expressed post stress. Importantly, through integrating publicly available transcriptomic and epigenomic datasets, we demonstrated that our newly discovered lncRNAs exhibited some unique functional and regulatory characteristics. Firstly, they were significantly more enriched in the heart as compared to protein coding genes (PCGs) or previously annotated lncRNAs. Much of this increased heart specificity is likely as a consequence of their functional genomic loci of origin that are active cardiac enhancer regions. Enhancers are important distally acting cis regulatory sequences (5-7). They are considered the key information processing units within the genome integrating temporal and spatial

cues to direct correct context-specific gene expression patterns. Thus, we found that the novel lncRNAs that were modulated post-MI were significantly more associated with active cardiac enhancer sequences, therefore implicating the enhancer landscape and their associated lncRNAs, in the transcriptional reprogramming that underpins pathological remodelling. Finally, and importantly from a translational and therapeutic perspective, we identified hundreds of predicted human orthologs and validated their expression in human samples of a pathological nature (8).

Despite the insightful perspective by El Azzouzi *et al.* (1), we would like to emphasise and clarify a few points. Firstly, what was somewhat unique and surprising during our initial analysis was the sheer number of novel lncRNAs we were able to discover. This, as we have discussed in depth elsewhere (9) and would like to emphasise here, is as a direct consequence of the RNA sequencing depth. We would strongly argue based on our previous experience, that sequencing to a depth of at least 300 million paired-end reads is essential for the identification of novel tissue- and context-specific lncRNAs. With this sequencing depth, we were able to identify the 1,500 novel lncRNAs described in this study but also 2,500 novel lncRNAs in mouse embryonic stem cells (ESC) differentiating towards cardiomyocytes (10). To support our argument, we performed a computational simulation, in which we counted how many novel lncRNAs would have been discovered at different sequencing depths. We found that if we would have sequenced to the widely used depth of



50 million paired-end reads, we would have not detected approximately 50% to 60% of novel lncRNAs in both our studies (9). Strikingly, one should consider that the novel lncRNAs that required the greatest depth of sequencing for discovery corresponded to those exhibiting the most interesting regulatory and functional characteristics, in particular heart-enriched lncRNAs associated with cardiac developmental and functional roles. Those lncRNAs that were discovered at the shallower sequence depth were typically already annotated housekeeping like and non-tissue enriched lncRNAs.

El Azzouzi *et al.* (1) highlighted the cardiac-enriched nature of our novel lncRNAs. Despite validating a number of them via qPCR, it is important to note that this conclusion was primarily based on the integration of publicly available tissue-specific RNA-seq datasets. By using ENCODE data, we quantified the expression of every transcript in the heart and 17 non-cardiac tissues (2,9). Heart specificity was then determined for each transcript by calculating the expression in the heart versus the mean expression in all 17 non-cardiac tissues, generating a specificity score for each transcript (9). This approach is powerful and has been successfully used for lncRNAs discovered in a number of cell types and tissues demonstrating that newly discovered lncRNAs are typically more tissue-enriched *vs.* PCGs and also miRNAs (11). This is of important therapeutic potential, as targeting cell- and tissue-restricted lncRNAs could increase the specificity of future therapies and reduce off-target side effects. These observations also support the notion that lncRNAs could represent highly specific biomarkers for pathological processes (11,12). Indeed, a number of recent studies are beginning to validate this hypothesis. Along the same line, we demonstrated that cardiac lncRNAs expression is extremely well correlated with electrocardiographic traits. It is important to emphasise here that this correlation was executed in mouse and not in human patients as mentioned by El Azzouzi *et al.* (1). Comparable observations have however been made in human patients (11,12), supporting our work in mice. Furthermore, a number of recent studies have demonstrated the presence of lncRNAs circulating in the plasma (12). The detection of cardiac-enriched lncRNAs in the plasma therefore warrants interrogation and promises to be an exciting future application of our work with respect to post infarction remodelling and heart failure.

Despite the relative ease in discovering lncRNAs implicated in cardiovascular development and disease, elucidating their functions remains a major challenge.

We applied a novel approach to infer functions for lncRNAs based on chromatin state transitions during the differentiation of ESCs to cardiomyocytes (2,9). We believe this approach represents a relatively simple method to begin to prioritise cardiovascular lncRNAs for functional characterisation. The analysis is based on the observation that PCGs in differentiating cells cannot be assigned to a particular function based on their expression kinetics alone. However, Wamstad *et al.* (13) found using genomewide epigenomic profiling that PCGs co-expressed during cardiac differentiation can be functionally grouped based on distinct chromatin state transitions at their promoters. More specifically, they concluded that sub-groups of co-expressed PCGs, clustered based on unique chromatin state transitions, were involved in highly specific and distinct biological processes pertinent to cardiac development and function. These biological processes included cardiac signalling, metabolism, development and muscle contraction. We therefore surmised that novel lncRNAs sharing these unique chromatin state patterns at their promoters were likely to be involved in parallel biological processes. This approach therefore provides a very powerful and unbiased proxy to infer functions for novel lncRNAs. Supporting this, we found that heart-enriched novel lncRNAs were preferentially associated with clusters linked to fundamental cardiac biological processes including cardiac contraction and development. Finally, we also used this approach to predict targets of novel lncRNAs based on shared chromatin state patterns. For example, we demonstrated that Novlnc6 was able to regulate Bmp10, which shared the same chromatin patterns during differentiation. This resulted in a specific impact on the key core cardiac transcription factor, Nkx2-5, a known target of Bmp10 (2).

To conclude, our work, in addition to a number of other studies characterising the long noncoding transcriptome in disease (14,15), opens a new era of vast therapeutic and diagnostic potential. This highly integrated layer of noncoding RNAs exhibits interactions with components of various effector complexes to dictate the activity of developmental and pathological cardiac gene regulatory networks. Our data clearly demonstrates that pathological states within the heart are intrinsically linked to the dynamics of lncRNA expression. In the future, it will be of critical importance that investigators discovering novel cardiovascular lncRNAs leverage both computational and experimental approaches to functionally characterise the landscape of lncRNAs within the heart (9). In particular, one should focus on deciphering the mechanisms and language

through which lncRNAs interact with their respective protein, RNA or DNA partners. These approaches should illuminate our understanding of these very exciting therapeutic and diagnostic molecular targets.

### Acknowledgements

*Funding:* This work is in part funded by a grant from the Swiss National Science Foundation within the framework of the National Research Program 63 on ‘Stem cells and regenerative medicine’ (T Pedrazzini; Grant no 406340-128129).

### Footnote

*Conflicts of Interest:* The authors have filed a patent about the diagnostic and therapeutic use of several heart enriched lncRNAs.

### References

1. El Azzouzi H, Doevendans PA, Sluijter JP. Long non-coding RNAs in heart failure: an obvious lnc. *Ann Transl Med* 2016;4:182.
2. Ounzain S, Micheletti R, Beckmann T, et al. Genome-wide profiling of the cardiac transcriptome after myocardial infarction identifies novel heart-specific long non-coding RNAs. *Eur Heart J* 2015;36:353-68a.
3. Ounzain S, Crippa S, Pedrazzini T. Small and long non-coding RNAs in cardiac homeostasis and regeneration. *Biochim Biophys Acta* 2013;1833:923-33.
4. Batista PJ, Chang HY. Long noncoding RNAs: cellular address codes in development and disease. *Cell* 2013;152:1298-1307.
5. Lam MT, Li W, Rosenfeld MG, et al. Enhancer RNAs and regulated transcriptional programs. *Trends Biochem Sci* 2014;39:170-182.
6. Ounzain S, Pedrazzini T. The promise of enhancer-associated long noncoding RNAs in cardiac regeneration. *Trends Cardiovasc Med* 2015;25:592-602.
7. Ounzain S, Pedrazzini T. Super-enhancer lncs to cardiovascular development and disease. *Biochim Biophys Acta* 2016;1863:1953-60.
8. Ounzain S, Micheletti R, Arnan C, et al. CARMEN, a human super enhancer-associated long noncoding RNA controlling cardiac specification, differentiation and homeostasis. *J Mol Cell Cardiol* 2015;89:98-112.
9. Ounzain S, Burdet F, Ibberson M, et al. Discovery and functional characterization of cardiovascular long noncoding RNAs. *J Mol Cell Cardiol* 2015;89:17-26.
10. Ounzain S, Pezzuto I, Micheletti R, et al. Functional importance of cardiac enhancer-associated noncoding RNAs in heart development and disease. *J Mol Cell Cardiol* 2014;76:55-70.
11. Yang KC, Yamada KA, Patel AY, et al. Deep RNA sequencing reveals dynamic regulation of myocardial noncoding RNAs in failing human heart and remodeling with mechanical circulatory support. *Circulation* 2014;129:1009-1021.
12. Kumarswamy R, Bauters C, Volkmann I, et al. Circulating long noncoding RNA, LIPCAR, predicts survival in patients with heart failure. *Circ Res* 2014;114:1569-1575.
13. Wamstad JA, Alexander JM, Truty RM, et al. Dynamic and coordinated epigenetic regulation of developmental transitions in the cardiac lineage. *Cell* 2012;151:206-220.
14. Uchida S, Dimmeler S. Long noncoding RNAs in cardiovascular diseases. *Circ Res* 2015;116:737-750.
15. Devaux Y, Zangrando J, Schroen B, et al. Long noncoding RNAs in cardiac development and ageing. *Nat Rev Cardiol* 2015;12:415-425.

**Cite this article as:** Ounzain S, Pedrazzini T. Long non-coding RNAs in heart failure: a promising future with much to learn. *Ann Transl Med* 2016;4(15):298. doi: 10.21037/atm.2016.07.13

# Long non-coding RNAs in heart failure: an obvious Inc

Hamid El Azzouzi<sup>1,2</sup>, Pieter Adrianus Doevendans<sup>1,3</sup>, Joost Petrus Gerardus Sluijter<sup>1,2,3</sup>

<sup>1</sup>Department of Cardiology, Experimental Cardiology Laboratory, University Medical Center Utrecht, the Netherlands; <sup>2</sup>UMC Utrecht Regenerative Medicine Center, University Medical Center Utrecht, the Netherlands; <sup>3</sup>Netherlands Heart Institute (ICIN), Utrecht, the Netherlands  
*Correspondence to:* Dr. Joost Petrus Gerardus Sluijter. Department of Cardiology, UMC Utrecht, Heidelberglaan 100, Room G03.550, 3508 GA Utrecht, the Netherlands. Email: j.sluijter@umcutrecht.nl.

*Provenance:* This is a Guest Perspective commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

**Abstract:** Heart failure is a life-threatening and costly ailment characterized by structural and functional impairment of the heart. Despite major advances in understanding protein-mediated transcriptional control and signaling pathways that underlie the cellular and interstitial alterations of heart failure, significant therapeutical breakthroughs for innovative treatments of this disease are still missing. The recent extensive profiling of the mammalian transcriptome has revealed a large number of long non-coding RNAs (lncRNAs) that play a diversity of important regulatory roles in gene expression. In here, we focus on a recent work by Ounzain and colleagues comprising genome-wide profiling of the cardiac transcriptome after myocardial infarction with an emphasis on the identification of novel heart-specific lncRNAs.

**Keywords:** Long non-coding RNA (lncRNA); heart failure; gene networks

Submitted Mar 25, 2016. Accepted for publication Apr 05, 2016.

doi: 10.21037/atm.2016.05.06

**View this article at:** <http://dx.doi.org/10.21037/atm.2016.05.06>

Heart failure is a life-threatening and costly condition categorized by structural and functional impairment of ventricular filling or ejection of blood by the heart. Considering the overall increase in lifespan of the general population and high incidence of risk factors and improved survival from acute cardiovascular events, it is expected that the prevalence of heart failure will increase along with the associated emotional and socio-economic burdens. Upon injury, the heart responds to a diversity of biochemical and hemodynamic stressors through a process that entails molecular, cellular and interstitial alterations that are reflected further in size, shape and function of the heart (1). Although the initial response is adaptive and mainly serves to balance the decay in the pumping capacity of the heart, prolonged stress stimulation progressively contributes to deteriorate ventricular function which ultimately can lead to heart failure. Despite major advances in understanding the mechanisms that underlie cardiac remodeling and the worsened cardiac performance,

this did not translate into significant therapeutical breakthroughs for innovative treatments of heart failure in patients.

Once considered transcriptional noise, several transcriptome studies have shown prevalent transcription of a large number of non-coding RNAs (ncRNAs) (2). These ncRNAs are classified into two groups according to their length, whereby small ncRNAs are arbitrarily set at less than 200 nucleotides in size and include several ncRNAs species including small interfering RNAs (siRNAs), Piwi-interacting RNAs (piRNAs) and microRNAs (miRNAs) (3-5). In addition, transcripts of ncRNAs that are larger than 200 nucleotides are called long non-coding RNAs (lncRNAs). Of all currently studied noncoding genes, lncRNAs are a mystery species of growing scientific interest. Just like the extensively studied miRNAs, lncRNAs have been reported to have diverse biological functions such as regulation of chromatin remodeling, maintenance of the nuclear structure

integrity, and transcriptional and post-transcriptional processing (6,7). Despite their length, lncRNAs are distinct from protein coding genes as they are expressed at lower levels, are less evolutionarily conserved and mainly consist of a distinctive gene structure of usually 1–2 exons (8,9). In contrast to other ncRNAs, some lncRNAs have been shown to give rise to small peptides (10), showcasing the diversity of lncRNAs. The importance is underlined by the fact that specific lncRNA knockout strategies in mice showed severe effects that caused pre- and postnatal lethal phenotypes (11,12). Whereas deletion of lncRNAs *Peril* and *Mdgt* resulted in viability defects, mice lacking the lncRNA *Fendrr* died at the embryonic stage due to defects in several organs including the heart (11). These developmental defects were shown to be independent of adjacent genetic regions, e.g., *cis* or enhancer-like regulatory domains, as deletion of these lncRNAs did not phenocopy deletion of adjacent genes, which suggests a distinct function for these lncRNAs (12). One of the first reports on ncRNA expression showed that infants with non-syndromic tetralogy of Fallot had higher levels of an ALC-1 antisense lncRNA that disturbed ALC-1 translation in hypertrophied ventricles compared to their healthy counterparts (13,14). Next to the fact that some lncRNAs have been found to be altered in the developing or diseased heart, several single nucleotide polymorphism (SNP) in lncRNAs have shown to be strongly correlated with cardiovascular disease (15,16). For example, SNPs in myocardial infarction associated transcript (MIAT) and antisense non-coding RNA in the INK4 locus (ANRIL) forecast increased risk of cardiovascular disease (15,16). Moreover, lncRNA H19 was significantly upregulated in failing murine hearts, suggesting a role for hypoxia regulated lncRNA expression in heart failure (17). This is further highlighted by a recent study employing a comprehensive myocardial transcript profiling of coding and non-coding RNA species in failing human hearts and hearts that were unloaded with a left ventricular assist device (LVAD) (18). LVAD support has been known to diminish cardiac stress by reducing tissue hypoxia, auxiliary to the concept of restoration of cardiac normoxia by mechanical unloading (19,20). Fascinatingly, a significantly higher proportion of differentially expressed lncRNAs was normalized in hearts that were unloaded compared to mRNA and miRNA levels (18). In fact, lncRNA levels not only responded more sensitively to LVAD support

but their expression profile allowed to distinguish left ventricular samples from patients with ischemic and non-ischemic heart failure before and after LVAD support (18). It is no longer a question whether lncRNAs have a role in the cardiovascular system under normal physiological condition and in disease states. However, understanding the comprehensive role of lncRNA played herein is still in its infancy.

In an attempt to elucidate this, Ounzain and colleagues report an exciting genome-wide profiling of the cardiac transcriptome after myocardial infarction with an emphasis on the identification of heart-specific lncRNAs (21). In the article by Ounzain *et al.*, the authors performed RNA sequencing, ab initio transcript reconstruction and integrated genome-wide approaches to supplement gene regulatory networks with lncRNA profile in hearts after myocardial infarction. Of the multi-exonic transcripts, 14.3% were allocated as lncRNAs of which a smaller part (5.6%) corresponded to the UCSC annotation and a larger part (8.7%) was still not annotated. These unannotated transcripts showed several typical lncRNA physiognomies, such as genomic orientations relative to nearby coding genes and the low protein coding potential, and were therefore designated as novel lncRNAs. To verify whether these novel lncRNAs are cardiac specific and are associated with enhancer sequences active in the heart, qRT-PCR verification and association with flanking *cis*-regulatory proximal coding genes showed that differentially expressed novel lncRNAs were significantly more enriched in the myocardium than other transcript classes. Intriguingly, although gene ontology classification of the proximal genes revealed enrichment in processes associated with cardiac function and transcriptional regulation, novel lncRNAs were significantly associated with transcriptional control. Moreover, these lncRNAs were also located proximal to cardiac developmental genes, implicating a role for these novel lncRNAs in the stress responses and transcriptional reprogramming of the remodeling heart. Indeed, it was suggested that lncRNAs expression profiles in general are more profound to different heart failure etiologies when compared to other RNA transcripts (18). Mechanistically, Ounzain *et al.* demonstrated in the current study that the lncRNAs act as competitive endogenous RNAs specifically for microRNAs. Correlation network analysis of miRNA-lncRNA-mRNA indicated that lncRNAs correlated positively with miRNAs that have been known to be

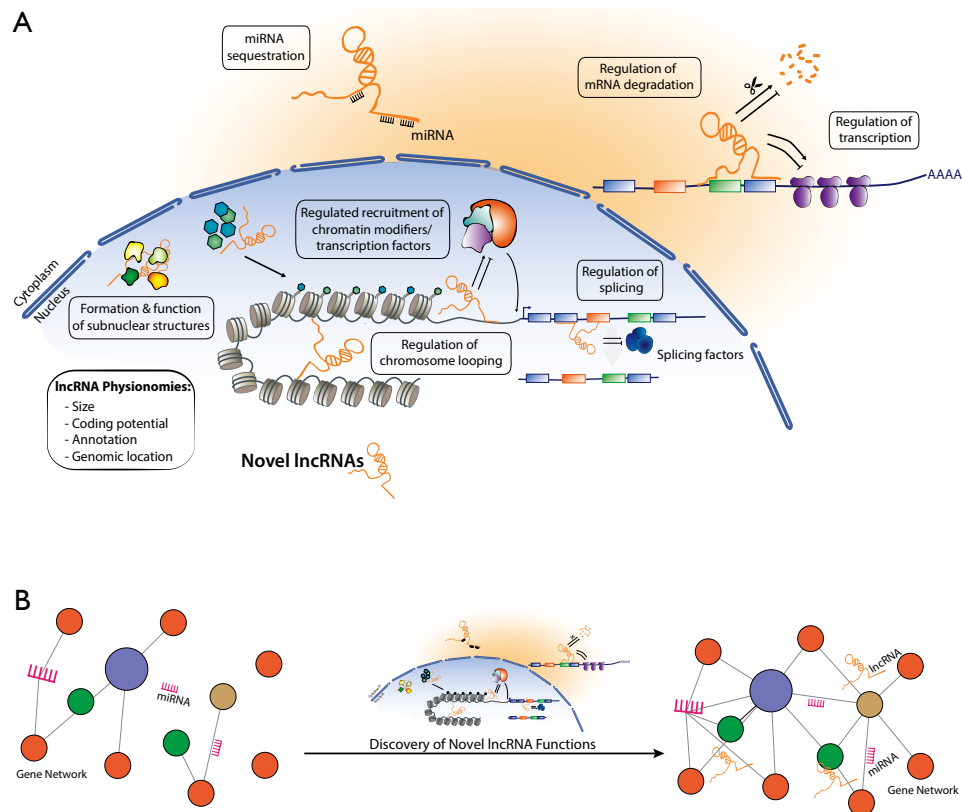
modulated in the post-infarcted myocardium, whereas mRNA levels correlated negatively. The likelihood that the predominant mechanism of action of cardiac lncRNAs is *cis*-, rather than *trans*-, gene regulation implicates a role as decoys for miRNAs. This is further exemplified by a study showing that linc-MD1, a muscle-specific lncRNA, absorbs miR-133 to regulate the expression of transcription factors that activate muscle-specific gene expression such as MAML1 and MEF2C (22). Moreover, linc-MD1 correlated strongly with aberrant muscle loss, as is seen in patients with Duchenne muscular dystrophy (23). In a cardiac setting, Wang *et al.* reported that cardiac apoptosis-related lncRNA (CARL) indirectly regulated mitochondrial fission and apoptosis, via an endogenous miR-539 sponge and thereby regulates PHB2 expression (24).

To assess whether the cardiac transcriptome, in particular the novel lncRNAs, correlated with the contractile and remodeling parameters of the heart *in vivo*, the cardiac transcriptome was associated with physiological traits in human patients. Though the novel reported lncRNAs correlated better with all physiological traits assessed, clustering for both coding transcripts and novel lncRNA indicated a group of related lncRNAs that correlated positively with cardiac function and negatively with remodeling parameters. The complexity to delineate how these particular physiological traits contribute to cardiac function and the function of lncRNAs herein is underscored by a single cluster showing diverse correlation to these interlinked functional parameters. Moreover, determining critical regulatory elements of transcriptional activity such as distinct chromatin states and methylation and acetylation of histone lysine residues is a major advance in understanding transcriptional regulation of lncRNAs. In line, although the novel lncRNAs were associated with more active chromatin states, their expressions showed to be more dependent on a canonical promoter signature rather than an exclusively active enhancer signature.

Although the lncRNAs complied with the UCSC annotation and have typical lncRNA physiognomies, validation of these transcripts is necessary to filter out false reads. The authors used qRT-PCR to validate a select group that was associated with functional parameters and to determine timely and regional cardiac expression. In contrast to mRNAs, lncRNA themselves must physically localize to their particular site of action, making knowledge

of lncRNA subcellular localization patterns vital for insights into their biology and function. Nuclear and cytoplasmic fractionation could indicate subcellular enrichment of an lncRNA and give insights into the functionality of the lncRNA. Indeed, an exclusive nuclear localization would argue against putative short peptide sequences encoded by the lncRNA, since translation occurs in the cytoplasm. However, to fully delineate the exact role of an lncRNA, further analysis on localization to particular subcellular areas within the cytosol or nucleus may lead to better insights on the role of an lncRNA. For example, lncRNAs such as NEAT1 and MALAT1 have been shown to localize to nuclear bodies within the nucleus and the lncRNA GAS5 was shown to shuttle between the nucleus and cytoplasm (25,26). More elegantly, Clemson and colleagues showed that the lncRNA XIST specifically accumulates on the inactive X-chromosome (27). Hence, RNA fluorescence *in situ* hybridization (RNA FISH) could better address these questions and reveal potential mechanisms for the described novel lncRNAs. Moreover, this technique would directly discriminate the cell type presence *in situ* of these novel lncRNA in the injured myocardium instead of the laborious and indirect detection in isolated different cell types

For a long time heart function was considered to be orchestrated through regulatory networks that are composed solely of a protein-mediated transcriptional control and signaling pathways, and which were perturbed during cardiac disease. In line, efforts for therapeutical interventions were systematized around these networks with little translational outcomes so far. Clearly the path of lncRNAs from background noise and serendipitous transcription to key regulators has led to significant insights into gene regulatory networks (*Figure 1*). This previously hidden layer of gene regulation underpins the sophisticated regulatory networks to not only coordinate physiological responses and environmental adaptation, but also pathophysiological responses in multifaceted heart diseases. In order to take the next step towards modulation of lncRNA levels as an approach for tackling cardiac disease, reviewing their function and role in cardiac biology is of utmost importance. Therefore, efforts to unveil the multiple lncRNAs and their role in cardiac disease, exemplified by the study of Ounzain and colleagues, provides an excellent toolkit to assist in exploring the possibilities for ranging from biomarkers of disease to targets for therapeutical intervention.



**Figure 1** Long non-coding RNAs (lncRNAs) and their functions. (A) Possible functional mechanisms for the novel lncRNAs; (B) lncRNAs as an additional layer of controllers of gene regulatory networks.

## Acknowledgements

**Funding:** This work was supported by the PLN foundation and a HUSTCARE grant (CVON2011-12) from the Netherlands CardioVascular Research Initiative (CVON): the Dutch Heart Foundation, Dutch Federation of University Medical Centers, the Netherlands Organization for Health Research and Development, and the Royal Netherlands Academy of Sciences.

## Footnote

**Conflicts of Interest:** The authors have no conflicts of interest to declare.

## References

1. Kehat I, Molkenkin JD. Molecular pathways underlying cardiac remodeling during pathophysiological stimulation. *Circulation* 2010;122:2727-2735.
2. Scheuermann JC, Boyer LA. Getting to the heart of the matter: long non-coding RNAs in cardiac development and disease. *EMBO J* 2013;32:1805-1816.
3. Castel SE, Martienssen RA. RNA interference in the nucleus: roles for small RNAs in transcription, epigenetics and beyond. *Nat Rev Genet* 2013;14:100-112.
4. Malone CD, Hannon GJ. Small RNAs as guardians of the genome. *Cell* 2009;136:656-668.
5. Sluijter JP. MicroRNAs in Cardiovascular Regenerative Medicine: Directing Tissue Repair and Cellular Differentiation. Available online: <http://dx.doi.org/>
6. Fatica A, Bozzoni I. Long non-coding RNAs: new players in cell differentiation and development. *Nat Rev Genet* 2014;15:7-21.
7. Zhu YG, Feng XM, Abbott J, et al. Human mesenchymal stem cell microvesicles for treatment of Escherichia coli endotoxin-induced acute lung injury in mice. *Stem Cells* 2014;32:116-125.
8. Dinger ME, Pang KC, Mercer TR, et al. Differentiating protein-coding and noncoding RNA: challenges and ambiguities. *PLoS Comput Biol* 2008;4:e1000176.

9. Hangauer MJ, Vaughn IW, McManus MT. Pervasive transcription of the human genome produces thousands of previously unidentified long intergenic noncoding RNAs. *PLoS Genet* 2013;9:e1003569.
10. Cooper C, Vincett D, Yan Y, et al. Steroid Receptor RNA Activator bi-faceted genetic system: Heads or Tails? *Biochimie* 2011;93:1973-1980.
11. Sauvageau M, Goff LA, Lodato S, et al. Multiple knockout mouse models reveal lincRNAs are required for life and brain development. *Elife* 2013;2:e01749.
12. Trajkovski M, Hausser J, Soutschek J, et al. MicroRNAs 103 and 107 regulate insulin sensitivity. *Nature* 2011;474:649-653.
13. O'Brien JE Jr, Kibiryeveva N, Zhou XG, et al. Noncoding RNA expression in myocardium from infants with tetralogy of Fallot. *Circ Cardiovasc Genet* 2012;5:279-286.
14. Ritter O, Haase H, Schulte HD, et al. Remodeling of the hypertrophied human myocardium by cardiac bHLH transcription factors. *J Cell Biochem* 1999;74:551-561.
15. Carlock LR, Skarecky D, Dana SL, et al. Deletion mapping of human chromosome 5 using chromosome-specific DNA probes. *Am J Hum Genet* 1985;37:839-852.
16. Ishii N, Ozaki K, Sato H, et al. Identification of a novel non-coding RNA, MIAT, that confers risk of myocardial infarction. *J Hum Genet* 2006;51:1087-1099.
17. Lee JH, Gao C, Peng G, et al. Analysis of transcriptome complexity through RNA sequencing in normal and failing murine hearts. *Circ Res* 2011;109:1332-1341.
18. Yang KC, Yamada KA, Patel AY, et al. Deep RNA sequencing reveals dynamic regulation of myocardial noncoding RNAs in failing human heart and remodeling with mechanical circulatory support. *Circulation* 2014;129:1009-1021.
19. Ambardekar AV, Buttrick PM. Reverse remodeling with left ventricular assist devices: a review of clinical, cellular, and molecular effects. *Circ Heart Fail* 2011;4:224-233.
20. Alcalay J, Ullrich SE, Kripke ML. Local suppression of contact hypersensitivity in mice by a monofunctional psoralen plus UVA radiation. *Photochem Photobiol* 1989;50:217-220.
21. Ounzain S, Micheletti R, Beckmann T, et al. Genome-wide profiling of the cardiac transcriptome after myocardial infarction identifies novel heart-specific long non-coding RNAs. *Eur Heart J* 2015;36:353-68a.
22. Cesana M, Cacchiarelli D, Legnini I, et al. A long noncoding RNA controls muscle differentiation by functioning as a competing endogenous RNA. *Cell* 2011;147:358-369.
23. Legnini I, Morlando M, Mangiavacchi A, et al. A feedforward regulatory loop between HuR and the long noncoding RNA linc-MD1 controls early phases of myogenesis. *Mol Cell* 2014;53:506-514.
24. Wang K, Long B, Zhou LY, et al. CARL lincRNA inhibits anoxia-induced mitochondrial fission and apoptosis in cardiomyocytes by impairing miR-539-dependent PHB2 downregulation. *Nat Commun* 2014;5:3596.
25. Ip JY, Nakagawa S. Long non-coding RNAs in nuclear bodies. *Dev Growth Differ* 2012;54:44-54.
26. Puig JG, Michán AD, Jiménez ML, et al. Female gout. Clinical spectrum and uric acid metabolism. *Arch Intern Med* 1991;151:726-732.
27. Clemson CM, McNeil JA, Willard HF, et al. XIST RNA paints the inactive X chromosome at interphase: evidence for a novel RNA involved in nuclear/chromosome structure. *J Cell Biol* 1996;132:259-275.

**Cite this article as:** EL Azzouzi H, Doevendans PA, Sluijter JP. Long non-coding RNAs in heart failure: an obvious Inc. *Ann Transl Med* 2016;4(9):182. doi: 10.21037/atm.2016.05.06



# Association between microRNAs and coronary collateral circulation: is there a new role for the small non-coding RNAs?

Nikolaos Papageorgiou<sup>1</sup>, Effimia Zacharia<sup>2</sup>, Dimitris Tousoulis<sup>2</sup>

<sup>1</sup>Barts Heart Centre, St Bartholomew's Hospital, London, UK; <sup>2</sup>Department of Cardiology, Hippokraton Hospital, University of Athens, Athens, Greece

*Correspondence to:* Dr. Nikolaos Papageorgiou, MD, PhD. Barts Heart Centre, St Bartholomew's Hospital, West Smithfield, London, EC1A 7BE, UK. Email: drnpapageorgiou@yahoo.com.

*Provenance:* This is a Guest Editorial commissioned by Section Editor Zhijun Han, MD (Department of Laboratory Medicine, Wuxi Second Hospital, Nanjing Medical University, Wuxi, China).

*Comment on:* Hakimzadeh N, Nossent AY, van der Laan AM, *et al.* Circulating MicroRNAs Characterizing Patients with Insufficient Coronary Collateral Artery Function. PLoS One 2015;10:e0137035.

**Abstract:** We read with interest the article entitled “Circulating microRNAs characterizing patients with insufficient coronary collateral artery function” which was recently published in the PLOS ONE journal. It was demonstrated for the first time that specific circulating microRNAs (miRNAs) can distinguish patients with sufficient from those with insufficient coronary collateral circulation. Circulating miRNAs in the plasma of patients with stable CAD and chronic CTO could provide information with regard to the coronary collateral artery capacity. However, several aspects need to be taken into consideration before the use of miRNAs in the clinical practice. A risk model that would incorporate risk factors for cardiovascular disease and miRNAs could prove to be very useful. Although an association between the levels of miRNAs and the collateral artery capacity appears promising, it still does not confirm any causal role for miRNAs. Therefore, large clinical studies in populations with CTO are warranted to evaluate this finding.

**Keywords:** microRNAs (miRNAs); coronary artery collaterals; angiogenesis

Submitted Apr 26, 2016. Accepted for publication May 06, 2016.

doi: 10.21037/atm.2016.05.51

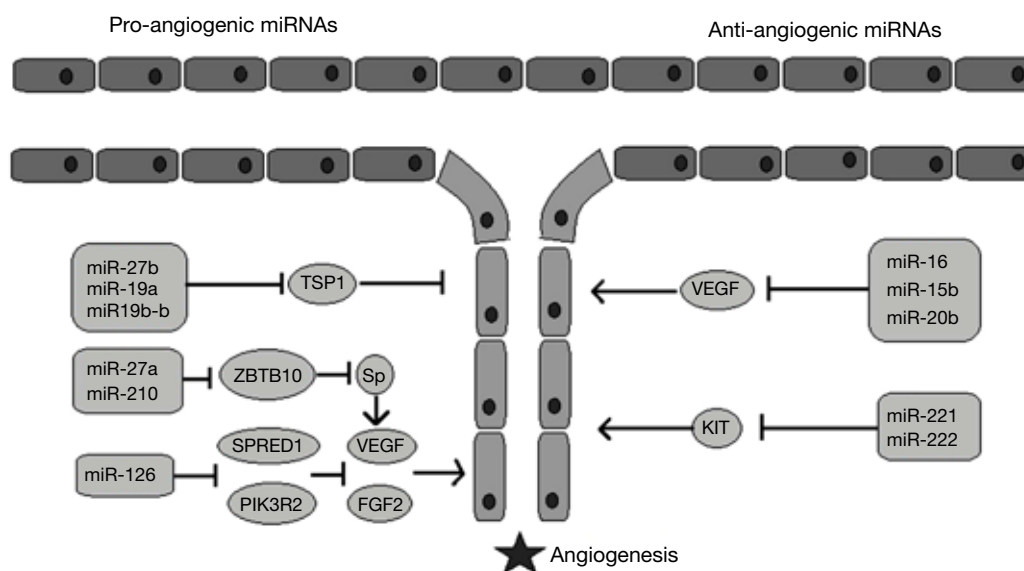
**View this article at:** <http://dx.doi.org/10.21037/atm.2016.05.51>

Studies have shown that the presence of sufficient collateral circulation is beneficial for patients with stable CAD, as it improves the survival rates (1). A circulating biomarker that would provide us with the appropriate information regarding to the existence of collateral circulation in these patients would be very helpful in the clinical setting. microRNAs (miRNAs) actively participate in cardiovascular homeostasis and play an important role in the initiation and progression of cardiovascular disease (2). Recent data suggest that miRNAs contribute to the formation of vulnerable atherosclerotic plaques (3), while they enhance angiogenesis as well. Therefore, it has been speculated that circulating miRNAs can provide information about the collateral artery network of patients with chronic total occlusion (CTO). In the present article, Hakimzadeh *et al.* (4)

showed for the first time that specific circulating miRNAs can distinguish patients with sufficient and patients with insufficient coronary collateral circulation.

## miRNAs and angiogenesis

It was an interesting discovery that the enzymes involved in miRNAs maturation also participate in angiogenesis: Dicer inhibition resulted in a decrease in angiogenesis *in vivo*, while inhibition of Drosha resulted in an anti-angiogenic effect *in vitro* (5). Since then, studies have shown that several miRNAs upregulate angiogenesis, while others suppress angiogenic pathways (6) (*Figure 1*). However, the data were mainly derived from preclinical studies, until Nie *et al.* (7) showed that miR-126 levels, along with vascular endothelial



**Figure 1** Pro-angiogenic versus anti-angiogenic miRNAs. Some miRNAs upregulate angiogenesis by promoting angiogenic factors (i.e., VEGF) or by inhibiting anti-angiogenic mediators (i.e., TSP1), while others downregulate angiogenesis by inhibiting angiogenic molecular pathways. VEGF, vascular endothelial growth factor; TSP1, thrombospondin 1; ZBTB10, zinc finger and BTB domain-containing protein 10; SPRED1, sprout-related EVH1 domain-containing protein 1; PIK3R2, phosphatidylinositol 3-kinase regulatory subunit beta; Sp, substance P; FGF2, fibroblast growth factor; KIT, receptor kinase protein/gene.

growth factor (VEGF) levels, were higher in healthy people and in patients with well-developed collateral arteries compared to patients with under-developed collateral circulation. In addition, miR-126 levels could independently predict coronary collateral circulation formation. Nevertheless, Hakimzadeh *et al.* (4) provided further insight to this issue. Their study included patients undergoing successful percutaneous coronary intervention and had CTO of a coronary artery. The levels of miR-423-5p, miR-30d, miR-10b and miR-126 were increased in the setting of insufficient coronary collateral artery capacity. In comparison to healthy controls, though, only the levels of miR-30d and miR-126 were found to be elevated. Indeed, there is evidence that gene modulation can discriminate between patients with well developed and patients with poorly developed collateral arteries. van der Laan (8) showed in patients with CTO that the mRNA expression of galectin-2 was increased in monocytes of patients with low collateral flow index (CFI). The rs7291467 polymorphism was associated with increased galectin-2 levels and a lower angiogenic response. Nevertheless, the role of miRNAs in angiogenesis was only recently examined.

Several studies have proved the angiogenic potential of miR-126. A study that included patients with right ventricular heart failure and pulmonary hypertension

showed that lower levels of miR-126 were expressed in right ventricular tissues of patients with decompensated heart failure. Of note, this was associated with decreased capillary density. The *in vivo* upregulation of miR-126 improved vascular density in an experimental animal model of pulmonary artery hypertension (9). More recently, the administration of miR-126 through ultrasound-targeted microbubble destruction resulted in increased vascular density in an animal model of chronic ischemia, as it enhanced VEGF and promoted angiopoietin-1 signaling (10).

In turn miRNA-10b has been shown to regulate angiogenesis in glioblastoma multiforme (11) and it was found to be implicated in vascular smooth muscle cell proliferation which is associated with atherosclerosis progression (12). miR-423-5p has been recently recognized as a novel biomarker for congestive heart failure and correlates with pro-brain natriuretic peptide (pro-BNP) levels. As opposed to miR-30d, though, miR-423-5p has been linked to increased cardiomyocyte apoptosis (13).

### miRNAs as circulating biomarkers for coronary collateral circulation: challenges to meet

A circulating biomarker that would determine patients with poorly developed collateral coronary artery network would be

**Table 1** Selected studies investigating predictors for poor coronary collateral capacity

Studies	Year	Population	N	Collateral flow stratification	Result
Hakimzadeh <i>et al.</i> (4)	2015	CTO patients undergoing coronary angiography	41	Poor CCC: CFI <0.39 Good CCC: CFI >0.39	(+) miRNA-126, miRNA-10b, miRNA-30d and miRNA-423-5p in patients with poor CCC
Nie <i>et al.</i> (7)	2014	CAD patients with ≥95% stenosis in a coronary artery	120	Poor CCC: grade 0 and grade 1 Rentrop Good CCC: grade 1 and grade 2 Rentrop	miR-126 and VEGF levels independently predicted CCC development
İleri <i>et al.</i> (15)	2016	Patients with NSTEMI	224	Poor CCC: grade 0 and grade 1 Rentrop Good CCC: grade 1 and grade 2 Rentrop	DM, WBC, neutrophil counts and NLR independently predicted low CCC; age negatively predicted poor CCC
Kalkan <i>et al.</i> (16)	2014	Patients with CTO	274	Poor CCC: grade 0 and grade 1 Rentrop Good CCC: grade 1 and grade 2 Rentrop	NLR, hs-CRP, WBC independently predicted poor CCC;
Baykan <i>et al.</i> (17)	2015	Patients with CTO	163	Poor CCC: grade 0 and grade 1 Rentrop Good CCC: grade 1 and grade 2 Rentrop	(+) Aix, PWV, fasting glucose, creatine, uric acid, neutrophil count and NLR in patients with low CCC
Yetkin <i>et al.</i> (18)	2015	Patients with at least one coronary stenosis of ≥95% that underwent coronary angiography	502 (228 with CTO)	Poor CCC: grade 0 and grade 1 Rentrop Good CCC: grade 1 and grade 2 Rentrop	DM and female gender predicted poor CCC; monocyte count was independent of CCC
van der Hoeven <i>et al.</i> (19)	2013	Patients with CTO	295	Poor CCC: CFI <0.39 Good CCC: CFI >0.39	Beta blockers, hypertension and angina pectoris were positively associated with CFI; WBC, prior MI and high DBP were negatively associated with CFI
van der Laan <i>et al.</i> (8)	2012	Patients with CTO	50	Dichotomized according to CFI	(+) mRNA expression of galectin-2 in monocytes of patients with poor CCC; (+) polymorphism rs7291467 CC genotype in patients with poor CCC

(+), increased. CCC, coronary collateral circulation; CTO, chronic total occlusion; CFI, collateral flow index; CAD, coronary artery disease; VEGF, vascular endothelial growth factor; NSTEMI, non ST-elevation myocardial infarction; WBC, white blood cell; NLR, neutrophil to monocyte ratio; hs-CRP, high sensitivity CRP; Aix, augmentation index; PWV, pulse wave velocity; MI, myocardial infarction; DBP, diastolic blood pressure.

of great clinical significance, since the invasive procedure of coronary angiography could potentially be avoided. In addition, coronary angiography can disclose arteries of >100 μm (14), thus cannot provide insight into microcirculation. It appears that the findings of Hakimzadeh *et al.* (4) are promising. However, up to date, no equivalent circulating biomarker exists (*Table 1*). That makes it difficult to compare the discriminatory capacity of miR 423-5p, miR-30d, miR-10b and miR-126 with the discriminatory capacity of an established circulating biomarker. It should be mentioned

that a generally accepted definition of low collateral capacity is necessary, so that the results of the future studies will be comparable. For example, Nie *et al.* (7) used Rentrop grades to assess coronary collateral circulation, while other studies (1) considered collateral circulation capacity to be insufficient when CFI was <0.25. Hakimzadeh *et al.* (4) defined insufficient collateral circulation as the one with CFI<0.39 and this was in agreement with the study of van der Hoeven *et al.* (19). It should be stressed out that the intracoronary assessment of CFI in healthy individuals

is not feasible, thereby limiting the information about the collateral network in this population. Despite this limitation, the levels of miR-30d and miR-126 were found to be lower in healthy individuals compared to patients with CTO (4).

Several co-existing parameters might affect the diagnostic ability of miRNAs in the general population. Diabetes mellitus interferes with the formation of collateral coronary artery network (15), while it has been found to down regulate the expression of several miRNAs, including miR-126 (20). A recent risk scoring model that predicts poor collateral coronary circulation showed that a combination of white blood cell count, age and history of myocardial infarction can predict poorly developed collaterals (15). Therefore it is necessary to stratify patients' risk for poor collateral coronary circulation before attempting to evaluate the levels of a circulating biomarker. In the present article (4), patients with diabetes mellitus and a history of myocardial infarction were excluded from the study so that the expression patterns of miRNAs could be examined independently. Of note, the discriminatory power of miR-10b, miR-30d, miR-423-5p was evident only after multivariate analysis, which underscores the significance of taking all clinical parameters into consideration. Since leukocytes were found to be implicated in collateral artery growth (15), their assessment seems mandatory. Hakimzadeh *et al.* (4) found an association between miRNA-10b levels and monocyte/leukocyte count, suggesting a possible link between these parameters. Finally, aspirin administration has been found to decrease miR-126 levels, since platelets are a major pool of circulating miR-126 (21). Nevertheless, in the present article, aspirin administration did not blunt the discriminatory efficacy of miR-126 (4).

## Conclusions

Circulating miRNAs in the plasma of patients with stable CAD and CTO have been shown to provide information about the coronary collateral artery capacity. This could possibly suggest an alternative diagnostic route to the invasive coronary angiography. However, several aspects need to be taken into consideration before the use of specific miRNAs could be applied into clinical practice. Co-existing parameters, such as diabetes mellitus and leukocyte count, affect angiogenesis and might interfere with the diagnostic efficacy of miRNAs; therefore, a risk model that would incorporate such parameters could be useful. It has become evident that an association between

the levels of circulating miRNAs and the collateral artery capacity is not enough to confirm an underlying causative mechanism. Therefore, many more large clinical studies in populations with CTO are warranted to evaluate this finding.

## Acknowledgements

None.

## Footnote

*Conflicts of Interest:* The authors have no conflicts of interest to declare.

## References

1. Meier P, Gloekler S, Zbinden R, et al. Beneficial effect of recruitable collaterals: a 10-year follow-up study in patients with stable coronary artery disease undergoing quantitative collateral measurements. *Circulation* 2007;116:975-983.
2. Papageorgiou N, Tousoulis D, Androulakis E, et al. The role of microRNAs in the initiation and progression of stable atheromatous plaque. *Curr Pharm Des* 2013;19:1651-1657.
3. Haver VG, Slart RH, Zeebregts CJ, et al. Rupture of vulnerable atherosclerotic plaques: microRNAs conducting the orchestra? *Trends Cardiovasc Med* 2010;20:65-71.
4. Hakimzadeh N, Nossent AY, van der Laan AM, et al. Circulating MicroRNAs Characterizing Patients with Insufficient Coronary Collateral Artery Function. *PLoS One* 2015;10:e0137035.
5. Kuehbach A, Urbich C, Zeiher AM, et al. Role of Dicer and Drosha for endothelial microRNA expression and angiogenesis. *Circ Res* 2007;101:59-68.
6. Chen Y, Banda M, Speyer CL, et al. Regulation of the expression and activity of the antiangiogenic homeobox gene GAX/MEOX2 by ZEB2 and microRNA-221. *Mol Cell Biol* 2010;30:3902-3913.
7. Nie X, Su L, Zhou Y, et al. Association between plasma levels of microRNA-126 and coronary collaterals in patients with coronary artery disease. *Zhonghua Xin Xue Guan Bing Za Zhi* 2014;42:561-565.
8. van der Laan AM, Schirmer SH, de Vries MR, et al. Galectin-2 expression is dependent on the rs7291467 polymorphism and acts as an inhibitor of arteriogenesis. *Eur Heart J* 2012;33:1076-1084.
9. Potus F, Ruffenach G, Dahou A, et al. Downregulation

- of MicroRNA-126 Contributes to the Failing Right Ventricle in Pulmonary Arterial Hypertension. *Circulation* 2015;132:932-943.
10. Cao WJ, Rosenblat JD, Roth NC, et al. Therapeutic Angiogenesis by Ultrasound-Mediated MicroRNA-126-3p Delivery. *Arterioscler Thromb Vasc Biol* 2015;35:2401-2411.
  11. Lin J, Teo S, Lam DH, et al. MicroRNA-10b pleiotropically regulates invasion, angiogenicity and apoptosis of tumor cells resembling mesenchymal subtype of glioblastoma multiforme. *Cell Death Dis* 2012;3:e398.
  12. Yu X, Li Z, Chen G, et al. MicroRNA-10b Induces Vascular Muscle Cell Proliferation Through Akt Pathway by Targeting TIP30. *Curr Vasc Pharmacol* 2015;13:679-686.
  13. Luo P, He T, Jiang R, et al. MicroRNA-423-5p targets O-GlcNAc transferase to induce apoptosis in cardiomyocytes. *Mol Med Rep* 2015;12:1163-1168.
  14. Kolibash AJ, Bush CA, Wepsic RA, et al. Coronary collateral vessels: spectrum of physiologic capabilities with respect to providing rest and stress myocardial perfusion, maintenance of left ventricular function and protection against infarction. *Am J Cardiol* 1982;50:230-238.
  15. İleri M, Güray Ü, Yetkin E, et al. A new risk scoring model for prediction of poor coronary collateral circulation in acute non-ST-elevation myocardial infarction. *Cardiol J* 2016;23:107-113.
  16. Kalkan M, Sahin M, Kalkan A, et al. The relationship between the neutrophil-lymphocyte ratio and the coronary collateral circulation in patients with chronic total occlusion. *Perfusion* 2014;29:360-366.
  17. Baykan AO, Gür M, Acele A, et al. Coronary collateral development and arterial stiffness in patients with chronic coronary total occlusions. *Scand Cardiovasc J* 2015;49:228-234.
  18. Yetkin E, Topal E, Erguzel N, et al. Diabetes mellitus and female gender are the strongest predictors of poor collateral vessel development in patients with severe coronary artery stenosis. *Angiogenesis* 2015;18:201-207.
  19. van der Hoeven NW, Teunissen PF, Werner GS, et al. Clinical parameters associated with collateral development in patients with chronic total coronary occlusion. *Heart* 2013;99:1100-1105.
  20. Jansen F, Yang X, Hoelscher M, et al. Endothelial microparticle-mediated transfer of MicroRNA-126 promotes vascular endothelial cell repair via SPRED1 and is abrogated in glucose-damaged endothelial microparticles. *Circulation* 2013;128:2026-2038.
  21. de Boer HC, van Solingen C, Prins J, et al. Aspirin treatment hampers the use of plasma microRNA-126 as a biomarker for the progression of vascular disease. *Eur Heart J* 2013;34:3451-3457.

**Cite this article as:** Papageorgiou N, Zacharia E, Tousoulis D. Association between microRNAs and coronary collateral circulation: is there a new role for the small non-coding RNAs? *Ann Transl Med* 2016;4(11):223. doi: 10.21037/atm.2016.05.51



# AME JOURNALS

创立于2009年7月的AME Publishing Company (简称AME, 代表Academic Made Easy, Excellent and Enthusiastic), 是一家崇尚创新、具有国际化视野和互联网思维的医学出版公司。AME拥有专业的期刊运营团队, 提供以国际组稿为核心竞争力的全流程出版服务, 专注于国际医学期刊、书籍的出版和医疗科研资讯成果的推广, 已在香港、台北、悉尼、广州、长沙、上海、北京、杭州、南京和成都等地设立办公室。目前出版了62本涵盖肿瘤、心血管、胸部疾病、影像和外科等不同领域的学术期刊, 已有18本被PubMed收录, 9本被SCI收录, 出版中英文医学专业图书近百本。



期刊名称: JTD  
 创刊时间: 2009年12月  
 PubMed收录: 2011年12月  
 SCI收录: 2013年2月



期刊名称: QIMS  
 创刊时间: 2011年12月  
 PubMed收录: 2012年9月  
 SCI收录: 2017年12月



期刊名称: TCR  
 创刊时间: 2012年6月  
 SCI收录: 2015年10月



期刊名称: ATM  
 创刊时间: 2013年4月  
 PubMed收录: 2014年9月  
 SCI收录: 2018年3月



期刊名称: HBSN  
 创刊时间: 2012年12月  
 PubMed收录: 2014年9月  
 SCI收录: 2017年6月



期刊名称: ACS  
 创刊时间: 2012年  
 PubMed收录: 2014年  
 SCI收录: 2018年5月



期刊名称: TLCR  
 创刊时间: 2012年3月  
 PubMed收录: 2014年12月  
 SCI收录: 2018年10月



期刊名称: TAU  
 创刊时间: 2012年3月  
 PubMed收录: 2015年11月  
 SCI收录: 2018年12月



期刊名称: GS  
 创刊时间: 2012年5月  
 PubMed收录: 2014年6月  
 SCI收录: 2018年12月

AME  
Books  
AME图书



## AME 图书 2.0 正式上线

随手，随时，随地关注医学健康与人文

### 精品医学书籍

囊括AME全系列图书及学术期刊

- 最前沿医学知识
- 最实用科研干货
- 最独到学术见解

### 多种分类书目

- 按专家分类
- 按专科分类
- 按系列分类

随心所欲，找书不再烦恼！

### 支持快币兑换

攒了快币没地花？  
从此买书不花钱！

### 目录一键跳转

不再一页一页翻资料，目录  
一目了然，一键快捷跳转！